



INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁵ : C07H 15/12, 17/00, A61K 37/00 C07K 13/00, 15/00, C12N 5/00	A1	(11) International Publication Number: WO 93/10136 (43) International Publication Date: 27 May 1993 (27.05.93)
(21) International Application Number: PCT/US92/09893 (22) International Filing Date: 16 November 1992 (16.11.92) (30) Priority data: 07/793,065 15 November 1991 (15.11.91) US (71) Applicant: THE TRUSTEES OF PRINCETON UNIVERSITY [US/US]; New South Building, 5th Floor, Princeton, NJ 08544 (US). (72) Inventor: LEMISCHKA, Ihor, R. ; 5T Hibben Apartments, Faculty Road, Princeton, NJ 08540 (US). (74) Agent: FEIT, Irving, N.; ImClone Systems Incorporated, 180 Varick Street, New York, NY 10014 (US).		(81) Designated States: AU, CA, FI, HU, JP, KP, NO, RO, RU, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, SE). Published <i>With international search report.</i>
(54) Title: TOTIPOTENT HEMATOPOIETIC STEM CELL RECEPTORS AND THEIR LIGANDS		
(57) Abstract Isolated mammalian nucleic acid molecules encoding receptor protein tyrosine kinases expressed in primitive hematopoietic cells and not expressed in mature hematopoietic cells are provided. Also included are the receptors encoded by such nucleic acid molecules; the nucleic acid molecules encoding receptor protein tyrosine kinases having the sequences shown in Figure 1a (murine flk-2), Figure 1b (human flk-2) and Figure 2 (murine flk-1); the receptor protein tyrosine kinases having the amino acid sequences shown in Figure 1a, Figure 1b and Figure 2; ligands for the receptors; nucleic acid sequences that encode the ligands; and methods of stimulating the proliferation and/or differentiation of primitive mammalian hematopoietic stem cells comprising contacting the stem with a ligand that binds to a receptor protein tyrosine kinase expressed in primitive mammalian hematopoietic cells and not expressed in mature hematopoietic cells.		

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**TOTIPOTENT HEMATOPOIETIC STEM CELL
RECEPTORS AND THEIR LIGANDS**

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The invention described in this application was made with U.S. government support from Grant Numbers R01-CA45339 and R01-DK42989 awarded by the National Institutes of Health. The government has certain rights in this invention.

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FIELD OF THE INVENTION

The present invention relates to hematopoietic stem cell receptors, ligands for such receptors, and nucleic acid molecules encoding such receptors and ligands.

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BACKGROUND OF THE INVENTION

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The mammalian hematopoietic system comprises red and white blood cells. These cells are the mature cells that result from more primitive lineage-restricted cells. The cells of the hematopoietic system have been reviewed by Dexter and Spooner in the Annual Review of Cell Biology 3, 423-441 (1987).

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The red blood cells, or erythrocytes, result from primitive cells referred to by Dexter and Spooner as erythroid burst-forming units (BFU-E). The immediate progeny of the erythroid burst-forming units are called erythroid colony-forming units (CFU-E).

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The white blood cells contain the mature cells of the lymphoid and myeloid systems. The lymphoid cells include B lymphocytes and T lymphocytes. The B and T lymphocytes result from earlier progenitor cells referred to by Dexter and Spooner as preT and preB cells.

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The myeloid system comprises a number of cells including granulocytes, platelets, monocytes, macrophages, and megakaryocytes. The granulocytes are further divided into

neutrophils, eosinophils, basophils and mast cells.

Each of the mature hematopoietic cells are specialized for specific functions. For example, erythrocytes are responsible for oxygen and carbon dioxide transport. T and B lymphocytes are responsible for cell-and antibody-mediated immune responses, respectively. Platelets are involved in blood clotting. Granulocytes and macrophages act generally as scavengers and accessory cells in the immune response against invading organisms and their by-products.

At the center of the hematopoietic system lie one or more totipotent hematopoietic stem cells, which undergo a series of differentiation steps leading to increasingly lineage-restricted progenitor cells. The more mature progenitor cells are restricted to producing one or two lineages. Some examples of lineage-restricted progenitor cells mentioned by Dexter and Spooncer include granulocyte/macrophage colony-forming cells (GM-CFC), megakaryocyte colony-forming cells (Meg-CFC), eosinophil colony-forming cells (Eos-CFC), and basophil colony-forming cells (Bas-CFC). Other examples of progenitor cells are discussed above.

The hematopoietic system functions by means of a precisely controlled production of the various mature lineages. The totipotent stem cell possesses the ability both to self renew and to differentiate into committed progenitors for all hematopoietic lineages. These most primitive of hematopoietic cells are both necessary and sufficient for the complete and permanent hematopoietic reconstitution of a radiation-ablated hematopoietic system in mammals. The ability of stem cells to reconstitute the entire hematopoietic system is the basis of bone marrow transplant therapy.

It is known that growth factors play an important role in the development and operation of the mammalian hematopoietic system. The role of growth factors is complex,

however, and not well understood at the present time. One reason for the uncertainty is that much of what is known about hematopoietic growth factors results from in vitro experiments. Such experiments do not necessarily reflect in vivo realities.

In addition, in vitro hematopoiesis can be established in the absence of added growth factors, provided that marrow stromal cells are added to the medium. The relationship between stromal cells and hematopoietic growth factors in vivo is not understood. Nevertheless, hematopoietic growth factors have been shown to be highly active in vivo.

From what is known about them, hematopoietic growth factors appear to exhibit a spectrum of activities. At one end of the spectrum are growth factors such as erythropoietin, which is believed to promote proliferation only of mature erythroid progenitor cells. In the middle of the spectrum are growth factors such as IL-3, which is believed to facilitate the growth and development of early stem cells as well as of numerous progenitor cells. Some examples of progenitor cells induced by IL-3 include those restricted to the granulocyte/macrophage, eosinophil, megakaryocyte, erythroid and mast cell lineages.

At the other end of the spectrum is the hematopoietic growth factor that, along with the corresponding receptor, was discussed in a series of articles in the October 5, 1990 edition of Cell. The receptor is the product of the W locus, c-kit, which is a member of the class of receptor protein tyrosine kinases. The ligand for c-kit, which is referred to by various names such as stem cell factor (SCF) and mast cell growth factor (MGF), is believed to be essential for the development of early hematopoietic stem cells and cells restricted to the erythroid and mast cell lineages in mice; see, for example, Copeland et al., Cell 63, 175-183 (1990).

It appears, therefore, that there are growth factors

that exclusively affect mature cells. There also appear to be growth factors that affect both mature cells and stem cells. The growth factors that affect both types of cells may affect a small number or a large number of mature cells.

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There further appears to be an inverse relationship between the ability of a growth factor to affect mature cells and the ability of the growth factor to affect stem cells. For example, the c-kit ligand, which stimulates a small number of mature cells, is believed to be more important in the renewal and development of stem cells than is IL-3, which is reported to stimulate proliferation of many mature cells (see above).

15

Prior to the present specification, there have been no reports of growth factors that exclusively stimulate stem cells in the absence of an effect on mature cells. The discovery of such growth factors would be of particular significance.

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As mentioned above, c-kit is a protein tyrosine kinase (pTK). It is becoming increasingly apparent that the protein tyrosine kinases play an important role as cellular receptors for hematopoietic growth factors. Other receptor pTKs include the receptors of colony stimulating factor 1 (CSF-1) and PDGF.

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The pTK family can be recognized by the presence of several conserved amino acid regions in the catalytic domain. These conserved regions are summarized by Hanks et al. in Science 241, 42-52 (1988), see Figure 1 starting on page 46 and by Wilks in Proc. Natl. Acad. Sci. USA 86, 1603-1607 (1989), see Figure 2 on page 1605.

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Additional protein tyrosine kinases that represent hematopoietic growth factor receptors are needed in order more effectively to stimulate the self-renewal of the totipotent hematopoietic stem cell and to stimulate the

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development of all cells of the hematopoietic system both in vitro and in vivo. Novel hematopoietic growth factor receptors that are present only on primitive stem cells, but are not present on mature progenitor cells, are particularly desired. Ligands for the novel receptors are also desirable to act as hematopoietic growth factors. Nucleic acid sequences encoding the receptors and ligands are needed to produce recombinant receptors and ligands.

SUMMARY OF THE INVENTION

These and other objectives as will be apparent to those with ordinary skill in the art have been met by providing isolated mammalian nucleic acid molecules encoding receptor protein tyrosine kinases expressed in primitive hematopoietic cells and not expressed in mature hematopoietic cells. Also included are the receptors encoded by such nucleic acid molecules; the nucleic acid molecules encoding receptor protein tyrosine kinases having the sequences shown in Figure 1a (murine flk-2), Figure 1b (human flk-2) and Figure 2 (murine flk-1); the receptor protein tyrosine kinases having the amino acid sequences shown in Figure 1a, Figure 1b and Figure 2; ligands for the receptors; nucleic acid sequences that encode the ligands; and methods of stimulating the proliferation of primitive mammalian hematopoietic stem cells comprising contacting the stem cells with a ligand that binds to a receptor protein tyrosine kinase expressed in primitive mammalian hematopoietic cells and not expressed in mature hematopoietic cells.

DESCRIPTION OF THE FIGURES

Figure 1a.1 through 1a.5 shows the cDNA and amino acid sequences of murine flk-2. All subsequent references to Figure 1a are intended to refer to Figure 1a.1 through 1a.5. The amino acid residues occur directly below the nucleotides in the open reading frame. Amino acids -27 to -1 constitute the hydrophobic leader sequence. Amino acids 1 to 517 constitute the extracellular receptor domain. Amino acids

518 to 537 constitute the transmembrane region. Amino acids 538 to 966 constitute the intracellular catalytic domain. Counting amino acid residue -27 as residue number 1, the following amino acid residues in the intracellular domain are catalytic sub-domains identified by Hanks (see above): 618-623, 811-819, 832-834, 857-862, 872-878. The sequence at residues 709-785 is a signature sequence characteristic of flk-2. The protein tyrosine kinases generally have a signature sequence in this region. (See SEQ. ID. NOS. 1-2)

Figure 1b.1 through 1b.5 shows the complete cDNA and amino acid sequences of human flk-2 receptor. All subsequent references to Figure 1b are intended to refer to Figure 1b.1 through 1b.5. Amino acids -27 to -1 constitute the hydrophobic leader sequence. Amino acids 1 to 516 constitute the extracellular receptor domain. Amino acids 517 to 536 constitute the transmembrane region. Amino acids 537 to 966 constitute the intracellular catalytic domain. (See SEQ. ID. NOS. 3-4)

Figure 2.1 through 2.7 shows the cDNA and amino acid sequences of murine flk-1. All subsequent references to Figure 2 are intended to refer to Figure 2.1 through 2.7. Amino acids -19 to -1 constitute the hydrophobic leader sequence. Amino acids 1 to 743 constitute the extracellular receptor domain. Amino acids 744 to 765 constitute the transmembrane region. Amino acids 766 to 1348 constitute the intracellular catalytic domain. (See SEQ. ID. NOS. 5-6)

Figure 3 shows the time response of binding between a murine stromal cell line (2018) and APTag-flk-2 as well as APTag-flk-1. APTag without receptor (SEAP) is used as a control. See Example 8.

Figure 4 shows the dose response of binding between stromal cells (2018) and APTag-flk-2 as well as APTag-flk-1. APTag without receptor (SEAP) is used as a control. See Example 8.

DETAILED DESCRIPTION OF THE INVENTIONR ceptors

5 In one embodiment, the invention relates to an isolated mammalian nucleic acid molecule encoding a receptor protein tyrosine kinase expressed in primitive mammalian hematopoietic cells and not expressed in mature hematopoietic cells.

10 The nucleic acid molecule may be a DNA, cDNA, or RNA molecule. The mammal in which the nucleic acid molecule exists may be any mammal, such as a mouse, rat, rabbit, or human.

15 The nucleic acid molecule encodes a protein tyrosine kinase (pTK). Members of the pTK family can be recognized by the conserved amino acid regions in the catalytic domains. Examples of pTK consensus sequences have been provided by
20 Hanks et al. in Science 241, 42-52 (1988); see especially Figure 1 starting on page 46 and by Wilks in Proc. Natl. Acad. Sci. USA 86, 1603-1607 (1989); see especially Figure 2 on page 1605. A methionine residue at position 205 in the conserved sequence WMAPES is characteristic of pTK's that are
25 receptors.

 The Hanks et al article identifies eleven catalytic sub-domains containing pTK consensus residues and sequences. The pTKs of the present invention will have most or all of these
30 consensus residues and sequences.

 Some particularly strongly conserved residues and sequences are shown in Table 1.

TABLE 1Conserved Residues and Sequences in pTKs¹

	<u>Position²</u>	<u>Residue or Sequence</u>	<u>Catalytic Domain</u>
5	50	G	I
	52	G	I
	57	V	I
10	70	A	II
	72	K	II
	91	E	III
	166	D	VI
	171	N	VI
15	184-186	DFG	VII
	208	E	VIII
	220	D	IX
	225	G	IX
	280	R	XI

20

1. See Hanks et al., Science 241, 42-52 (1988)
2. Adjusted in accordance with Hanks et al., Id.

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A pTK of the invention may contain all thirteen of these highly conserved residues and sequences. As a result of natural or synthetic mutations, the pTKs of the invention may contain fewer than all thirteen strongly conserved residues and sequences, such as 11, 9, or 7 such sequences.

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The receptors of the invention generally belong to the same class of pTK sequences that c-kit belongs to. It has surprisingly been discovered, however, that a new functional class of receptor pTKs exists. The new functional class of receptor pTKs is expressed in primitive hematopoietic cells, but not expressed in mature hematopoietic cells.

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For the purpose of this specification, a primitive hematopoietic cell is totipotent, i.e. capable of reconstituting all hematopoietic blood cells in vivo. A mature hematopoietic cell is non-self-renewing, and has limited proliferative capacity - i.e., a limited ability to give rise to multiple lineages. Mature hematopoietic cells, for the purposes of this specification, are generally capable of giving rise to only one or two lineages in vitro or in vivo.

It should be understood that the hematopoietic system is complex, and contains many intermediate cells between the primitive totipotent hematopoietic stem cell and the totally committed mature hematopoietic cells defined above. As the stem cell develops into increasingly mature, lineage-restricted cells, it gradually loses its capacity for self-renewal.

The receptors of the present invention may and may not be expressed in these intermediate cells. The necessary and sufficient condition that defines members of the new class of receptors is that they are present in the primitive, totipotent stem cell or cells, and not in mature cells restricted only to one or, at most, two lineages.

An example of a member of the new class of receptor pTKs is called fetal liver kinase 2 (flk-2) after the organ in which it was found. There is approximately 1 totipotent stem cell per 10^4 cells in mid-gestation (day 14) fetal liver in mice. In addition to fetal liver, flk-2 is also expressed in fetal spleen, fetal thymus, adult brain, and adult marrow.

For example, flk-2 is expressed in individual multipotential CFU-Blast colonies capable of generating numerous multilineage colonies upon replating. It is likely, therefore, that flk-2 is expressed in the entire primitive (i.e. self-renewing) portion of the hematopoietic hierarchy. This discovery is consistent with flk-2 being important in transducing putative self-renewal signals from the environment.

It is particularly relevant that the expression of flk-2 mRNA occurs in the most primitive thymocyte subset. Even in two closely linked immature subsets that differ in expression of the IL-2 receptor, flk-2 expression segregates to the more primitive subset lacking an IL-2 receptor. The earliest thymocyte subset is believed to be uncommitted. Therefore, the thymocytes expressing flk-2 may be multipotential. flk-2 is the first receptor tyrosine kinase known to be expressed

in the T-lymphoid lineage.

5 The fetal liver mRNA migrates relative to 28S and 18S ribosomal bands on formaldehyde agarose gels at approximately 3.5 kb, while the brain message is considerably larger. In adult tissues, flk-2 m-RNA from both brain and bone marrow migrated at approximately 3.5 kb.

10 A second pTK receptor is also included in the present invention. This second receptor, which is called fetal liver kinase 1 (flk-1), is not a member of the same class of receptors as flk-2, since flk-1 may be found in some more mature hematopoietic cells. The amino acid sequence of murine flk-1 is given in Figure 2.

15 The present invention includes the flk-1 receptor as well as DNA, cDNA and RNA encoding flk-1. The DNA sequence of murine flk-1 is also given in Figure 2. Flk-1 may be found in the same organs as flk-2, as well as in fetal brain, stomach, kidney, lung, heart and intestine; and in adult kidney, heart, spleen, lung, muscle, and lymph nodes.

25 The receptor protein tyrosine kinases of the invention are known to be divided into easily found domains. The DNA sequence corresponding to the pTKs encode, starting at their 5'-ends, a hydrophobic leader sequence followed by a hydrophilic extracellular domain, which binds to, and is activated by, a specific ligand. Immediately downstream from the extracellular receptor domain, is a hydrophobic transmembrane region. The transmembrane region is immediately followed by a basic catalytic domain, which may easily be identified by reference to the Hanks et al. and Wilks articles discussed above.

35 The following table shows the nucleic acid and amino acid numbers that correspond to the signal peptide, the extracellular domain, the transmembrane region and the intracellular domain for murine flk-1 (mflk-1), murine flk-2 (mflk-2) and human flk-2 (hflk-2).

mFLK-1

	<u>Signal Peptide</u>	<u>Extracellular</u>	<u>Transmembrane</u>	<u>Intracellular</u>
	aa # -19 to -1	1 to 743	744 to 765	766 to 1348
	aa code M A	A E	V V	R A
5	na # 208-264	265-2493	2494-2559	2560-4308

mFLK-2

	<u>Signal Peptide</u>	<u>Extracellular</u>	<u>Transmembrane</u>	<u>Intracellular</u>
	aa # -27 to -1	1 to 517	518 to 537	538 to 966
10	aa code M T	N S	F C	H S
	na # 31-111	112-1662	1663-1722	1723-3006

hFLK-2

	<u>Signal Peptide</u>	<u>Extracellular</u>	<u>Transmembrane</u>	<u>Intracellular</u>
15	aa # -27 to -1	1 to 516	517 to 536	537 to 966
	aa code M N	Q F	Y C	H S
	na # 58-138	139-1689	1690-1746	1747-3036

20 The present invention includes the extracellular receptor domain lacking the transmembrane region and catalytic domain. Preferably, the hydrophobic leader sequence is also removed from the extracellular domain. In the case of human and murine flk-2, the hydrophobic leader sequence includes amino acids 1-27.

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These regions and domains may easily be visually identified by those having ordinary skill in the art by reviewing the amino acid sequence in a suspected pTK and comparing it to known pTKs. For example, referring to Figure 30 1a, the transmembrane region of flk-2, which separates the extracellular receptor domain from the catalytic domain, is encoded by nucleotides 1663 (T) to 1722 (C). These nucleotides correspond to amino acid residues 545 (Phe) to 564 (Cys). The amino acid sequence between the transmembrane 35 region and the catalytic sub-domain (amino acids 618-623) identified by Hanks et al. as sub-domain I (i.e., GXGXXG) is characteristic of receptor protein tyrosine kinases.

The extracellular domain may also be identified through

commonly recognized criteria of extracellular amino acid sequences. The determination of appropriate criteria is known to those skilled in the art, and has been described, for example, by Hopp et al, Proc. Nat'l Acad. Sci. USA 78, 3824-3828 (1981); Kyte et al, J. Mol. Biol. 157, 105-132 (1982); Emini, J. Virol. 55, 836-839 (1985); Jameson et al, CA BIOS 4, 181-186 (1988); and Karplus et al, Naturwissenschaften 72, 212-213 (1985). Amino acid domains predicted by these criteria to be surface exposed characteristic of extracellular domains.

As will be discussed in more detail below, the nucleic acid molecules that encode the receptors of the invention may be inserted into known vectors for use in standard recombinant DNA techniques. Standard recombinant DNA techniques are those such as are described in Sambrook et al., "Molecular Cloning," Second Edition, Cold Spring Harbor Laboratory Press (1987) and by Ausubel et al., Eds, "Current Protocols in Molecular Biology," Green Publishing Associates and Wiley-Interscience, New York (1987). The vectors may be circular (i.e. plasmids) or non-circular. Standard vectors are available for cloning and expression in a host. The host may be prokaryotic or eucaryotic. Prokaryotic hosts are preferably E. coli. Preferred eucaryotic hosts include yeast, insect and mammalian cells. Preferred mammalian cells include, for example, CHO, COS and human cells.

Ligands

The invention also includes ligands that bind to the receptor pTKs of the invention. In addition to binding, the ligands stimulate the proliferation of additional primitive stem cells, differentiation into more mature progenitor cells, or both.

The ligand may be a growth factor that occurs naturally in a mammal, preferably the same mammal that produces the corresponding receptor. The growth factor may be isolated and purified, or be present on the surface of an isolated

population of cells, such as stromal cells.

The ligand may also be a molecule that does not occur naturally in a mammal. For example, antibodies, preferably
5 monoclonal, raised against the receptors of the invention or against anti-ligand antibodies mimic the shape of, and act as, ligands if they constitute the negative image of the receptor or anti-ligand antibody binding site. The ligand may also be a non-protein molecule that acts as a ligand when
10 it binds to, or otherwise comes into contact with, the receptor.

In another embodiment, nucleic acid molecules encoding the ligands of the invention are provided. The nucleic acid
15 molecule may be RNA, DNA or cDNA.

Stimulating Proliferation of Stem Cells

The invention also includes a method of stimulating the
20 proliferation and/or differentiation of primitive mammalian hematopoietic stem cells as defined above. The method comprises contacting the stem cells with a ligand in accordance with the present invention. The stimulation of proliferation and/or differentiation may occur in vitro or in vivo.
25 vivo.

The ability of a ligand according to the invention to stimulate proliferation of stem cells in vitro and in vivo has important therapeutic applications. Such applications
30 include treating mammals, including humans, whose primitive stem cells do not sufficiently undergo self-renewal. Example of such medical problems include those that occur when defects in hematopoietic stem cells or their related growth factors depress the number of white blood cells. Examples of
35 such medical problems include anemia, such as macrocytic and aplastic anemia. Bone marrow damage resulting from cancer chemotherapy and radiation is another example of a medical problem that would be helped by the stem cell factors of the invention.

Functional Equivalents

The invention includes functional equivalents of the pTK receptors, receptor domains, and ligands described above as well as of the nucleic acid sequences encoding them. A protein is considered a functional equivalent of another protein for a specific function if the equivalent protein is immunologically cross-reactive with, and has the same function as, the receptors and ligands of the invention. The equivalent may, for example, be a fragment of the protein, or a substitution, addition or deletion mutant of the protein.

For example, it is possible to substitute amino acids in a sequence with equivalent amino acids. Groups of amino acids known normally to be equivalent are:

- (a) Ala(A) Ser(S) Thr(T) Pro(P) Gly(G);
- (b) Asn(N) Asp(D) Glu(E) Gln(Q);
- (c) His(H) Arg(R) Lys(K);
- (d) Met(M) Leu(L) Ile(I) Val(V); and
- (e) Phe(F) Tyr(Y) Trp(W).

Substitutions, additions and/or deletions in the receptors and ligands may be made as long as the resulting equivalent receptors and ligands are immunologically cross reactive with, and have the same function as, the native receptors and ligands.

The equivalent receptors and ligands will normally have substantially the same amino acid sequence as the native receptors and ligands. An amino acid sequence that is substantially the same as another sequence, but that differs from the other sequence by means of one or more substitutions, additions and/or deletions is considered to be an equivalent sequence. Preferably, less than 25%, more preferably less than 10%, and most preferably less than 5% of the number of amino acid residues in the amino acid sequence of the native receptors and ligands are substituted for, added to, or deleted from.

Equivalent nucleic acid molecules include nucleic acid sequences that encode equivalent receptors and ligands as defined above. Equivalent nucleic acid molecules also include nucleic acid sequences that differ from native
5 nucleic acid sequences in ways that do not affect the corresponding amino acid sequences.

ISOLATION OF NUCLEIC ACID MOLECULES AND PROTEINS

10 Isolation of Nucleic Acid Molecules Encoding Receptors

In order to produce nucleic acid molecules encoding mammalian stem cell receptors, a source of stem cells is provided. Suitable sources include fetal liver, spleen, or
15 thymus cells or adult marrow or brain cells.

For example, suitable mouse fetal liver cells may be obtained at day 14 of gestation. Mouse fetal thymus cells may be obtained at day 14-18, preferably day 15, of
20 gestation. Suitable fetal cells of other mammals are obtained at gestation times corresponding to those of mouse.

Total RNA is prepared by standard procedures from stem cell receptor-containing tissue. The total RNA is used to
25 direct cDNA synthesis. Standard methods for isolating RNA and synthesizing cDNA are provided in standard manuals of molecular biology such as, for example, in Sambrook et al., "Molecular Cloning," Second Edition, Cold Spring Harbor Laboratory Press (1987) and in Ausubel et al., (Eds),
30 "Current Protocols in Molecular Biology," Greene Associates/Wiley Interscience, New York (1990).

The cDNA of the receptors is amplified by known methods. For example, the cDNA may be used as a template for
35 amplification by polymerase chain reaction (PCR); see Saiki et al., Science, 239, 487 (1988) or Mullis et al., U.S. patent 4,683,195. The sequences of the oligonucleotide primers for the PCR amplification are derived from the sequences of known receptors, such as from the sequences

given in Figures 1 and 2 for flk-2 and flk-1, respectively, preferably from flk-2. The oligonucleotides are synthesized by methods known in the art. Suitable methods include those described by Caruthers in Science 230, 281-285 (1985).

5 In order to isolate the entire protein-coding regions for the receptors of the invention, the upstream oligonucleotide is complementary to the sequence at the 5' end, preferably encompassing the ATG start codon and at least 10 5-10 nucleotides upstream of the start codon. The downstream oligonucleotide is complementary to the sequence at the 3' end, optionally encompassing the stop codon. A mixture of upstream and downstream oligonucleotides are used in the PCR amplification. The conditions are optimized for each 15 particular primer pair according to standard procedures. The PCR product is analyzed by electrophoresis for the correct size cDNA corresponding to the sequence between the primers.

20 Alternatively, the coding region may be amplified in two or more overlapping fragments. The overlapping fragments are designed to include a restriction site permitting the assembly of the intact cDNA from the fragments.

25 The amplified DNA encoding the receptors of the invention may be replicated in a wide variety of cloning vectors in a wide variety of host cells. The host cell may be prokaryotic or eukaryotic. The DNA may be obtained from natural sources and, optionally, modified, or may be 30 synthesized in whole or in part.

The vector into which the DNA is spliced may comprise segments of chromosomal, non-chromosomal and synthetic DNA sequences. Some suitable prokaryotic cloning vectors include 35 plasmids from E. coli, such as colE1, pCR1, pBR322, pMB9, pUC, pKSM, and RP4. Prokaryotic vectors also include derivatives of phage DNA such as M13 and other filamentous single-stranded DNA phages.

Isolation of Receptors

DNA encoding the receptors of the invention are inserted into a suitable vector and expressed in a suitable prokaryotic or eucaryotic host. Vectors for expressing proteins in bacteria, especially E.coli, are known. Such vectors include the PATH vectors described by Dieckmann and Tzagoloff in J. Biol. Chem. 260, 1513-1520 (1985). These vectors contain DNA sequences that encode anthranilate synthetase (TrpE) followed by a polylinker at the carboxy terminus. Other expression vector systems are based on beta-galactosidase (pEX); lambda P_L; maltose binding protein (pMAL); and glutathione S-transferase (pGST) - see Gene 67, 31 (1988) and Peptide Research 3, 167 (1990).

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Vectors useful in yeast are available. A suitable example is the 2 μ plasmid.

Suitable vectors for use in mammalian cells are also known. Such vectors include well-known derivatives of SV-40, adenovirus, retrovirus-derived DNA sequences and shuttle vectors derived from combination of functional mammalian vectors, such as those described above, and functional plasmids and phage DNA.

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Further eukaryotic expression vectors are known in the art (e.g., P.J. Southern and P. Berg, J. Mol. Appl. Genet. 1, 327-341 (1982); S. Subramani et al, Mol. Cell. Biol. 1, 854-864 (1981); R.J. Kaufmann and P.A. Sharp, "Amplification And Expression Of Sequences Cotransfected with A Modular Dihydrofolate Reductase Complementary DNA Gene," J. Mol. Biol. 159, 601-621 (1982); R.J. Kaufmann and P.A. Sharp, Mol. Cell. Biol. 159, 601-664 (1982); S.I. Scahill et al, "Expression And Characterization Of The Product Of A Human Immune Interferon DNA Gene In Chinese Hamster Ovary Cells," Proc. Natl. Acad. Sci. USA 80, 4654-4659 (1983); G. Urlaub and L.A. Chasin, Proc. Natl. Acad. Sci. USA 77, 4216-4220, (1980).

35

The expression vectors useful in the present invention contain at least one expression control sequence that is operatively linked to the DNA sequence or fragment to be expressed. The control sequence is inserted in the vector in order to control and to regulate the expression of the cloned DNA sequence. Examples of useful expression control sequences are the lac system, the trp system, the tac system, the trc system, major operator and promoter regions of phage lambda, the control region of fd coat protein, the glycolytic promoters of yeast, e.g., the promoter for 3-phosphoglycerate kinase, the promoters of yeast acid phosphatase, e.g., Pho5, the promoters of the yeast alpha-mating factors, and promoters derived from polyoma, adenovirus, retrovirus, and simian virus, e.g., the early and late promoters or SV40, and other sequences known to control the expression of genes of prokaryotic or eukaryotic cells and their viruses or combinations thereof.

Vectors containing the receptor-encoding DNA and control signals are inserted into a host cell for expression of the receptor. Some useful expression host cells include well-known prokaryotic and eukaryotic cells. Some suitable prokaryotic hosts include, for example, E. coli, such as E. coli SG-936, E. coli HB 101, E. coli W3110, E. coli X1776, E. coli X2282, E. coli DHI, and E. coli MRC1, Pseudomonas, Bacillus, such as Bacillus subtilis, and Streptomyces. Suitable eukaryotic cells include yeast and other fungi, insect, animal cells, such as COS cells and CHO cells, human cells and plant cells in tissue culture.

The human homologs of the mouse receptors described above are isolated by a similar strategy. RNA encoding the receptors are obtained from a source of human cells enriched for primitive stem cells. Suitable human cells include fetal spleen, thymus and liver cells, and umbilical cord blood as well as adult brain and bone marrow cells. The human fetal cells are preferably obtained on the day of gestation corresponding to mid-gestation in mice. The amino acid sequences of the human flk receptors as well as of the

nucleic acid sequences encoding them are homologous to the amino acid and nucleotide sequences of the mouse receptors.

In the present specification, the sequence of a first protein, such as a receptor or a ligand, or of a nucleic acid molecule that encodes the protein, is considered homologous to a second protein or nucleic acid molecule if the amino acid or nucleotide sequence of the first protein or nucleic acid molecule is at least about 30% homologous, preferably at least about 50% homologous, and more preferably at least about 65% homologous to the respective sequences of the second protein or nucleic acid molecule. In the case of proteins having high homology, the amino acid or nucleotide sequence of the first protein or nucleic acid molecule is at least about 75% homologous, preferably at least about 85% homologous, and more preferably at least about 95% homologous to the amino acid or nucleotide sequence of the second protein or nucleic acid molecule.

Combinations of mouse oligonucleotide pairs are used as PCR primers to amplify the human homologs from the cells to account for sequence divergence. The remainder of the procedure for obtaining the human flk homologs are similar to those described above for obtaining mouse flk receptors. The less than perfect homology between the human flk homologs and the mouse oligonucleotides is taken into account in determining the stringency of the hybridization conditions.

30 Assay for expression of Receptors on Stem Cells

In order to demonstrate the expression of flk receptors on the surface of primitive hematopoietic stem cells, antibodies that recognize the receptor are raised. The receptor may be the entire protein as it exists in nature, or an antigenic fragment of the whole protein. Preferably, the fragment comprises the predicted extra-cellular portion of the molecule.

Antigenic fragments may be identified by methods known in the art. Fragments containing antigenic sequences may be selected on the basis of generally accepted criteria of potential antigenicity and/or exposure. Such criteria include the hydrophilicity and relative antigenic index, as determined by surface exposure analysis of proteins. The determination of appropriate criteria is known to those skilled in the art, and has been described, for example, by Hopp et al, Proc. Nat'l Acad. Sci. USA 78, 3824-3828 (1981); Kyte et al, J. Mol. Biol. 157, 105-132 (1982); Emini, J. Virol. 55, 836-839 (1985); Jameson et al, CA BIOS 4, 181-186 (1988); and Karplus et al, Naturwissenschaften 72, 212-213 (1985). Amino acid domains predicted by these criteria to be surface exposed are selected preferentially over domains predicted to be more hydrophobic or hidden.

The proteins and fragments of the receptors to be used as antigens may be prepared by methods known in the art. Such methods include isolating or synthesizing DNA encoding the proteins and fragments, and using the DNA to produce recombinant proteins, as described above.

Fragments of proteins and DNA encoding the fragments may be chemically synthesized by methods known in the art from individual amino acids and nucleotides. Suitable methods for synthesizing protein fragments are described by Stuart and Young in "Solid Phase Peptide Synthesis," Second Edition, Pierce Chemical Company (1984). Suitable methods for synthesizing DNA fragments are described by Caruthers in Science 230, 281-285 (1985).

If the receptor fragment defines the epitope, but is too short to be antigenic, it may be conjugated to a carrier molecule in order to produce antibodies. Some suitable carrier molecules include keyhole limpet hemocyanin, Ig sequences, TrpE, and human or bovine serum albumen. Conjugation may be carried out by methods known in the art. One such method is to combine a cysteine residue of the fragment with a cysteine residue on the carrier molecule.

The antibodies are preferably monoclonal. Monoclonal antibodies may be produced by methods known in the art. These methods include the immunological method described by Kohler and Milstein in Nature 256, 495-497 (1975) and
5 Campbell in "Monoclonal Antibody Technology, The Production and Characterization of Rodent and Human Hybridomas" in Burdon et al., Eds, Laboratory Techniques in Biochemistry and Molecular Biology, Volume 13, Elsevier Science Publishers, Amsterdam (1985); as well as by the recombinant DNA method
10 described by Huse et al in Science 246, 1275-1281 (1989).

Polyclonal or monoclonal antisera shown to be reactive with receptor-encoded native proteins, such as with flk-1 and flk-2 encoded proteins, expressed on the surface of viable
15 cells are used to isolate antibody-positive cells. One method for isolating such cells is flow cytometry; see, for example, Loken et al., European patent application 317,156. The cells obtained are assayed for stem cells by engraftment into radiation-ablated hosts by methods known in the art;
20 see, for example, Jordan et al., Cell 61, 953-963 (1990).

Criteria for Novel Stem Cell Receptor Tyrosine Kinases Expressed in Stem Cells

25 Additional novel receptor tyrosine kinase cDNAs are obtained by amplifying cDNAs from stem cell populations using oligonucleotides as PCR primers; see above. Examples of suitable oligonucleotides are PTK1 and PTK2, which were described by Wilks et al. in Proc. Natl. Acad. Sci. USA 86,
30 1603-1607 (1989). Novel cDNA is selected on the basis of differential hybridization screening with probes representing known kinases. The cDNA clones hybridizing only at low stringency are selected and sequenced. The presence of the amino acid triplet DFG confirms that the sequence represents
35 a kinase. The diagnostic methionine residue in the WMAPES motif is indicative of a receptor-like kinase, as described above. Potentially novel sequences obtained are compared to available sequences using databases such as Genbank in order to confirm uniqueness. Gene-specific oligonucleotides are
40 prepared as described above based on the sequence obtained.

The oligonucleotides are used to analyze stem cell enriched and depleted populations for expression. Such cell populations in mice are described, for example, by Jordan et al. in Cell 61, 953-956 (1990); Ikuta et al. in Cell 62, 863-864 (1990); Spangrude et al. in Science 241, 58-62 (1988); and Szilvassy et al. in Blood 74, 930-939 (1989). Examples of such human cell populations are described as CD33⁻CD34⁺ by Andrews et al. in the Journal of Experimental Medicine 169, 1721-1731 (1989). Other human stem cell populations are described, for example, in Civin et al., European Patent Application 395,355 and in Loken et al., European Patent Application 317,156.

15

Isolating Ligands and Nucleic Acid Molecules Encoding Ligands

Cells that may be used for obtaining ligands include stromal cells, for example stromal cells from fetal liver, fetal spleen, fetal thymus and fetal or adult bone marrow. Cell lines expressing ligands are established and screened.

For example, cells such as stromal (non-hematopoietic) cells from fetal liver are immortalized by known methods. Examples of known methods of immortalizing cells include transduction with a temperature sensitive SV40 T-antigen expressed in a retroviral vector. Infection of fetal liver cells with this virus permits the rapid and efficient establishment of multiple independent cell lines. These lines are screened for ligand activity by methods known in the art, such as those outlined below.

Ligands for the receptors of the invention, such as flk-1 and flk-2, may be obtained from the cells in several ways. For example, a bioassay system for ligand activity employs chimeric tagged receptors; see, for example, Flanagan et al., Cell 63, 185-194 (1990). One strategy measures ligand binding directly via a histochemical assay. Fusion proteins comprising the extracellular receptor domains and secretable alkaline phosphatase (SEAP) are constructed and transfected

into suitable cells such as NIH/3T3 or COS cells. Flanagan et al. refer to such DNA or amino acid constructs as APTag followed by the name of the receptor - i.e. APTag-c-kit. The fusion proteins bind with high affinity to cells expressing surface-bound ligand. Binding is detectable by the enzymatic activity of the alkaline phosphatase secreted into the medium. The bound cells, which are often stromal cells, are isolated from the APTag-receptor complex.

For example, some stromal cells that bind APTag-flk1 and APTag-flk2 fusion proteins include mouse fetal liver cells (see example 1); human fetal spleen cells (see example 3); and human fetal liver (example 3). Some stromal fetal thymus cells contain flk-1 ligand (example 3).

To clone the cDNA that encodes the ligand, a cDNA library is constructed from the isolated stromal cells in a suitable expression vector, preferably a phage such as CDM8, pSV Sport (BRL Gibco) or pIH3, (Seed et al., Proc. Natl. Acad. Sci. USA 84, 3365-3369 (1987)). The library is transfected into suitable host cells, such as COS cells. Cells containing ligands on their surface are detected by known methods, see above.

In one such method, transfected COS cells are distributed into single cell suspensions and incubated with the secreted alkaline phosphatase-flk receptor fusion protein, which is present in the medium from NIH/3T3 or COS cells prepared by the method described by Flanagan et al., see above. Alkaline phosphatase-receptor fusion proteins that are not bound to the cells are removed by centrifugation, and the cells are panned on plates coated with antibodies to alkaline phosphatase. Bound cells are isolated following several washes with a suitable wash reagent, such as 5% fetal bovine serum in PBS, and the DNA is extracted from the cells. Additional details of the panning method described above may be found in an article by Seed et al., Proc. Natl. Acad. Sci. USA 84, 3365-3369 (1987).

In a second strategy, the putative extracellular ligand binding domains of the receptors are fused to the transmembrane and kinase domains of the human c-fms tyrosine kinase and introduced into 3T3 fibroblasts. The human c-fms kinase is necessary and sufficient to transduce proliferative signals in these cells after appropriate activation i.e. with the flk-1 or flk-2 ligand. The 3T3 cells expressing the chimeras are used to screen putative sources of ligand in a cell proliferation assay.

An alternate approach for isolating ligands using the fusion receptor-expressing 3T3 cells and insertional activation is also possible. A retrovirus is introduced into random chromosomal positions in a large population of these cells. In a small fraction, the retrovirus is inserted in the vicinity of the ligand-encoding gene, thereby activating it. These cells proliferate due to autocrine stimulation of the receptor. The ligand gene is "tagged" by the retrovirus, thus facilitating its isolation.

Examples

Example 1. Cells containing mouse flk-1 and flk-2 ligands. Murine stromal cell line 2018.

In order to establish stromal cell lines, fetal liver cells are disaggregated with collagen and grown in a mixture of Dulbecco's Modified Eagle's Medium (DMEM) and 10% heat-inactivated fetal calf serum at 37°C. The cells are immortalized by standard methods. A suitable method involves introducing DNA encoding a growth regulating- or oncogene-encoding sequence into the target host cell. The DNA may be introduced by means of transduction in a recombinant viral particle or transfection in a plasmid. See, for example, Hammerschmidt et al., Nature 340, 393-397 (1989) and Abcouwer et al, Biotechnology 7, 939-946 (1989). Retroviruses are the preferred viral vectors, although SV40 and Epstein-Barr virus can also serve as donors of the growth-enhancing sequences. A suitable retrovirus is the ecotropic retrovirus containing

a temperature sensitive SV40 T-antigen (tsA58) and a G418 resistance gene described by McKay in Cell 66, 713-729 (1991). After several days at 37°C, the temperature of the medium is lowered to 32°C. Cells are selected with G418 (0.5 mg/ml). The selected cells are expanded and maintained.

A mouse stromal cell line produced by this procedure is called 2018 and was deposited on October 30, 1991 in the American Type Culture Collection, Rockville, Maryland, USA (ATCC); accession number CRL 10907.

Example 2. Cells containing human flk-1 and flk-2 ligands.

Human fetal liver (18, 20, and 33 weeks after abortion), spleen (18 weeks after abortion), or thymus (20 weeks after abortion) is removed at the time of abortion and stored on ice in a balanced salt solution. After mincing into 1 mm fragments and forcing through a wire mesh, the tissue is washed one time in Hanks Balanced Salt Solution (HBSS).

The disrupted tissue is centrifuged at 200 xg for 15 minutes at room temperature. The resulting pellet is resuspended in 10-20 ml of a tissue culture grade trypsin-EDTA solution (Flow Laboratories). The resuspended tissue is transferred to a sterile flask and stirred with a stirring bar at room temperature for 10 minutes. One ml of heat-inactivated fetal bovine calf serum (Hyclone) is added to a final concentration of 10% in order to inhibit trypsin activity. Collagenase type IV (Sigma) is added from a stock solution (10 mg/ml in HBSS) to a final concentration of 100 ug/ml in order to disrupt the stromal cells. The tissue is stirred at room temperature for an additional 2.5 hours; collected by centrifugation (400xg, 15 minutes); and resuspended in "stromal medium," which contains Iscove's modification of DMEM supplemented with 10% heat-inactivated fetal calf serum, 5% heat-inactivated human serum (Sigma), 4 mM L-glutamine, 1x sodium pyruvate, (stock of 100x Sigma), 1x non-essential amino acids (stock of 100x, Flow), and a mixture of antibiotics kanomycin, neomycin, penicillin,

streptomycin. Prior to resuspending the pellet in the stromal medium, the pellet is washed one time with HBSS. It is convenient to suspend the cells in 60 ml of medium. The number of cultures depends on the amount of tissue.

5 Example 3. Isolating Stromal cells

10 Resuspended Cells (example 2) that are incubated at 37°C with 5% carbon dioxide begin to adhere to the plastic plate within 10-48 hours. Confluent monolayers may be observed within 7-10 days, depending upon the number of cells plated in the initial inoculum. Non-adherent and highly refractile cells adhering to the stromal cell layer as colonies are
15 separately removed by pipetting and frozen. Non-adherent cells are likely sources of populations of self-renewing stem cells containing flk-2. The adherent stromal cell layers are frozen in aliquots for future studies or expanded for growth in culture.

20 An unexpectedly high level of APTag-flk-2 fusion protein binding to the fetal spleen cells is observed. Two fetal spleen lines are grown in "stromal medium," which is described in example 2.

25 Non-adherent fetal stem cells attach to the stromal cells and form colonies (colony forming unit - CFU). Stromal cells and CFU are isolated by means of sterile glass cylinders and expanded in culture. A clone, called Fsp
30 62891, contains the flk-2 ligand. Fsp 62891 was deposited in the American Type Culture Collection, Rockville, Maryland, U.S.A on November 21, 1991, accession number CRL 10935.

35 Fetal liver and fetal thymus cells are prepared in a similar way. Both of these cell types produce ligands of flk-1 and, in the case of liver, some flk-2. One such fetal thymus cell line, called F.thy 62891, and one such fetal liver cell line, called FL 62891, were deposited in the American Type Culture Collection, Rockville, Maryland, U.S.A
40 on November 21, 1991 and April 2, 1992, respectively,

accession numbers CRL 10936 and CRL 11005, respectively.

Stable human cell lines are prepared from fetal cells with the same temperature sensitive immortalizing virus used to prepare the murine cell line described in example 1.

Example 4. Isolation of human stromal cell clone

Highly refractile cells overgrow patches of stromal cells, presumably because the stromal cells produce factors that allow the formation of the CFU. To isolate stromal cell clones, sterile glass cylinders coated with vacuum grease are positioned over the CFU. A trypsin-EDTA solution (100 ml) is added in order to detach the cells. The cells are added to 5 ml of stromal medium and each (clone) plated in a single well of 6-well plate.

Example 5. Plasmid (AP-tag) for expressing secretable alkaline phosphatase (SEAP)

20

Plasmids that express secretable alkaline phosphatase are described by Flanagan and Leder in Cell 63, 185-194 (1990). The plasmids contain a promoter, such as the LTR promoter; a polylinker, including HindIII and BglII; DNA encoding SEAP; a poly-A signal; and ampicillin resistance gene; and replication site.

Example 6. Plasmid for expressing APtag-flk-2 and APtag-flk-1 fusion proteins

30

Plasmids that express fusion proteins of SEAP and the extracellular portion of either flk-1 or flk-2 are prepared in accordance with the protocols of Flanagan and Leder in Cell 63, 185-194 (1990) and Berger et al., Gene 66, 1-10 (1988). Briefly, a HindIII-Bam HI fragment containing the extracellular portion of flk-1 or flk-2 is prepared and inserted into the HindIII-BglII site of the plasmid described in example 5.

35

40

Example 7. Production Of APTaq-flk-1 Or -flk-2 Fusion
Prot in

5 The plasmids from Example 6 are transfected into Cos-7
cells by DEAE-dextran (as described in Current Protocols in
Molecular Biology, Unit 16.13, "Transient Expression of
Proteins Using Cos Cells," 1991); and cotransfected with a
selectable marker, such as pSV7neo, into NIH/3T3 cells by
10 calcium precipitation. The NIH/3T3 cells are selected with
600 µg/ml G418 in 100 mm plates. Over 300 clones are screened
for secretion of placental alkaline phosphatase activity.
The assay is performed by heating a portion of the
supernatant at 65°C for 10 minutes to inactivate background
15 phosphatase activity, and measuring the OD₄₀₅ after incubating
with 1M diethanolamine (pH 9.8), 0.5 mM MgCl₂, 10 mM L-
homoarginine (a phosphatase inhibitor), 0.5 mg/ml BSA, and 12
mM p-nitrophenyl phosphate. Human placental alkaline
phosphatase is used to perform a standard curve. The APTaq-
20 flk-1 clones (F-1AP21-4) produce up to 10 µg alkaline
phosphatase activity/ml and the APTaq-flk-2 clones (F-2AP26-
0) produce up to 0.5 µg alkaline phosphatase activity/ml.

25 Example 8. Assay For APTaq-flk-1 Or APTaq-flk-2 Binding To
Cells

30 The binding of APTaq-flk-1 or APTaq-flk-2 to cells
containing the appropriate ligand is assayed by standard
methods. See, for example, Flanagan and Leder, Cell 63:185-
194, 1990). Cells (i.e., mouse stromal cells, human fetal
liver, spleen or thymus, or various control cells) are grown
to confluency in six-well plates and washed with HBHA (Hank's
balanced salt solution with 0.5 mg/ml BSA, 0.02% NaN₃, 20 mM
35 HEPES, pH 7.0). Supernatants from transfected COS or NIH/3T3
cells containing either APTaq-flk-1 fusion protein, APTaq-
flk-2 fusion protein, or APTaq without a receptor (as a
control) are added to the cell monolayers and incubated for
two hours at room temperature on a rotating platform. The
40 concentration of the APTaq-flk-1 fusion protein, APTaq-flk-2
fusion protein, or APTaq without a receptor is 60 ng/ml of

alkaline phosphatase as determined by the standard alkaline phosphatase curve (see above). The cells are then rinsed seven times with HBHA and lysed in 350 μ l of 1% Triton X-100, 10 mM Tris-HCl (pH 8.0). The lysates are transferred to a microfuge tube, along with a further 150 μ l rinse with the same solution. After vortexing vigorously, the samples are centrifuged for five minutes in a microfuge, heated at 65°C for 12 minutes to inactivate cellular phosphatases, and assayed for phosphatase activity as described previously. Results of experiments designed to show the time and dose responses of binding between stromal cells containing the ligands to flk-2 and flk-1 (2018) and APTag-flk-2, APTag-flk-1 and APTag without receptor (as a control) are shown in Figures 3 and 4, respectively.

Example 8A. Plasmids for expressing flk1/fms and flk2/fms fusion proteins

Plasmids that express fusion proteins of the extracellular portion of either flk-1 or flk-2 and the intracellular portion of c-fms (also known as colony-stimulating factor-1 receptor) are prepared in a manner similar to that described under Example 6 (Plasmid for expressing APTag-flk-2 and APTag-flk-1 fusion proteins). Briefly, a Hind III - Bam HI fragment containing the extracellular portion of flk1 or flk2 is prepared and inserted into the Hind III - Bgl II site of a pLH expression vector containing the intracellular portion of c-fms.

8B. Expression of flk1/fms or flk2/fms in 3T3 cells

The plasmids from Example 11 are transfected into NIH/3T3 cells by calcium. The intracellular portion of c-fms is detected by Western blotting.

Example 9. Cloning and Expression of cDNA Coding For Mouse Ligand T flk-1 and flk-2 Receptors

cdNA expressing mouse ligand for flk-1 and flk-2 is prepared by known methods. See, for example, Seed, B., and Aruffo, A. PNAS 84:3365-3369, 1987; Simmons, D. and Seed, B. J. Immunol. 141:2797-2800; and D'Andrea, A.D., Lodish, H.F. and Wong, G.G. Cell 57:277-285, 1989).

The protocols are listed below in sequence: (a) RNA isolation; (b) poly A RNA preparation; (c) cdNA synthesis; (d) cdNA size fractionation; (e) propagation of plasmids (vector); (f) isolation of plasmid DNA; (g) preparation of vector pSV Sport (BRL Gibco) for cloning; (h) compilation of buffers for the above steps; (i) Transfection of cdNA encoding Ligands in Cos 7 Cells; (j) panning procedure; (k) Expression cloning of flk-1 or flk-2 ligand by establishment of an autocrine loop.

9a. Guanidinium thiocyanate/LiCl Protocol for RNA Isolation

For each ml of mix desired, 0.5 g guanidine thiocyanate (GuSCN) is dissolved in 0.55 ml of 25% LiCl (stock filtered through 0.45 micron filter). 20 μ l of mercaptoethanol is added. (The resulting solution is not good for more than about a week at room temperature.)

The 2018 stromal cells are centrifuged, and 1 ml of the solution described above is added to up to 5×10^7 cells. The cells are sheared by means of a polytron until the mixture is non-viscous. For small scale preparations ($<10^8$ cells), the sheared mixture is layered on 1.5 ml of 5.7M CsCl (RNase free; 1.26 g CsCl added to every ml 10 mM EDTA pH8), and overlaid with RNase-free water if needed. The mixture is spun in an SW55 rotor at 50 krpm for 2 hours. For large scale preparations, 25 ml of the mixture is layered on 12 ml CsCl in an SW28 tube, overlaid as above, and spun at 24 krpm for 8 hours. The contents of the tube are aspirated carefully with a sterile pasteur pipet connected to a vacuum flask. Once past the CsCl interface, a band around the tube is scratched with the pipet tip to prevent creeping of the layer on the wall down the tube. The remaining CsCl

solution is aspirated. The resulting pellet is taken up in water, but not redissolved. 1/10 volume of sodium acetate and three volumes of ethanol are added to the mixture, and spun. The pellet is resuspended in water at 70°C, if necessary. The concentration of the RNA is adjusted to 1 mg/ml and frozen.

It should be noted that small RNA molecules (e.g., 5S) do not come down. For small amounts of cells, the volumes are scaled down, and the mixture is overlaid with GuSCN in RNase-free water on a gradient (precipitation is inefficient when RNA is dilute).

9b. Poly A⁺ RNA preparation

(All buffers mentioned are compiled separately below)

A disposable polypropylene column is prepared by washing with 5M NaOH and then rinsing with RNase-free water. For each milligram of total RNA, approximately 0.3 ml (final packed bed) of oligo dT cellulose is added. The oligo dT cellulose is prepared by resuspending approximately 0.5 ml of dry powder in 1 ml of 0.1M NaOH and transferring it into the column, or by percolating 0.1M NaOH through a previously used column. The column is washed with several column volumes of RNase-free water until the pH is neutral, and rinsed with 2-3 ml of loading buffer. The column bed is transferred to a sterile 15 ml tube using 4-6 ml of loading buffer.

Total RNA from the 2018 cell line is heated to 70°C for 2-3 minutes. LiCl from RNase-free stock is added to the mixture to a final concentration of 0.5M. The mixture is combined with oligo dT cellulose in the 15 ml tube, which is vortexed or agitated for 10 minutes. The mixture is poured into the column, and washed with 3 ml loading buffer, and then with 3 ml of middle wash buffer. The mRNA is eluted directly into an SW55 tube with 1.5 ml of 2 mM EDTA and 0.1% SDS, discarding the first two or three drops.

The eluted mRNA is precipitated by adding 1/10 volume of 3M sodium acetate and filling the tube with ethanol. The contents of the tube are mixed, chilled for 30 minutes at -20°C, and spun at 50 krpm at 5°C for 30 minutes. After the ethanol is decanted, and the tube air dried, the mRNA pellet is resuspended in 50-100 µl of RNase-free water. 5 µl of the resuspended mRNA is heated to 70°C in MOPS/EDTA/formaldehyde, and examined on an RNase-free 1% agarose gel.

9c. cDNA Synthesis

The protocol used is a variation of the method described by Gubler and Hoffman in Gene 25, 263-270 (1983).

1. First Strand. 4 µg of mRNA is added to a microfuge tube, heated to approximately 100°C for 30 seconds, quenched on ice. The volume is adjusted to 70 µl with RNase-free water. 20 µl of RT1 buffer, 2 µl of RNase inhibitor (Boehringer 36 u/µl), 1 µl of 5 µg/µl of oligo dT (Collaborative Research), 2.5 µl of 20 mM dXTP's (ultrapure - US Biochemicals), 1 µl of 1M DTT and 4 µl of RT-XL (Life Sciences, 24 u/µl) are added. The mixture is incubated at 42°C for 40 minutes, and inactivated by heating at 70°C for 10 minutes.

2. Second Strand. 320 µl of RNase-free water, 80 µl of RT2 buffer, 5 µl of DNA Polymerase I (Boehringer, 5 U/µl), 2 µl RNase H (BRL 2 u/µl) are added to the solution containing the first strand. The solution is incubated at 15°C for one hour and at 22°C for an additional hour. After adding 20 µl of 0.5M EDTA, pH 8.0, the solution is extracted with phenol and precipitated by adding NaCl to 0.5M linear polyacrylamide (carrier) to 20 µg/ml, and filling the tube with EtOH. The tube is spun for 2-3 minutes in a microfuge, vortexed to dislodge precipitated material from the wall of the tube, and respun for one minute.

3. Adaptors. Adaptors provide specific restriction sites to facilitate cloning, and are available from BRL

Gibco, New England Biolabs, etc. Crude adaptors are resuspended at a concentration of 1 $\mu\text{g}/\mu\text{l}$. MgSO_4 is added to a final concentration of 10 mM, followed by five volumes of EtOH. The resulting precipitate is rinsed with 70% EtOH and resuspended in TE at a concentration of 1 $\mu\text{g}/\mu\text{l}$. To kinase, 5 25 μl of resuspended adaptors is added to 3 μl of 10X kinasing buffer and 20 units of kinase. The mixture is incubated at 37°C overnight. The precipitated cDNA is resuspended in 240 μl of TE (10/1). After adding 30 μl of 10 10X low salt buffer, 30 μl of 10X ligation buffer with 0.1mM ATP, 3 μl (2.4 μg) of kinased 12-mer adaptor sequence, 2 μl (1.6 μg) of kinased 8-mer adaptor sequence, and 1 μl of T4 DNA ligase (BioLabs, 400 u/ μl , or Boehringer, 1 Weiss unit ml), the mixture is incubated at 15°C overnight. The cDNA is 15 extracted with phenol and precipitated as above, except that the extra carrier is omitted, and resuspended in 100 μl of TE.

9d. cDNA Size Fractionation.

20 A 20% KOAc, 2 mM EDTA, 1 $\mu\text{g}/\text{ml}$ ethidium bromide solution and a 5% KOAc, 2 mM EDTA, 1 $\mu\text{g}/\text{ml}$ ethidium bromide solution are prepared. 2.6 ml of the 20% KOAc solution is added to the back chamber of a small gradient maker. Air bubbles are 25 removed from the tube connecting the two chambers by allowing the 20% solution to flow into the front chamber and forcing the solution to return to the back chamber by tilting the gradient maker. The passage between the chambers is closed, and 2.5 ml of 5% solution is added to the front chamber. Any 30 liquid in the tubing from a previous run is removed by allowing the 5% solution to flow to the end of the tubing, and then to return to its chamber. The apparatus is placed on a stirplate, and, with rapid stirring, the topcock connecting the two chambers and the front stopcock are 35 opened. A polyallomer SW55 tube is filled from the bottom with the KOAc solution. The gradient is overlaid with 100 μl of cDNA solution, and spun for three hours at 50k rpm at 22°C. To collect fractions from the gradient, the SW55 tube is pierced close to the bottom of the tube with a butterfly

infusion set (with the luer hub clipped off). Three 0.5 ml fractions and then six 0.25 ml fractions are collected in microfuge tubes (approximately 22 and 11 drops, respectively). The fractions are precipitated by adding linear polyacrylamide to 20 μ g/ml and filling the tube to the top with ethanol. The tubes are cooled, spun in a microfuge tube for three minutes, vortexed, and respun for one minute. The resulting pellets are rinsed with 70% ethanol and respun, taking care not to permit the pellets to dry to completion. Each 0.25 ml fraction is resuspended in 10 μ l of TE, and 1 μ l is run on a 1% agarose minigel. The first three fractions, and the last six which contain no material smaller than 1 kb are pooled.

15 9e. Propagation of Plasmids

SupF plasmids are selected in nonsuppressing bacterial hosts containing a second plasmid, p3, which contains amber mutated ampicillin and tetracycline drug resistance elements. See Seed, Nucleic Acids Res., 11, 2427-2445 (1983). The p3 plasmid is derived from RP1, is 57 kb in length, and is a stably maintained, single copy episome. The ampicillin resistance of this plasmid reverts at a high rate so that amp^r plasmids usually cannot be used in p3-containing strains. Selection for tetracycline resistance alone is almost as good as selection for ampicillin-tetracycline resistance. However, spontaneous appearance of chromosomal suppressor tRNA mutations presents an unavoidable background (frequency about 10^{-9}) in this system. Colonies arising from spontaneous suppressor mutations are usually larger than colonies arising from plasmid transformation. Suppressor plasmids are selected in Luria broth (LB) medium containing ampicillin at 12.5 μ g/ml and tetracycline at 7.5 μ g/ml. For scaled-up plasmid preparations, M9 Casamino acids medium containing glycerol (0.8%) is employed as a carbon source. The bacteria are grown to saturation.

Alternatively, pSV Sport (BRL, Gaithersburg, Maryland) may be employed to provide SV40 derived sequences for

replication, transcription initiation and termination in COS 7 cells, as well as those sequences necessary for replication and ampicillin resistance in E. coli.

5 9f. Isolation of Vector DNA/Plasmid

One liter of saturated bacterial cells are spun down in J6 bottles at 4.2k rpm for 25 minutes. The cells are resuspended in 40 ml 10 mM EDTA, pH 8. 80 ml 0.2M NaOH and 1% SDS are added, and the mixture is swirled until it is clear and viscous. 40 ml 5M KOAc, pH 4.7 (2.5M KOAc, 2.5M HOAc) is added, and the mixture is shaken semi-vigorously until the lumps are approximately 2-3 mm in size. The bottle is spun at 4.2k rpm for 5 minutes. The supernatant is poured through cheesecloth into a 250 ml bottle, which is then filled with isopropyl alcohol and centrifuged at 4.2k rpm for 5 minutes. The bottle is gently drained and rinsed with 70% ethanol, taking care not to fragment the pellet. After inverting the bottle and removing traces of ethanol, the mixture is resuspended in 3.5 ml Tris base/EDTA (20 mM/10 mM). 3.75 ml of resuspended pellet and 0.75 ml 10 mg/ml ethidium bromide are added to 4.5 g CsCl. VTi80 tubes are filled with solution, and centrifuged for at least 2.5 hours at 80k rpm. Bands are extracted by visible light with 1 ml syringe and 20 gauge or lower needle. The top of the tube is cut off with scissors, and the needle is inserted upwards into the tube at an angle of about 30 degrees with respect to the tube at a position about 3 mm beneath the band, with the bevel of the needle up. After the band is removed, the contents of the tube are poured into bleach. The extracted band is deposited in a 13 ml Sarstedt tube, which is then filled to the top with n-butanol saturated with 1M NaCl extract. If the amount of DNA is large, the extraction procedure may be repeated. After aspirating the butanol into a trap containing 5M NaOH to destroy ethidium, an approximately equal volume of 1M ammonium acetate and approximately two volumes of 95% ethanol are added to the DNA, which is then spun at 10k rpm for 5 minutes. The pellet is rinsed carefully with 70% ethanol, and dried with a swab

or lyophilizer.

9g. Preparation of Vector for Cloning

5 20 μ g of vector is cut in a 200 μ l reaction with 100
units of BstXI (New York Biolabs) at 50°C overnight in a well
thermostated, circulating water bath. Potassium acetate
solutions (5 and 20%) are prepared in 5W55 tubes as described
above. 100 μ l of the digested vector is added to each tube
10 and spun for three hours, 50k rpm at 22°C. Under 300 nm UV
light, the desired band is observed to migrate 2/3 of the
length of the tube. Forward trailing of the band indicates
that the gradient is overloaded. The band is removed with a
1 ml syringe fitted with a 20 gauge needle. After adding
15 linear polyacrylamide and precipitating the plasmid by adding
three volumes of ethanol, the plasmid is resuspended in 50 μ l
of TE. Trial ligations are carried out with a constant
amount of vector and increasing amounts of cDNA. Large scale
ligation are carried out on the basis of these trial
20 ligations. Usually the entire cDNA prep requires 1-2 μ g of
cut vector.

9h. Buffers

25 Loading Buffer: .5M LiCl, 10 mM Tris pH 7.5, 1 mM
EDTA .1% SDS.
Middle Wash Buffer: .15M LiCl, 10 mM Tris pH 7.5, 1 mM
EDTA .1% SDS.
RT1 Buffer: .25M Tris pH 8.8 (8.2 at 42°), .25M
30 KCl, 30 mM MgCl₂.
RT2 Buffer: .1M Tris pH 7.5, 25 mM MgCl₂, .5M
KCl, .25 mg/ml BSA, 50 mM
dithiothreitol (DTT).
10X Low Salt: 60 mM Tris pH 7.5, 60 mM MgCl₂, 50 mM
NaCl, 2.5 mg/ml BSA 70 mM DME
35 10X Ligation Additions: 1 mM ATP, 20 mM DTT, 1 mg/ml BSA 10
mM spermidine.
10X Kinasing Buffer: .5M Tris pH 7.5, 10 mM ATP, 20 mM
DTT, 10 mM spermidine, 1 mg/ml BSA

100 mM MgCl₂9i. Transfection of cDNA encoding Ligands in Cos 7 Cells

5 Cos 7 cells are split 1:5 into 100 mm plates in
Dulbecco's modified Eagles medium (DME)/10% fetal calf serum
(FCS), and allowed to grow overnight. 3 ml Tris/DME (0.039M
Tris, pH 7.4 in DME) containing 400 µg/ml DEAE-dextran
(Sigma, D-9885) is prepared for each 100 mm plate of Cos 7
10 cells to be transfected. 10 µg of plasmid DNA preparation
per plate is added. The medium is removed from the Cos-7
cells and the DNA/DEAE-dextran mixture is added. The cells
are incubated for 4.5 hours. The medium is removed from the
cells, and replaced with 3 ml of DME containing 2% fetal calf
15 serum (FCS) and 0.1 mM chloroquine. The cells are incubated
for one hour. After removing the chloroquine and replacing
with 1.5 ml 20% glycerol in PBS, the cells are allowed to
stand at room temperature for one minute. 3 ml Tris/DME is
added, and the mixture is aspirated and washed two times with
20 Tris/DME. 10 ml DME/10% FCS is added and the mixture is
incubated overnight. The transfected Cos 7 cells are split
1:2 into fresh 100 mm plates with (DME)/10% FCS and allowed
to grow.

25 9j. Panning Procedure for Cos 7 cells Expressing Ligand1) Antibody-coated plates:

Bacteriological 100 mm plates are coated for 1.5 hours
30 with rabbit anti-human placental alkaline phosphatase (Dako,
California) diluted 1:500 in 10 ml of 50 mM Tris.HCl, pH 9.5.
The plates are washed three times with 0.15M NaCl, and
incubated with 3 mg BSA/ml PBS overnight. The blocking
solution is aspirated, and the plates are utilized
35 immediately or frozen for later use.

2) Panning cells:

The medium from transfected Cos 7 cells is aspirated,

and 3 ml PBS/0.5 mM EDTA/0.02% sodium azide is added. The plates are incubated at 37°C for thirty minutes in order to detach the cells. The cells are triturated vigorously with a pasteur pipet and collected in a 15 ml centrifuge tube. The plate is washed with a further 2 ml PBS/EDTA/azide solution, which is then added to the centrifuge tube. After centrifuging at 200 xg for five minutes, the cells are resuspended in 3 ml of Aptaqlk-1 (F-1AP21-4) or flk-2 (F-2AP26-0) supernatant from transfected NIH/3T3 cells (see Example 7.), and incubated for 1.5 hours on ice. The cells are centrifuged again at 200 xg for five minutes. The supernatant is aspirated, and the cells are resuspended in 3 ml PBS/EDTA/azide solution. The cell suspension is layered carefully on 3 ml PBS/EDTA/azide/2% Ficoll, and centrifuged at 200 xg for four minutes. The supernatant is aspirated, and the cells are resuspended in 0.5 ml PBS/EDTA/azide solution. The cells are added to the antibody-coated plates containing 4 ml PBS/EDTA/azide/5% FBS, and allowed to stand at room temperature one to three hours. Non-adhering cells are removed by washing gently two or three times with 3 ml PBS/5% FBS.

3) Hirt Supernatant:

0.4 ml 0.6% SDS and 10 mM EDTA are added to the panned plates, which are allowed to stand 20 minutes. The viscous mixture is added by means of a pipet into a microfuge tube. 0.1 ml 5M NaCl is added to the tube, mixed, and chilled on ice for at least five hours. The tube is spun for four minutes, and the supernatant is removed carefully. The contents of the tube are extracted with phenol once, or, if the first interface is not clean, twice. Ten micrograms of linear polyacrylamide (or other carrier) is added, and the tube is filled to the top with ethanol. The resulting precipitate is resuspended in 0.1 ml water or TE. After adding 3 volumes of EtOH/NaOAc, the cells are reprecipitated and resuspended in 0.1 ml water or TE. The cDNA obtained is transfected into any suitable E. coli host by electroporation. Suitable hosts are described in various

catalogs, and include MC1061/p3 or Electromax DH10B Cells of BRL Gibco. The cDNA is extracted by conventional methods.

5 The above panning procedure is repeated until a pure E. coli clone bearing the cDNA as a unique plasmid recombinant capable of transfecting mammalian cells and yielding a positive panning assay is isolated. Normally, three repetitions are sufficient.

10 9k. Expression cloning of flk1 or flk2 ligand by establishment of an autocrine loop

15 Cells expressing flk1/fms or flk2/fms (Example 10) are transfected with 20-30 μ g of a cDNA library from either flk1 ligand or flk2 ligand expressing stromal cells, respectively. The cDNA library is prepared as described above (a-h). The cells are co-transfected with 1 μ g pLTR neo cDNA. Following transfection the cells are passaged 1:2 and cultured in 800 μ g/ml of G418 in Dulbecco's medium (DME) supplemented with 10% CS. Approximately 12 days later the colonies of cells are passaged and plated onto dishes coated with poly-D-lysine (1 mg/ml) and human fibronectin (15 μ g/ml). The culture medium is defined serum-free medium which is a mixture (3:1) of DME and Ham's F12 medium. The medium supplements are 8 mM NaHCO_3 , 15 mM HEPES pH 7.4, 3 mM histidine, 4 μ M MnCl_2 , 10 μ M ethanolamine, 0.1 μ M selenous acid, 2 μ M hydrocortisone, 5 μ g/ml transferrin, 500 μ g/ml bovine serum albumin/linoleic acid complex, and 20 μ g/ml insulin (Ref. Zhan, X, et al. Oncogene 1: 369-376, 1987). The cultures are refed the next day and every 3 days until the only cells capable of growing under the defined medium condition remain. The remaining colonies of cells are expanded and tested for the presence of the ligand by assaying for binding of APTag - flk1 or APTag - flk2 to the cells (as described in Example 8). The DNA would be rescued from cells demonstrating the presence of the flk1 or flk2 ligand and the sequence.

Example 10. Expression of Ligand cDNA

5 The cDNA is sequenced, and expressed in a suitable host cell, such as a mammalian cell, preferably COS, CHO or NIH/3T3 cells. The presence of the ligand is confirmed by demonstrating binding of the ligand to APTag-flk2 fusion protein (see above).

Example 11. Chemical Cross Linking of Receptor and Ligand

10 Cross linking experiments are performed on intact cells using a modification of the procedure described by Blume-Jensen et al et al., EMBO J., 10, 4121-4128 (1991). Cells are cultured in 100mm tissue culture plates to subconfluence and washed once with PBS-0.1% BSA.

20 To examine chemical cross linking of soluble receptor to membrane-bound ligand, stromal cells from the 2018 stromal cell line are incubated with conditioned media (CM) from transfected 3T3 cells expressing the soluble receptor Flk2-APTAg. Cross linking studies of soluble ligand to membrane bound receptor are performed by incubating conditioned media from 2018 cells with transfected 3T3 cells expressing a Flk2-fms fusion construct.

25 Binding is carried out for 2 hours either at room temperature with CM containing 0.02% sodium azide to prevent receptor internalization or at 4°C with CM (and buffers) supplemented with sodium vanadate to prevent receptor dephosphorylation. Cells are washed twice with PBS-0.1% BSA and four times with PBS.

35 Cross linking is performed in PBS containing 250 mM disuccinimidyl suberate (DSS; Pierce) for 30 minutes at room temperature. The reaction is quenched with Tris-HCL pH7.4 to a final concentration of 50 mM.

Cells are solubilized in solubilization buffer: 0.5% Triton - X100, 0.5% deoxycholic acid, 20 mM Tris pH 7.4, 150

mM NaCl, 10mM EDTA, 1mM PMFS, 50 mg/ml aprotinin, 2 mg/ml bestatin, 2 mg/ml pepstatin and 10mg/ml leupeptin. Lysed cells are immediately transferred to 1.5 ml Nalgene tubes and solubilized by rolling end to end for 45 minutes at 4°C. Lysates are then centrifuged in a microfuge at 14,000g for 10 minutes. Solubilized cross linked receptor complexes are then retrieved from lysates by incubating supernatants with 10% (v/v) wheat germ lectin-Sepharose 6MB beads (Pharmacia) at 4°C for 2 hours or overnight.

Beads are washed once with Tris-buffered saline (TBS) and resuspended in 2X SDS-polyacrylamide nonreducing sample buffer. Bound complexes are eluted from the beads by heating at 95°C for 5 minutes. Samples are analyzed on 4-12% gradient gels (NOVEX) under nonreducing and reducing conditions (0.35 M 2-mercaptoethanol) and then transferred to PVDF membranes for 2 hours using a Novex blotting apparatus. Blots are blocked in TBS-3% BSA for 1 hour at room temperature followed by incubation with appropriate antibody.

Cross linked Flk2-Aptag and Flk2-fms receptors are detected using rabbit polyclonal antibodies raised against human alkaline phosphatase and fms protein, respectively. The remainder of the procedure is carried out according to the instructions provided in the ABC Kit (Pierce). The kit is based on the use of a biotinylated secondary antibody and avidin-biotinylated horseradish peroxidase complex for detection.

Example 12. Expression and purification of Flag-Flk-2.

1. Design of the Flag-Flk2 expression plasmids.

A synthetic DNA fragment (Fragment 1) is synthesized using complementary oligonucleotides BP1 and BP2 (see below and SEQ. ID. NOS. 7 and 8). The fragment encoded the following features in the 5' to 3' order: Sal I restriction

site, 22 base pair (bp) 5' untranslated region containing an eukaryotic ribosome binding site, an ATG initiation codon, preprotrypsinogen signal sequence, coding region for the FLAG peptide (DYKDDDDKI) and Bgl II restriction site.

5. A cDNA fragment (Fragment 2) encoding Asn 27 to Ser 544 of murine Flk2 is obtained by polymerase chain reaction (PCR) using primers designed to introduce an in frame Bgl II site at the 5' end (oligonucleotide BP5, see below and SEQ. ID. NO. 9) and a termination codon followed by a Not I site at 10 the 3' end (oligonucleotide BP10, see below and SEQ. ID. NO. 10). The template for the PCR reaction is full length Flk2 cDNA (Matthews et al., Cell 65:1143 (1991)). Fragment 2 is 15 extensively digested with Bgl II and Not I restriction enzymes prior to ligation.

To assemble the complete Flag-Flk2 gene, Fragments 1 and 2 are ligated in a tripartate ligation into Sal I and Not I digested plasmid pSPORT (Gibco/BRL, Grand Island, NY) to give 20 the plasmid pFlag-Flk2.

Preferably, the Flag-Flk2 protein is attached at either end to the Fc portion of an immunoglobulin (Ig). The Ig is preferably attached to the Flk2 portion of the Flag-Flk2 25 protein. To assemble the construct pFlag-FLK2-Ig, the sequences coding for the CH¹ domain of human immunoglobulin G (IgG¹) are placed downstream of the Flk2 coding region in the plasmid pFlag-Flk2 as per the method described by Zettlemessl et al., DNA and Cell Biology 2: 347-352 (1990).

30 The sequences of oligonucleotides used to construct the Flag-Flk2 gene are given below:

Oligonucleotide BP1:

35 5'-AATTCGTCGACTTTCTGTCACCATGAGTGCACTTCTGATCCTAGCCCTTGTG
GGAGCTGCTGTTGCTGACTACAAAGATGATGATGACAAGATCTA-3'

Oligonucleotide BP2:

5'-AGCTTAGATCTTGTGTCATCATCATCTTTGTAGTCAGCAACAGCAGCTCCCA

AGGGCTAGGATCAGAAGTGCACTCATGGTGACAGAAAGTCGACG-3'

Oligonucleotide BP5:

5'-TGAGAAGATCTCAAACCAAGACCTGCCTGT-3'

5

Oligonucleotide BP10:

5'-CCAATGGCGGCCGCTCAGGAGATGTTGTCTTGGA-3'

2. Expression of the Flag-Flk2 construct.

10

For transient expression of the Flag-Flk2 construct, the SalI to Not I fragment from pFlag-Flk2 is subcloned into the plasmid pSVSPORT (Gibco/BRL) to give the plasmid pSVFlag-Flk2. For expression of the Flag-Flk2 protein pSVFlag-Flk2 is transfected into COS monkey cells using the DEAE-dextran method.

15

For stable expression in eukaryotic cells, the Sal I-Not I fragment of pFlag-Flk2 is cloned into the EcoRV and Not I sites of the plasmid pCDNA I/Neo (Invitrogen Co., San Diego, CA). The Sal I 3' recessed terminus of pFlag-Flk2 is filled with the Klenow fragment of DNA polymerase I and a mixture of deoxyribonucleotides to make the site compatible with the EcoRV site of the vector. The resulting construct is introduced into cultured mamalian cells using either the Lipofectin (Gibco/BRL) or the calcium phosphate methods.

20

25

For expression in insect cells, the SalI to Hind III (from pSPORT polylinker) fragment of pFlag-Flk2 is subcloned into the BamHI-Hind III sites of the baculovirus transfer vector pBlueBac III (Invitrogen). The vector Bam HI site and the insert Sal I site are blunted with Klenow (see above). Production of the recombinant virus and infection of the Sf9 insect cells is performed as per manufacturers directions (Invitrogen).

30

35

Expression of the Flag-Flk2 protein is detected by Western blotting of SDS-PAGE separated conditioned media (mamalian cells) or cell lysates (insect cells) with the

anti-Flag monoclonal antibody (mAb) M1 (International Biotechnology, Inc. [IBI], New Haven, CT).

5 3. Affinity purification of the Flag-Flk2 protein from conditioned media or insect cell lysates is performed using immobilized mAb M1 (IBI) as per manufacturers specifications.

10 3.1 Affinity purification of the Flag-Flk2-Ig¹ protein from conditioned media is performed using immobilized Protein A (Pharmacia LKB, Piscataway, NJ) as per the manufacturers instructions.

15 II. Use of the Flag-Flk2 protein to search for the Flk2 ligand.

1. Binding and cross-linking studies to detect membrane-bound ligand:

A. Binding studies.

20 Murine stromal lines (eg. 2018 cells ATCC CRL 10907 (see below), see example 1, supra) considered to be candidates for expression of the Flk2 ligand were deposited at the American Type Culture Collection, ATCC CRL 10907 (see
25 below) and cultured in Dulbecco's modified Eagles medium (DMEM; Gibco/BRL) supplemented with 10% fetal calf serum. The cells are grown to confluency in 10 cm plates and washed once with PBS. Conditioned media containing Flag-Flk2 is incubated with the cells at 4°C for 2 hrs. The cell monolayers are
30 rinsed extensively to remove the non-bound protein, solubilized and centrifuged to remove insoluble cellular material. Glycoproteins in the lysates are partially purified with wheat germ agglutinin-Sepharose (Pharmacia LKB, Piscataway, NJ), boiled in an SDS sample buffer, separated on
35 SDS-PAGE gels and transferred to nitrocellulose membranes. The membranes are probed with the M1 antibody to detect the presence of cell-associated Flag-Flk2 protein.

B. In a cross-linking study, the above protocol is

followed except that prior to solubilization the monolayer are treated with the crosslinker disuccinimidyl suberate (DSS; Pierce, Rockford, IL). The presence of a putative ligand is detected by an upward shift in the apparent molecular weight of the Flag-Flk2 band on Western blots.

C. Purified Flag-Flk2 protein labelled with Na¹²⁵I via the Chloramine T method is used to assess the ability of the soluble extracellular domain of the Flk2 receptor to bind transmembrane form of the Flk2 ligand in cultured stromal lines. The labelled protein is added to monolayers of stromal cells on ice for 2 hr in the presence or absence of excess unlabelled protein. Specific binding is calculated by subtracting counts bound in the presence of excess unlabelled protein from the total counts bound.

2. Use of the Flag-Flk2 protein to search for secreted form of the ligand.

A. The Flag-Flk2 protein is used in attempts to identify the Flk2 ligand in conditioned media from stromal cell cultures via modification of the direct N-terminal sequencing method of Pan et al., Bioch. Biophys. Res. Comm. 166:201 (1990). Briefly, the Flag-Flk2 protein N-terminally sequenced by automatic Edman degradation chemistry on an ABI 477A sequencer with on line PTH amino acid analysis. Approximately 15 amino acids are determined. The protein is then immobilized on Nugal PAF silica beads via free NH₄⁺ groups. The immobilized Flag-Flk2 is incubated with conditioned media from putative ligand-producing cells for 30 min at 4°C and washed free off non-bound proteins with phosphate buffered saline adjusted to 2M NaCl. The resulting protein complex is resequenced. For each sequencing cycle, any amino acid not expected at this position in the FLAG-Flk2 protein is considered as possibly originating from a protein complexed to the Flk2 receptor.

B. For conventional affinity chromatography, the Flag-Flk2 protein is immobilized on a stable support such as

Sepharose. 35S-methionine labelled-conditioned media from stromal cell lines are passed over the affinity matrix and bound material is analyzed by SDS-PAGE gel electrophoresis and autoradiography.

5

3. Use of the Flag-Flk2 protein in expression cloning experiments.

10 A method of expression cloning of integral membrane proteins in COS cells has been described (Aruffo and Seed, Proc. Natl. Acad. Sci. 84:8573 (1987)). A cDNA library is prepared from an appropriate stromal cell line such as 2018 and is transfected into COS cells. Cells transiently expressing the Flk2 ligand are affinity adsorbed onto plastic plates coated with the Flag-Flk2 protein. The cells are
15 lysed, the plasmid DNA is recovered and amplified in a bacterial host. The cycle of transfection into COS cells is repeated until a single cDNA clone encoding the ligand molecule is isolated.

20

In a modification of the above technique, pools of transfected COS cells are screened for binding of 125I-Flag-Flk2. Positive cells pools are selected and plasmid DNA is recovered and amplified in E. coli. The resulting DNA
25 preparation is used in subsequent rounds of transfection and transient expression until all cells are positive for binding of 125I-Flag-Flk2. The cDNA in the final plasmid preparation is then sequenced to determine the sequence of the putative Flk-2 ligand.

30

SUPPLEMENTAL ENABLEMENT

The invention as claimed is enabled in accordance with the above specification and readily available references and
35 starting materials. Nevertheless, Applicants have deposited with the American Type Culture Collection, Rockville, Md., USA (ATCC) the cell lines listed below:

2018, ATCC accession no. CRL 10907, deposited

October 30, 1991.

Fsp 62891, ATCC accession no. CRL 10935, deposit d
November 21, 1991.

5

F.thy 62891, ATCC accession no. CRL 10936,
deposited November 21, 1991.

10

FL 62891, ATCC accession no. CRL 11005, deposited
April 2, 1992.

These deposits were made under the provisions of the
Budapest Treaty on the International Recognition of the
Deposit of Microorganisms for the Purposes of Patent
15 Procedure and the regulations thereunder (Budapest Treaty).
This assures maintenance of a viable culture for 30 years
from date of deposit. The organisms will be made available
by ATCC under the terms of the Budapest Treaty, and subject
to an agreement between Applicants and ATCC which assures
20 unrestricted availability upon issuance of the pertinent U.S.
patent. Availability of the deposited strains is not to be
construed as a license to practice the invention in
contravention of the rights granted under the authority of
any government in accordance with its patent laws.

25

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT: Lemischka, Ihor R.
- (ii) TITLE OF INVENTION: TOTIPOTENT HEMATOPOIETIC STEM CELL RECEPTORS AND THEIR LIGANDS
- (iii) NUMBER OF SEQUENCES: 10
- (iv) CORRESPONDENCE ADDRESS:
- (A) ADDRESSEE: ImClone Systems Incorporated
 - (B) STREET: 180 Varick Street
 - (C) CITY: New York
 - (D) STATE: New York
 - (E) COUNTRY: U.S.A.
 - (F) ZIP: 10014
- (v) COMPUTER READABLE FORM:
- (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
- (vi) CURRENT APPLICATION DATA:
- (A) APPLICATION NUMBER:
 - (B) FILING DATE:
 - (C) CLASSIFICATION:
- (vii) ATTORNEY/AGENT INFORMATION:
- (A) NAME: Feit, Irving N.
 - (B) REGISTRATION NUMBER: 28,601
 - (C) REFERENCE/DOCKET NUMBER: LEM-3-7PT
- (ix) TELECOMMUNICATION INFORMATION:
- (A) TELEPHONE: 212-645-1405

(B) TELEFAX: 212-645-2054

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 3453 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(v) FRAGMENT TYPE: N-terminal

(ix) FEATURE:

- (A) NAME/KEY: mat_peptide
- (B) LOCATION: 112..3006

(ix) FEATURE:

- (A) NAME/KEY: sig_peptide
- (B) LOCATION: 31..111

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 31..3009

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

CGCGCCTGGC TACCGCGCGC TCCGGAGGCC ATG CGG GCG TTG GCG CAG CGC AGC 54
Met Arg Ala Leu Ala Gln Arg Ser
-27 -25 -20

102 GAC CGG CGG CTG CTG CTT GTT GTT TTG TCA GTA ATG ATT CTT GAG
 Asp Arg Arg Leu Leu Leu Val Leu Ser Val Met Ile Leu Glu
 -15 -10 -5
 150 ACC GTT ACA AAC CAA GAC CTG CCT GTG ATC AAG TGT GTT TTA ATC AGT
 Thr Val Thr Asn Gln Asp Leu Pro Val Ile Lys Cys Val Leu Ile Ser
 1 5 10
 198 CAT GAG AAC AAT GGC TCA TCA GCG GGA AAG CCA TCA TCG TAC CGA ATG
 His Glu Asn Asn Gly Ser Ser Ala Gly Lys Pro Ser Ser Tyr Arg Met
 15 20 25
 246 GTG CGA GGA TCC CCA GAA GAC CTC CAG TGT ACC CCG AGG CGC CAG AGT
 Val Arg Gly Ser Pro Glu Asp Leu Gln Cys Thr Pro Arg Arg Gln Ser
 30 35 40 45
 294 GAA GGG ACG GTA TAT GAA GCG GCC ACC GTG GAG GTG GCC GAG TCT GGG
 Glu Gly Thr Val Tyr Glu Ala Ala Thr Val Glu Val Ala Glu Ser Gly
 50 55 60
 342 TCC ATC ACC CTG CAA GTG CAG CTC GCC ACC CCA GGG GAC CTT TCC TGC
 Ser Ile Thr Leu Gln Val Gln Leu Ala Thr Pro Gly Asp Leu Ser Cys
 65 70 75
 390 CTC TGG GTC TTT AAG CAC AGC TCC CTG GGC TGC CAG CCG CAC TTT GAT
 Leu Trp Val Phe Lys His Ser Ser Leu Gly Cys Gln Pro His Phe Asp
 80 85 90
 438 TTA CAA AAC AGA GGA ATC GTT TCC ATG GCC ATC TTG AAC GTG ACA GAG
 Leu Gln Asn Arg Gly Ile Val Ser Met Ala Ile Leu Asn Val Thr Glu
 95 100 105
 486 ACC CAG GCA GGA GAA TAC CTC CAT ATT CAG AGC GAA CGC GCC AAC
 Thr Gln Ala Gly Glu Tyr Leu Leu His Ile Gln Ser Glu Arg Ala Asn
 110 115 120 125
 534 TAC ACA GTA CTG TTC ACA GTG AAT GTA AGA GAT ACA CAG CTG TAT GTG
 Tyr Thr Val Leu Phe Thr Val Asn Val Arg Asp Thr Gln Leu Tyr Val

CTA AGG AGA CCT TAC TTT AGG AAG ATG GAA AAC CAG GAT GCA CTG CTC Leu Arg Arg Pro Tyr Phe Arg Lys Met Glu Asn Gln Asp Ala Leu Leu 145 150 155	130	135	140	582
TGC ATC TCC GAG GGT GTT CCG GAG CCC ACT GTG GAG TGG GTG CTC TGC Cys Ile Ser Glu Gly Val Pro Glu Pro Thr Val Glu Trp Val Leu Cys 160 165 170				630
AGC TCC CAC AGG GAA AGC TGT AAA GAA GAA GGC CCT GCT GTT GTC AGA Ser Ser His Arg Glu Ser Cys Lys Glu Glu Gly Pro Ala Val Val Arg 175 180 185				678
AAG GAG GAA AAG GTA CTT CAT GAG TTG TTC GGA ACA GAC ATC AGA TGC Lys Glu Glu Lys Val Leu His Glu Leu Phe Gly Thr Asp Ile Arg Cys 190 195 200 205				726
TGT GCT AGA AAT GCA CTG GGC CGC GAA TGC ACC AAG CTG TTC ACC ATA Cys Ala Arg Asn Ala Leu Gly Arg Glu Cys Thr Lys Leu Phe Thr Ile 210 215 220				774
GAT CTA AAC CAG GCT CCT CAG AGC ACA CTG CCC CAG TTA TTC CTG AAA Asp Leu Asn Gln Ala Pro Gln Ser Thr Leu Pro Gln Leu Phe Leu Lys 225 230 235				822
GTG GGG GAA CCC TTG TGG ATC AGG TGT AAG GCC ATC CAT GTG AAC CAT Val Gly Glu Pro Leu Trp Ile Arg Cys Lys Ala Ile His Val Asn His 240 245 250				870
GGA TTC GGG CTC ACC TGG GAG CTG GAA GAC AAA GCC CTG GAG GAG GGC Gly Phe Gly Leu Thr Trp Glu Leu Glu Asp Lys Ala Leu Glu Glu Gly 255 260 265				918
AGC TAC TTT GAG ATG AGT ACC TAC TCC ACA AAC AGG ACC ATG ATT CGG Ser Tyr Phe Glu Met Ser Thr Tyr Ser Thr Asn Arg Thr Met Ile Arg 270 275 280 285				966

1014
 ATT CTC TTG GCC TTT GTG TCT TCC GTG GGA AGG AAC GAC ACC GGA TAT
 Ile Leu Leu Ala Phe Val Ser Ser Val Gly Arg Asn Asp Thr Gly Tyr
 290 295 300

1062
 TAC ACC TGC TCT TCC TCA AAG CAC CCC AGC CAG TCA GCG TTG GTG ACC
 Tyr Thr Cys Ser Ser Ser Lys His Pro Ser Gln Ser Ala Leu Val Thr
 305 310 315

1110
 ATC CTA GAA AAA GGG TTT ATA AAC GCT ACC AGC TCG CAA GAA GAG TAT
 Ile Leu Glu Lys Gly Phe Ile Asn Ala Thr Ser Ser Gln Glu Glu Tyr
 320 325 330

1158
 GAA ATT GAC CCG TAC GAA AAG TTC TGC TTC TCA GTC AGG TTT AAA GCG
 Glu Ile Asp Pro Tyr Glu Lys Phe Cys Phe Ser Val Arg Phe Lys Ala
 335 340 345

1206
 TAC CCA CGA ATC CGA TGC AGC TGG ATC TTC TCT CAA GCC TCA TTT CCT
 Tyr Pro Arg Ile Arg Cys Thr Trp Ile Phe Ser Ser Gln Ala Ser Phe Pro
 350 355 360 365

1254
 TGT GAA CAG AGA GGC CTG GAG GAT GGG TAC AGC ATA TCT AAA TTT TGC
 Cys Glu Gln Arg Gly Leu Glu Asp Gly Tyr Ser Ile Ser Lys Phe Cys
 370 375 380

1302
 GAT CAT AAG AAC AAG CCA GGA GAG TAC ATA TTC TAT GCA GAA AAT GAT
 Asp His Lys Asn Lys Pro Gly Glu Tyr Ile Phe Tyr Ala Glu Asn Asp
 385 390 395

1350
 GAC GCC CAG TTC ACC AAA ATG TTC ACG CTG AAT ATA AGA AAG AAA CCT
 Asp Ala Gln Phe Thr Lys Met Phe Thr Leu Asn Ile Arg Lys Lys Pro
 400 405 410

1398
 CAA GTG CTA GCA AAT GCC TCA GCC AGC CAG GCG TCC TGT TCC TCT GAT
 Gln Val Leu Ala Asn Ala Ser Ala Ser Gln Ala Ser Cys Ser Ser Asp
 415 420 425

1446
 GGC TAC CCG CTA CCC TCT TGG ACC TGG AAG AAG TGT TCG GAC AAA TCT
 Gly Tyr Pro Leu Pro Ser Trp Thr Trp Lys Lys Cys Ser Asp Lys Ser

430	435	440	445	
CCC AAT TGC ACG GAG GAA ATC CCA GAA GGA GTT TGG AAT AAA AAG GCT				1494
Pro Asn Cys Thr Glu Glu Thr Glu Glu Ile Pro Glu Gly Val Trp Asn Lys Lys Ala	450	455	460	
AAC AGA AAA GTG TTT GGC CAG TGG GTG TCG AGC AGT ACT CTA AAT ATG				1542
Asn Arg Lys Val Phe Gly Gln Trp Val Ser Ser Thr Leu Asn Met	465	470	475	
AGT GAG GCC GGG AAA GGG CTT CTG GTC AAA TGC TGT GCG TAC AAT TCT				1590
Ser Glu Ala Gly Lys Glu Leu Leu Val Lys Cys Ala Tyr Asn Ser	480	485	490	
ATG GGC ACG TCT TGC GAA ACC ATC TTT TTA AAC TCA CCA GGC CCC TTC				1638
Met Gly Thr Ser Cys Glu Thr Ile Phe Leu Asn Ser Pro Gly Pro Phe	495	500		
CCT TTC ATC CAA GAC AAC ATC TCC TTC TAT GCG ACC ATT GGG CTC TGT				1686
Pro Phe Ile Gln Asp Asn Ile Ser Phe Tyr Ala Thr Ile Gly Leu Cys	510	515	520	
CTC CCC TTC ATT GTT CTC ATT GTG TTT GTG TGC CAC AAA TAC AAA				1734
Leu Pro Phe Ile Val Val Leu Ile Val Leu Ile Cys His Lys Tyr Lys	530	535	540	
AAG CAA TTT AGG TAC GAG AGT CAG CTG CAG ATG ATC CAG GTG ACT GGC				1782
Lys Gln Phe Arg Tyr Glu Ser Gln Leu Leu Met Ile Gln Val Thr Gly	545	550	555	
CCC CTG GAT AAC GAG TAC TTC TAC GTT GAC TTC AGG GAC TAT GAA TAT				1830
Pro Leu Asp Asn Glu Tyr Phe Tyr Val Asp Phe Arg Asp Tyr Glu Tyr	560	565	570	
GAC CTT AAG TGG GAG TTC CCG AGA GAG AAC TTA GAG TTT GGG AAG GTC				1878
Asp Leu Lys Trp Glu Phe Pro Arg Glu Asn Leu Phe Gly Lys Val	575	580	585	

1926 CTG GGG TCT GGC GCT TTC GGG AGG GTG ATG AAC GCC ACG GCC TAT GGC
 Leu Gly Ser Gly Ala Phe Gly Arg Val Met Asn Ala Thr Ala Tyr Gly
 590 595 600 605
 1974 ATT AGT AAA ACG GGA GTC TCA ATT CAG GTG GCG GTG AAG ATG CTA AAA
 Ile Ser Lys Thr Gly Val Ser Ile Gln Val Ala Val Lys Met Leu Lys
 610 615 620
 2022 GAG AAA GCT GAC AGC TGT GAA AAA GAA GCT CTC ATG TCG GAG CTC AAA
 Glu Lys Ala Asp Ser Cys Glu Lys Glu Ala Leu Met Ser Glu Leu Lys
 625 630 635
 2070 ATG ATG ACC CAC CTG GGA CAC CAT GAC AAC ATC GTG AAT CTG CTG GGG
 Met Met Thr His Leu Gly His His Asp Asn Ile Val Asn Leu Leu Gly
 640 645 650
 2118 GCA TGC ACA CTG TCA GGG CCA GTG TAC TTG ATT TTT GAA TAT TGT TGC
 Ala Cys Thr Leu Ser Gly Pro Val Tyr Leu Ile Phe Glu Tyr Cys Cys
 655 660 665
 2166 TAT GGT GAC CTC AAC TAC CTA AGA AGT AAA AGA GAG AAG TTT CAC
 Tyr Gly Asp Leu Leu Asn Tyr Leu Arg Ser Lys Arg Glu Lys Phe His
 670 675 680 685
 2214 AGG ACA TGG ACA GAG ATT TTT AAG GAA CAT AAT TTC AGT TCT TAC CCT
 Arg Thr Trp Thr Glu Ile Phe Lys Glu His Asn Phe Ser Ser Tyr Pro
 690 695 700
 2262 ACT TTC CAG GCA CAT TCA AAT TCC AGC ATG CCT GGT TCA CGA GAA GTT
 Thr Phe Gln Ala His Ser Asn Ser Ser Met Pro Gly Ser Arg Glu Val
 705 710 715
 2310 CAG TTA CAC CCG CCC TTG GAT CAG CTC TCA GGG TTC AAT GGG AAT TCA
 Gln Leu His Pro Pro Leu Asp Gln Leu Ser Gly Phe Asn Gly Asn Ser
 720 725 730
 2358 ATT CAT TCT GAA GAT GAG ATT GAA TAT GAA AAC CAG AAG AGG CTG GCA
 Ile His Ser Glu Asp Glu Ile Glu Tyr Glu Asn Gln Lys Arg Leu Ala

735	740	745	
GAA GAA GAG GAG GAA GAT TTG AAC GTG CTG ACG TTT GAA GAC CTC CTT			2406
Glu Glu Glu Glu Asp 755	Leu Thr Phe Glu Asp Leu 760		
750			
TGC TTT GCG TAC CAA GTG GCC AAA GGC ATG GAA TTC CTG GAG TTC AAG			2454
Cys Phe Ala Tyr Gln Val 770	Met Glu Phe Leu Glu Phe Lys 780		
775			
TCG TGT GTC CAC AGA GAC CTG GCA GCC AGG AAT GTG TTG GTC ACC CAC			2502
Ser Cys Val His Arg Asp Leu Ala 785	Asn Val Leu Val Thr His 795		
790			
GGG AAG GTG AAG ATC TGT GAC TTT GGA CTG GCC CGA GAC ATC CTG			2550
Gly Lys Val Val Lys 800	Phe Gly Leu Ala Arg Asp Ile Leu 810		
805			
AGC GAC TCC AGC TAC GTC GTC AGG GGC AAC GCA CCG CTG CCG GTG AAG			2598
Ser Asp Ser Ser Tyr Val 815	Arg Gly Asn Ala Arg Leu Pro Val Lys 825		
820			
TGG ATG GCA CCC GAG AGC TTA TTT GAA GGG ATC TAC ACA ATC AAG AGT			2646
Trp Met Ala Pro Glu Ser Leu 835	Ile Tyr Thr Ile Lys Ser 845		
830			
GAC GTC TGG TCC TAC GGC ATC CTT CTC TGG GAG ATA TTT TCA CTG GGT			2694
Asp Val Trp Ser Tyr Gly Ile Leu 850	Trp Glu Ile Phe Ser Leu Gly 860		
855			
GTG AAC CCT TAC CCT GGC ATT CCT GTC GAC GCT AAC TTC TAT AAA CTG			2742
Val Asn Pro Tyr Pro Gly Ile Pro 865	Val Asp Ala Asn Phe Tyr Lys Leu 875		
870			
ATT CAG AGT GGA TTT AAA ATG GAG CAG CCA TTC TAT GCC ACA GAA GGG			2790
Ile Gln Ser Gly Phe Lys Met 880	Gln Pro Phe Tyr Ala Thr Glu Gly 890		
885			

2838 ATA TAC TTT GTA ATG CAA TCC TGC TGG GCT TTT GAC TCA AGG AAG CGG
 Ile Tyr Phe Val Met Gln Ser Cys Trp Ala Phe Asp Ser Arg Lys Arg
 895 900 905
 2886 CCA TCC TTC CCC AAC CTG ACT TCA TTT TTA GGA TGT CAG CTG GCA GAG
 Pro Ser Phe Pro Asn 915 Leu Thr Ser Phe Leu Gly Cys Gln Leu Ala Glu
 910 920 925
 2934 GCA GAA GAA GCA TGT ATC AGA ACA TCC ATC CAT CTA CCA AAA CAG GCG
 Ala Glu Glu Ala Cys Ile Arg Thr Ser Ile His Leu Pro Lys Gln Ala
 930 935 940
 2982 GCC CCT CAG CAG AGA GGC GGC CTC AGA GCC CAG TCG CCA CAG CGC CAG
 Ala Pro Gln Gln Arg Gly Gly Leu Arg Ala Gln Ser Pro Gln Arg Gln
 945 950 955
 3036 GTG AAG ATT CAC AGA GAA AGA AGT TAGCGAGGAG GCCTTGGACC CCGCCACCCT
 Val Lys Ile His Arg Glu Arg Ser
 960
 3096 AGCAGGCTGT AGACCGCAGA GCCAAGATTA GCCTCGCCTC TGAGGAAGCG CCCTACAGCG
 3156 CGTTGCTTCG CTGGACTTTT CTCTAGATGC TGTCTGCCAT TACTCCAAAG TGACTTCTAT
 3216 AAAATCAAAC CTCTCCTCGC ACAGGCGGGA GAGCCAAATAA TGAGACTTGT TGGTGAGCCC
 3276 GCCTACCCCTG GGGGCCCTTC CACGAGCTTG AGGGGAAAGC CATGTATCTG AAATATAGTA
 3336 TATTCTTGTA AATACGTGAA ACAAAACCAA CCGTTT TTTT GCTAAGGGAA AGCTAAATAT
 3396 GATTTTAAA AATCTATGTT TTAATAACT ATGTAAC TTTT TATCATCTATT TAGTGATATA
 3453 TTTTATGGAT GAAATAAAC TTTCTACTGT AAAAAAAAAA AAAAAAAAAA AAAAAA

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 992 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Arg Ala Leu Ala Gln Arg Ser Asp Arg Arg Leu Leu Leu Val	-15
-27 -25	
Val Leu Ser Val Met Ile Leu Glu Thr Val Thr Asn Gln Asp Leu Pro	5
-10 -5	1
Val Ile Lys Cys Val Leu Ile Ser His Glu Asn Asn Gly Ser Ser Ala	20
10 15	
Gly Lys Pro Ser Ser Tyr Arg Met Val Arg Gly Ser Pro Glu Asp Leu	35
25 30	
Gln Cys Thr Pro Arg Arg Gln Ser Glu Gly Thr Val Tyr Glu Ala Ala	50
40 45	
Thr Val Glu Val Ala Glu Ser Gly Ser Ile Thr Leu Gln Val Gln Leu	65
55 60	
Ala Thr Pro Gly Asp Leu Ser Cys Leu Trp Val Phe Lys His Ser Ser	85
70 75	
Leu Gly Cys Gln Pro His Phe Asp Leu Gln Asn Arg Gly Ile Val Ser	100
90 95	
Met Ala Ile Leu Asn Val Thr Glu Thr Gln Ala Gly Glu Tyr Leu Leu	115
105 110	
His Ile Gln Ser Glu Arg Ala Asn Tyr Thr Val Leu Phe Thr Val Asn	130
120 125	

Val Arg Asp Thr Gln Leu Tyr Val Leu Arg Arg Pro Tyr Phe Arg Lys
 135 140 145
 Met Glu Asn Gln Asp Ala Leu Leu Cys Ile Ser Glu Gly Val Pro Glu
 150 155 160 165
 Pro Thr Val Glu Trp Val Leu Cys Ser Ser His Arg Glu Ser Cys Lys
 170 175 180
 Glu Glu Gly Pro Ala Val Val Arg Lys Glu Glu Lys Val Leu His Glu
 185 190 195
 Leu Phe Gly Thr Asp Ile Arg Cys Cys Ala Arg Asn Ala Leu Gly Arg
 200 205 210
 Glu Cys Thr Lys Leu Phe Thr Ile Asp Leu Asn Gln Ala Pro Gln Ser
 215 220 225
 Thr Leu Pro Gln Leu Phe Leu Lys Val Gly Glu Pro Leu Trp Ile Arg
 230 235 240 245
 Cys Lys Ala Ile His Val Asn His Gly Phe Gly Leu Thr Trp Glu Leu
 250 255 260
 Glu Asp Lys Ala Leu Glu Glu Gly Ser Tyr Phe Glu Met Ser Thr Tyr
 265 270 275
 Ser Thr Asn Arg Thr Met Ile Arg Ile Leu Leu Ala Phe Val Ser Ser
 280 285 290
 Val Gly Arg Asn Asp Thr Gly Tyr Tyr Thr Cys Ser Ser Ser Lys His
 295 300 305
 Pro Ser Gln Ser Ala Leu Val Thr Ile Leu Glu Lys Gly Phe Ile Asn
 310 315 320 325
 Ala Thr Ser Ser Gln Glu Glu Tyr Glu Ile Asp Pro Tyr Glu Lys Phe
 330 335 340

Cys Phe Ser Val Arg Phe Lys Ala Tyr Pro Arg Ile Arg Cys Thr Trp
 345 350 355
 Ile Phe Ser Gln Ala Ser Phe Pro Cys Glu Gln Arg Gly Leu Glu Asp
 360 365 370
 Gly Tyr Ser Ile Ser Lys Phe Cys Asp His Lys Asn Lys Pro Gly Glu
 375 380 385
 Tyr Ile Phe Tyr Ala Glu Asn Asp Asp Ala Gln Phe Thr Lys Met Phe
 390 395 400 405
 Thr Leu Asn Ile Arg Lys Lys Pro Gln Val Leu Ala Asn Ala Ser Ala
 410 415 420
 Ser Gln Ala Ser Cys Ser Ser Asp Gly Tyr Pro Leu Pro Ser Trp Thr
 425 430 435
 Trp Lys Lys Cys Ser Asp Lys Ser Pro Asn Cys Thr Glu Glu Ile Pro
 440 445 450
 Glu Gly Val Trp Asn Lys Lys Ala Asn Arg Lys Val Phe Gly Gln Trp
 455 460 465
 Val Ser Ser Ser Thr Leu Asn Met Ser Glu Ala Gly Lys Gly Leu Leu
 470 475 480 485
 Val Lys Cys Cys Ala Tyr Asn Ser Met Gly Thr Ser Cys Glu Thr Ile
 490 495 500
 Phe Leu Asn Ser Pro Gly Pro Phe Pro Phe Ile Gln Asp Asn Ile Ser
 505 510 515
 Phe Tyr Ala Thr Ile Gly Leu Cys Leu Pro Phe Ile Val Val Leu Ile
 520 525 530
 Val Leu Ile Cys His Lys Tyr Lys Lys Gln Phe Arg Tyr Glu Ser Gln
 535 540 545

Leu Gln Met Ile Gln Val Thr Gly Pro Leu Asp Asn Glu Tyr Phe Tyr 565
 550 555 560
 Val Asp Phe Arg Asp Tyr Glu Tyr Asp Leu Lys Trp Glu Phe Pro Arg 580
 570 575
 Glu Asn Leu Glu Phe Gly Lys Val Leu Gly Ser Gly Ala Phe Gly Arg 595
 585 590
 Val Met Asn Ala Thr Ala Tyr Gly Ile Ser Lys Thr Gly Val Ser Ile 610
 600 605
 Gln Val Ala Val Lys Met Leu Lys Glu Lys Ala Asp Ser Cys Glu Lys 625
 615 620
 Glu Ala Leu Met Ser Glu Leu Lys Met Met Thr His Leu Gly His His 645
 630 635 640
 Asp Asn Ile Val Asn Leu Leu Gly Ala Cys Thr Leu Ser Gly Pro Val 660
 650 655
 Tyr Leu Ile Phe Glu Tyr Cys Cys Tyr Gly Asp Leu Leu Asn Tyr Leu 675
 665 670
 Arg Ser Lys Arg Glu Lys Phe His Arg Thr Trp Thr Glu Ile Phe Lys 690
 680 685
 Glu His Asn Phe Ser Ser Tyr Pro Thr Phe Gln Ala His Ser Asn Ser 705
 695 700
 Ser Met Pro Gly Ser Arg Glu Val Gln Leu His Pro Pro Leu Asp Gln 725
 710 715 720
 Leu Ser Gly Phe Asn Gly Asn Ser Ile His Ser Glu Asp Glu Ile Glu 740
 730 735
 Tyr Glu Asn Gln Lys Arg Leu Ala Glu Glu Glu Glu Asp Leu Asn 755
 745 750

Val Leu Thr Phe Glu Asp Leu Leu Cys Phe Ala Tyr Gln Val Ala Lys
 760 765 770
 Gly Met Glu Phe Leu Glu Phe Lys Ser Cys Val His Arg Asp Leu Ala
 775 780 785
 Ala Arg Asn Val Leu Val Thr His Gly Lys Val Lys Ile Cys Asp
 790 795 800 805
 Phe Gly Leu Ala Arg Asp Ile Leu Ser Asp Ser Ser Tyr Val Val Arg
 810 815 820
 Gly Asn Ala Arg Leu Pro Val Lys Trp Met Ala Pro Glu Ser Leu Phe
 825 830 835
 Glu Gly Ile Tyr Thr Ile Lys Ser Asp Val Trp Ser Tyr Gly Ile Leu
 840 845 850
 Leu Trp Glu Ile Phe Ser Leu Gly Val Asn Pro Tyr Pro Gly Ile Pro
 855 860 865
 Val Asp Ala Asn Phe Tyr Lys Leu Ile Gln Ser Gly Phe Lys Met Glu
 870 875 880 885
 Gln Pro Phe Tyr Ala Thr Glu Gly Ile Tyr Phe Val Met Gln Ser Cys
 890 895 900
 Trp Ala Phe Asp Ser Arg Lys Arg Pro Ser Phe Pro Asn Leu Thr Ser
 905 910 915
 Phe Leu Gly Cys Gln Leu Ala Glu Ala Glu Glu Ala Cys Ile Arg Thr
 920 925 930
 Ser Ile His Leu Pro Lys Gln Ala Ala Pro Gln Gln Arg Gly Gly Leu
 935 940 945
 Arg Ala Gln Ser Pro Gln Arg Gln Val Lys Ile His Arg Glu Arg Ser
 950 955 960 965

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 3501 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(v) FRAGMENT TYPE: N-terminal

(ix) FEATURE:

 (A) NAME/KEY: CDS

 (B) LOCATION: 58..3039

(ix) FEATURE:

 (A) NAME/KEY: mat_peptide

 (B) LOCATION: 139..3036

(ix) FEATURE:

 (A) NAME/KEY: sig_peptide

 (B) LOCATION: 58..138

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

CGAGGCGGCA TCCGAGGGCT GGGCCGGCGC CCTGGGGGAC CCCGGGCTCC GGAGGCC

57

105
 ATG CCG GCG TTG GCG CGC GAC GCG GGC ACC GTG CCG CTG CTC GTT GTT
 Met Pro Ala Leu Ala Arg Asp Ala Gly Thr Val Pro Leu Leu Val Val
 -27 -25 -20 -15
 153
 TTT TCT GCA ATG ATA TTT GGG ACT ATT ACA AAT CAA GAT CTG CCT GTG
 Phe Ser Ala Met Ile Phe Gly Thr Ile Thr Asn Gln Asp Leu Pro Val
 -10 -5 1 5
 201
 ATC AAG TGT GTT TTA ATC AAT CAT AAG AAC AAT GAT TCA TCA GTG GGG
 Ile Lys Cys Val Leu Ile Asn His Lys Asn Asp Ser Ser Val Gly
 10 15 20
 249
 AAG TCA TCA TCA TAT CCC ATG GTA TCA GAA TCC CCG GAA GAC CTC GGG
 Lys Ser Ser Ser Tyr Pro Met Val Ser Glu Ser Pro Glu Asp Leu Gly
 25 30 35
 297
 TGT GCG TTG AGA CCC CAG AGC TCA GGG ACA GTG TAC GAA GCT GCC GCT
 Cys Ala Leu Arg Pro Gln Ser Ser Gly Thr Val Tyr Glu Ala Ala Ala
 40 45 50
 345
 GTG GAA GTG GAT GTA TCT GCT TCC ATC ACA CTG CAA GTG CTG GTC GAT
 Val Glu Val Asp Val Ser Ala Ser Ile Thr Leu Gln Val Leu Val Asp
 55 60 65
 393
 GCC CCA GCG AAC ATT TCC TGT CTC TGG GTC TTT AAG CAC AGC TCC CTG
 Ala Pro Gly Asn Ile Ser Cys Leu Trp Val Phe Lys His Ser Ser Leu
 70 75 80 85
 441
 AAT TGC CAG CCA CAT TTT GAT TTA CAA AAC AGA GGA GTT GTC TCC ATG
 Asn Cys Gln Pro His Phe Asp Leu Gln Asn Arg Gly Val Val Ser Met
 90 95 100
 489
 GTC ATT TTG AAA ATG ACA GAA ACC CAA GCT GGA GAA TAC CTA CTT TTT
 Val Ile Leu Lys Met Thr Glu Thr Gln Ala Gly Glu Tyr Leu Leu Phe
 105 110 115
 537
 ATT CAG AGT GAA GCT ACC AAT TAC ACA ATA TTG TTT ACA GTG AGT ATA
 Ile Gln Ser Glu Ala Thr Asn Tyr Thr Ile Leu Phe Thr Val Ser Ile

120	125	130	
AGA AAT ACC CTG CTT TAC ACA TTA AGA AGA CCT TAC TTT AGA AAA ATG			585
Arg Asn Thr Leu Leu Tyr Thr Leu Arg Arg Pro Tyr Phe Arg Lys Met	140	145	
135			
GAA AAC CAG GAC GCC CTG GTC TGC ATA TCT GAG AGC GTT CCA GAG CCG			633
Glu Asn Gln Asp Ala Leu Val Cys Ile Ser Glu Ser Val Pro Glu Pro	155	160	165
150			
ATC GTG GAA TGG GTG CTT TGC GAT TCA CAG GGG GAA AGC TGT AAA GAA			681
Ile Val Glu Trp Val Leu Cys Asp Ser Gln Gly Glu Ser Cys Lys Glu	170	175	180
GAA AGT CCA GCT GTT GTT AAA AAG GAG GAG GAA AAA GTG CTT CAT GAA TTA			729
Glu Ser Pro Ala Val Val Lys Lys Glu Glu Lys Val Leu His Glu Leu	185	190	195
TTT GGG ACG GAC ATA AGG TGC TGT GCC AGA AAT GAA CTG GGC AGG GAA			777
Phe Gly Thr Asp Ile Arg Cys Cys Ala Arg Asn Glu Leu Gly Arg Glu	200	205	210
TGC ACC AGG CTG TTC ACA ATA GAT CTA AAT CAA ACT CCT CAG ACC ACA			825
Cys Thr Arg Leu Phe Thr Ile Asp Leu Asn Gln Thr Pro Gln Thr Thr	215	220	225
TTG CCA CAA TTA TTT CTT AAA GTA GGG GAA CCC TTA TGG ATA AGG TGC			873
Leu Pro Gln Leu Phe Leu Lys Val Gly Glu Pro Leu Trp Ile Arg Cys	230	235	240
AAA GCT GTT CAT GTG AAC CAT GGA TTC GGG CTC ACC TGG GAA TTA GAA			921
Lys Ala Val His Val Asn His Gly Phe Gly Leu Thr Trp Glu Leu Glu	250	255	260
AAC AAA GCA CTC GAG GAG GGC AAC TAC TTT GAG ATG AGT ACC TAT TCA			969
Asn Lys Ala Leu Glu Glu Tyr Phe Glu Met Ser Thr Tyr Ser	265	270	275

1017
 ACA AAC AGA ACT ATG ATA CGG ATT CTG TTT GCT TTT GTA TCA TCA GTG
 Thr Asn Arg Thr Met Ile Arg Ile Leu Phe Ala Phe Val Ser Ser Val
 280 285 290

1065
 GCA AGA AAC GAC ACC GGA TAC TAC ACT TGT TCC TCT TCA AAG CAT CCC
 Ala Arg Asn Asp Thr Gly Tyr Tyr Thr Cys Ser Ser Ser Lys His Pro
 295 300 305

1113
 AGT CAA TCA GCT TTG GTT ACC ATC GTA GGA AAG GGA TTT ATA AAT GCT
 Ser Gln Ser Ala Leu Val Thr Ile Val Gly Lys Gly Phe Ile Asn Ala
 310 315 320 325

1161
 ACC AAT TCA AGT GAA GAT TAT GAA ATT GAC CAA TAT GAA GAG TTT TGT
 Thr Asn Ser Ser Ser Glu Asp Tyr Glu Ile Asp Gln Tyr Glu Glu Phe Cys
 330 335 340

1209
 TTT TCT GTC AGG TTT AAA GCC TAC CCA CAA ATC AGA TGT ACG TGG ACC
 Phe Ser Val Arg Phe Lys Ala Tyr Pro Gln Ile Arg Cys Thr Trp Thr
 345 350 355

1257
 TTC TCT CGA AAA TCA TTT CCT TGT GAG CAA AAG GGT CTT GAT AAC GGA
 Phe Ser Arg Lys Ser Phe Pro Cys Glu Gln Lys Gly Leu Asp Asn Gly
 360 365 370

1305
 TAC AGC ATA TCC AAG TTT TGC AAT CAT AAG CAC CAG CCA GGA GAA TAT
 Tyr Ser Ile Ser Lys Phe Cys Asn His Lys His Gln Pro Gly Glu Tyr
 375 380 385

1353
 ATA TTC CAT GCA GAA AAT GAT GAT GCC CAA TTT ACC AAA ATG TTC ACG
 Ile Phe His Ala Glu Asn Asp Ala Gln Phe Thr Lys Met Phe Thr
 390 395 400 405

1401
 CTG AAT ATA AGA AGG AAA CCT CAA GTG CTC GCA GAA GCA TCG GCA AGT
 Leu Asn Ile Arg Arg Lys Pro Gln Val Leu Ala Glu Ala Ser Ala Ser
 410 415 420

1449
 CAG GCG TCC TGT TTC TCG GAT GGA TAC CCA TTA CCA TCT TGG ACC TGG
 Gln Ala Ser Cys Phe Ser Asp Gly Tyr Pro Leu Pro Ser Trp Thr Trp

425	430	435	
AAG AAG TGT TCA GAC AAG TCT CCC AAC TGC ACA GAA GAG ATC ACA GAA			1497
Lys Lys Cys Ser Asp Lys Ser Pro Asn Cys Thr Glu Glu Ile Thr Glu	440 445 450		
GGA GTC TGG AAT AGA AAG GCT AAC AGA AAA GTG TTT GGA CAG TGG GTG			1545
Gly Val Trp Asn Arg Lys Ala Asn Arg Lys Val Phe Gly Gln Trp Val	455 460 465		
TCG AGC AGT ACT CTA AAC ATG AGT GAA GCC ATA AAA GGG TTC CTG GTC			1593
Ser Ser Ser Thr Leu Asn Met Ser Ser Glu Ala Ile Lys Gly Phe Leu Val	470 475 480 485		
AAG TGC TGT GCA TAC AAT TCC CTT GGC ACA TCT TGT GAG ACG ATC CTT			1641
Lys Cys Cys Ala Tyr Asn Ser Ser Leu Gly Thr Ser Cys Glu Thr Ile Leu	490 495 500		
TTA AAC TCT CCA GGC CCC TTC CCT TTC ATC CAA GAC AAC ATC TCA TTC			1689
Leu Asn Ser Pro Gly Pro Phe Pro Phe Ile Gln Asp Asn Ile Ser Phe	505 510 515		
TAT GCA ACA ATT GGT GTT TGT CTC CTC TTC ATT GTC GTT TTA ACC CTG			1737
Tyr Ala Thr Ile Gly Val Cys Leu Leu Phe Phe Ile Val Val Leu Thr Leu	520 525 530		
CTA ATT TGT CAC AAG TAC AAA AAG CAA TTT AGG TAT GAA AGC CAG CTA			1785
Leu Ile Cys His Lys Tyr Lys Lys Gln Phe Arg Tyr Glu Ser Gln Leu	535 540 545		
CAG ATG GTA CAG GTG ACC GGC TCC TCA GAT AAT GAG TAC TTC TAC GTT			1833
Gln Met Val Gln Val Thr Gly Ser Ser Asp Asn Glu Tyr Phe Tyr Val	550 555 560 565		
GAT TTC AGA GAA TAT GAA TAT GAT CTC AAA TGG GAG TTT CCA AGA GAA			1881
Asp Phe Arg Glu Tyr Glu Tyr Asp Leu Lys Trp Glu Phe Pro Arg Glu	570 575 580		

AAT TTA GAG TTT GGG AAG GTA CTA GGA TCA GGT GCT TTT GGA AAA GTG 1929
 Asn Leu Glu Phe Gly Lys Val Leu Gly Ser Gly Ala Phe Gly Lys Val 595
 585
 ATG AAC GCA ACA GCT TAT GGA ATT AGC AAA ACA GGA GTC TCA ATC CAG 1977
 Met Asn Ala Thr Ala Tyr Gly Ile Ser Lys Thr Gly Val Ser Ile Gln 610
 600
 GTT GCC GTC AAA ATG CTG AAA GAA AAA GCA GAC AGC TCT GAA AGA GAG 2025
 Val Ala Val Lys Met Leu Lys Glu Lys Ala Asp Ser Ser Glu Arg Glu 625
 615
 GCA CTC ATG TCA GAA CTC AAG ATG ATG ACC CAG CTG GGA AGC CAC GAG 2073
 Ala Leu Met Ser Glu Leu Lys Met Met Thr Gln Leu Gly Ser His Glu 645
 630
 AAT ATT GTG AAC CTG CTG GGG GCG TGC ACA CTG TCA GGA CCA ATT TAC 2121
 Asn Ile Val Asn Leu Leu Gly Ala Cys Thr Leu Ser Gly Pro Ile Tyr 660
 650
 TTG ATT TTT GAA TAC TGT TGC TAT GGT GAT CTT CTC AAC TAT CTA AGA 2169
 Leu Ile Phe Glu Tyr Cys Cys Tyr Gly Asp Leu Leu Asn Tyr Leu Arg 675
 665
 AGT AAA AGA GAA AAA TTT CAC AGG ACT TGG ACA GAG ATT TTC AAG GAA 2217
 Ser Lys Arg Glu Lys Phe His Arg Thr Trp Thr Glu Ile Phe Lys Glu 690
 680
 CAC AAT TTC AGT TTT TAC CCC ACT TTC CAA TCA CAT CCA AAT TCC AGC 2265
 His Asn Phe Ser Phe Tyr Pro Thr Phe Gln Ser His Pro Asn Ser Ser 705
 695
 ATG CCT GGT TCA AGA GAA GTT CAG ATA CAC CCG GAC TCG GAT CAA ATC 2313
 Met Pro Gly Ser Arg Glu Val Gln Ile His Pro Asp Ser Asp Gln Ile 725
 710
 TCA GGG CTT CAT GGG AAT TCA TTT CAC TCT GAA GAT GAA ATT GAA TAT 2361
 Ser Gly Leu His Gly Asn Ser Phe His Ser Glu Asp Glu Ile Glu Tyr

	730	735	740	
GAA AAC CAA AAA AGG CTG GAA GAA GAG GAG GAC TTG AAT GTG CTT ACA				2409
Glu Asn Gln Lys Arg Leu Glu Glu Glu Asp Leu Asn Val Leu Thr	745	750	755	
TTT GAA GAT CTT CTT TGC TTT GCA TAT CAA GTT GCC AAA GGA ATG GAA				2457
Phe Glu Asp Leu Leu Cys Phe Ala Tyr Gln Val Ala Lys Gly Met Glu	760	765	770	
TTT CTG GAA TTT AAG TCG TGT GTT CAC AGA GAC CTG GCC GCC AGG AAC				2505
Phe Leu Glu Phe Lys Ser Cys Val His Arg Asp Leu Ala Ala Arg Asn	775	780	785	
GTG CTT GTC ACC CAC GGG AAA GTG GTG AAG ATA TGT GAC TTT GGA TTG				2553
Val Leu Val Thr His Gly Lys Val Val Lys Ile Cys Asp Phe Gly Leu	790	795	800	
GCT CGA GAT ATC ATG AGT GAT TCC AAC TAT GTT GTC AGG GCC AAT GCC				2601
Ala Arg Asp Ile Met Ser Ser Asp Ser Asn Tyr Val Val Arg Gly Asn Ala	810	815	820	
CGT CTG CCT GTA AAA TGG ATG GCC CCC GAA AGC CTG TTT GAA GGC ATC				2649
Arg Leu Pro Val Lys Trp Met Ala Pro Glu Ser Leu Phe Glu Gly Ile	825	830	835	
TAC ACC ATT AAG AGT GAT GTC TGG TCA TAT GGA ATA TTA CTG TGG GAA				2697
Tyr Thr Ile Lys Ser Asp Val Trp Ser Tyr Gly Ile Leu Leu Trp Glu	840	845	850	
ATC TTC TCA CTT GGT GTG AAT CCT TAC CCT GGC ATT CCG GTT GAT GCT				2745
Ile Phe Ser Leu Gly Val Asn Pro Tyr Pro Gly Ile Pro Val Asp Ala	855	860	865	
AAC TTC TAC AAA CTG ATT CAA AAT GGA TTT AAA ATG GAT CAG CCA TTT				2793
Asn Phe Tyr Lys Leu Ile Gln Asn Gly Phe Lys Met Asp Gln Pro Phe	870	875	880	
			885	

2841 TAT GCT ACA GAA GAA ATA TAC ATT ATA ATG CAA TCC TGC TGG GCT TTT
 Tyr Ala Thr Glu Glu Ile Tyr Ile Ile Met Gln Ser Cys Trp Ala Phe
 890 895
 2889 GAC TCA AGG AAA CGG CCA TCC TTC CCT AAT TTG ACT TCG TTT TTA GGA
 Asp Ser Arg Lys Arg Pro Ser Phe Pro Asn Leu Thr Ser Phe Leu Gly
 905 910
 2937 TGT CAG CTG GCA GAT GCA GAA GAA GCG ATG TAT CAG AAT GTG GAT GGC
 Cys Gln Leu Ala Asp Ala Glu Glu Ala Met Tyr Gln Asn Val Asp Gly
 920 925
 2985 CGT GTT TCG GAA TGT CCT CAC ACC TAC CAA AAC AGG CGA CCT TTC AGC
 Arg Val Ser Glu Cys Pro His Thr Tyr Gln Asn Arg Arg Pro Phe Ser
 935 940 945
 3033 AGA GAG ATG GAT TTG GGG CTA CTC TCT CCG CAG GCT CAG GTC GAA GAT
 Arg Glu Met Asp Leu Gly Leu Leu Ser Pro Gln Ala Gln Val Glu Asp
 950 955 960
 3086 TCG TAGAGGAACA ATTTAGTTTT AAGGACTTCA TCCCTCCACC TATCCCTAAC
 Ser
 3146 AGGCTGTAGA TTACCAAAC AAGATTAATT TCATCACTAA AAGAAATCT ATTATCAACT
 3206 GCTGCTTCAC CAGACTTTTC TCTAGAAGCC GTCTGCGTTT ACTCTTGTTT TCAAAGGGAC
 3266 TTTTGTAAAA TCAAATCATC CTGTCACAAG GCAGGAGGAG CTGATAATGA ACTTTATGG
 3326 AGCATTGATC TGCATCCAAG GCCTTCTCAG GCCGGCTTGA GTGAATTGTG TACCTGAAGT
 3386 ACAGTATATT CTTGTAAATA CATAAACAA AAGCATTTTG CTAAGGAGAA GCTAATATGA
 3446 TTTTAAAGT CTATGTTTTA AAATAATATG TAAATTTTC AGCTATTTAG TGATATATTT
 3501 TATGGGTGGG AATAAAATTT CTACTACAGA AAAAAAAAAA AAAAAA

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 993 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

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Met Pro Ala Leu Ala Arg Asp Ala Gly Thr Val Pro Leu Leu Val Val
-27 -25 -20 -15
Phe Ser Ala Met Ile Phe Gly Thr Ile Thr Asn Gln Asp Leu Pro Val
-10 -5 1 5
Ile Lys Cys Val Leu Ile Asn His Lys Asn Asn Asp Ser Ser Val Gly
10 15 20
Lys Ser Ser Tyr Pro Met Val Ser Glu Ser Pro Glu Asp Leu Gly
25 30 35
Cys Ala Leu Arg Pro Gln Ser Ser Gly Thr Val Tyr Glu Ala Ala Ala
40 45 50
Val Glu Val Asp Val Ser Ala Ser Ile Thr Leu Gln Val Leu Val Asp
55 60 65
Ala Pro Gly Asn Ile Ser Cys Leu Trp Val Phe Lys His Ser Ser Leu
70 75 80 85
Asn Cys Gln Pro His Phe Asp Leu Leu Gln Asn Arg Gly Val Val Ser Met
90 95 100
Val Ile Leu Lys Met Thr Glu Thr Gln Ala Gly Glu Tyr Leu Leu Phe
105 110 115

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Ile Gln Ser Glu Ala Thr Asn Tyr Thr Ile Leu Phe Thr Val Ser Ile
 120 125 130
 Arg Asn Thr Leu Leu Tyr Thr Leu Arg Arg Pro Tyr Phe Arg Lys Met
 135 140 145
 Glu Asn Gln Asp Ala Leu Val Cys Ile Ser Glu Ser Val Pro Glu Pro
 150 155 160 165
 Ile Val Glu Trp Val Leu Cys Asp Ser Gln Gly Glu Ser Cys Lys Glu
 170 175 180
 Glu Ser Pro Ala Val Val Lys Lys Glu Glu Lys Val Leu His Glu Leu
 185 190 195
 Phe Gly Thr Asp Ile Arg Cys Cys Ala Arg Asn Glu Leu Gly Arg Glu
 200 205 210
 Cys Thr Arg Leu Phe Thr Ile Asp Leu Asn Gln Thr Pro Gln Thr Thr
 215 220 225
 Leu Pro Gln Leu Phe Leu Lys Val Gly Glu Pro Leu Trp Ile Arg Cys
 230 235 240 245
 Lys Ala Val His Val Asn His Gly Phe Gly Leu Thr Trp Glu Leu Glu
 250 255 260
 Asn Lys Ala Leu Glu Glu Gly Asn Tyr Phe Glu Met Ser Thr Tyr Ser
 265 270 275
 Thr Asn Arg Thr Met Ile Arg Ile Leu Phe Ala Phe Val Ser Ser Val
 280 285 290
 Ala Arg Asn Asp Thr Gly Tyr Tyr Thr Cys Ser Ser Ser Lys His Pro
 295 300 305
 Ser Gln Ser Ala Leu Val Thr Ile Val Gly Lys Gly Phe Ile Asn Ala
 310 315 320 325

Thr Asn Ser Ser Glu Asp Tyr Glu Ile Asp Gln Tyr Glu Glu Phe Cys
 330 335 340
 Phe Ser Val Arg Phe Lys Ala Tyr Pro Gln Ile Arg Cys Thr Trp Thr
 345 350 355
 Phe Ser Arg Lys Ser Phe Pro Cys Glu Gln Lys Gly Leu Asp Asn Gly
 360 365 370
 Tyr Ser Ile Ser Lys Phe Cys Asn His Lys His Gln Pro Gly Glu Tyr
 375 380 385
 Ile Phe His Ala Glu Asn Asp Ala Gln Phe Thr Lys Met Phe Thr
 390 395 400 405
 Leu Asn Ile Arg Arg Lys Pro Gln Val Leu Ala Glu Ala Ser Ala Ser
 410 415 420
 Gln Ala Ser Cys Phe Ser Asp Gly Tyr Pro Leu Pro Ser Trp Thr Trp
 425 430 435
 Lys Lys Cys Ser Asp Lys Ser Pro Asn Cys Thr Glu Glu Ile Thr Glu
 440 445 450
 Gly Val Trp Asn Arg Lys Ala Asn Arg Lys Val Phe Gly Gln Trp Val
 455 460 465
 Ser Ser Ser Thr Leu Asn Met Ser Glu Ala Ile Lys Gly Phe Leu Val
 470 475 480 485
 Lys Cys Cys Ala Tyr Asn Ser Leu Gly Thr Ser Cys Glu Thr Ile Leu
 490 495 500
 Leu Asn Ser Pro Gly Pro Phe Pro Phe Ile Gln Asp Asn Ile Ser Phe
 505 510 515
 Tyr Ala Thr Ile Gly Val Cys Leu Leu Phe Ile Val Val Leu Thr Leu
 520 525 530

Leu Ile Cys His Lys Tyr Lys Lys Gln Phe Arg Tyr Glu Ser Gln Leu
 535 540
 Gln Met Val Gln Val Thr Gly Ser Ser Asp Asn Glu Tyr Phe Tyr Val
 550 555 560
 Asp Phe Arg Glu Tyr Glu Tyr Asp Leu Lys Trp Glu Phe Pro Arg Glu
 570 575 580
 Asn Leu Glu Phe Gly Lys Val Leu Gly Ser Gly Ala Phe Gly Lys Val
 585 590
 Met Asn Ala Thr Ala Tyr Gly Ile Ser Lys Thr Gly Val Ser Ile Gln
 600 605
 Val Ala Val Lys Met Leu Lys Glu Lys Ala Asp Ser Ser Glu Arg Glu
 615 620 625
 Ala Leu Met Ser Glu Leu Lys Met Met Thr Gln Leu Gly Ser His Glu
 630 635 640
 Asn Ile Val Asn Leu Leu Gly Ala Cys Thr Leu Ser Gly Pro Ile Tyr
 650 655 660
 Leu Ile Phe Glu Tyr Cys Cys Tyr Gly Asp Leu Leu Asn Tyr Leu Arg
 665 670 675
 Ser Lys Arg Glu Lys Phe His Arg Thr Trp Thr Glu Ile Phe Lys Glu
 680 685 690
 His Asn Phe Ser Phe Tyr Pro Thr Phe Gln Ser His Pro Asn Ser Ser
 695 700 705
 Met Pro Gly Ser Arg Glu Val Gln Ile His Pro Asp Ser Asp Gln Ile
 710 715 720 725
 Ser Gly Leu His Gly Asn Ser Phe His Ser Glu Asp Glu Ile Glu Tyr
 730 735 740

Glu Asn Gln Lys Arg Leu Glu Glu Glu Asp Leu Asn Val Leu Thr
 745 750 755
 Phe Glu Asp Leu Leu Cys Phe Ala Tyr Gln Val Ala Lys Gly Met Glu
 760 765 770
 Phe Leu Glu Phe Lys Ser Cys Val His Arg Asp Leu Ala Ala Arg Asn
 775 780 785
 Val Leu Val Thr His Gly Lys Val Val Lys Ile Cys Asp Phe Gly Leu
 790 795 800 805
 Ala Arg Asp Ile Met Ser Asp Ser Asn Tyr Val Val Arg Gly Asn Ala
 810 815 820
 Arg Leu Pro Val Lys Trp Met Ala Pro Glu Ser Leu Phe Glu Gly Ile
 825 830 835
 Tyr Thr Ile Lys Ser Asp Val Trp Ser Tyr Gly Ile Leu Leu Trp Glu
 840 845 850
 Ile Phe Ser Leu Gly Val Asn Pro Tyr Pro Gly Ile Pro Val Asp Ala
 855 860 865
 Asn Phe Tyr Lys Leu Ile Gln Asn Gly Phe Lys Met Asp Gln Pro Phe
 870 875 880 885
 Tyr Ala Thr Glu Glu Ile Tyr Ile Ile Met Gln Ser Cys Trp Ala Phe
 890 895 900
 Asp Ser Arg Lys Arg Pro Ser Phe Pro Asn Leu Thr Ser Phe Leu Gly
 905 910 915
 Cys Gln Leu Ala Asp Ala Glu Glu Ala Met Tyr Gln Asn Val Asp Gly
 920 925 930
 Arg Val Ser Glu Cys Pro His Thr Tyr Gln Asn Arg Arg Pro Phe Ser
 935 940 945

Arg Glu Met Asp Leu Gly Leu Ser Pro Gln Ala Gln Val Glu Asp
950 955 960 965

Ser

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 5406 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(v) FRAGMENT TYPE: N-terminal

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 208..4311

(ix) FEATURE:

- (A) NAME/KEY: mat_peptide
- (B) LOCATION: 265..4308

(ix) FEATURE:

- (A) NAME/KEY: sig_peptide
- (B) LOCATION: 208..264

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

CTGTGTCCCG CAGCCGGATA ACCTGGCTGA CCCGATTCCG CGGACACCCG TGCAGCCGCG 60
 GCTGGAGCCA GGGCGCCGGT GCCCGCGCTC TCCCCGGTCT TCGCGTGCGG GGGCCGATAC 120
 CGCCTCTGTG ACTTCTTTGC GGGCCAGGGA CGGAGAAGGA GTCTGTGCCT GAGAAACTGG 180
 GCTCTGTGCC CAGGCGCGAG GTGCAGG ATG GAG AGC AAG GGC CTG CTA GCT 231
 Met Glu Ser Lys Gly Leu Leu Ala
 -19
 -15
 GTC GCT CTG TGG TTC TGC GTG GAG ACC CGA GCC GCC TCT GTG GGT TTG 279
 Val Ala Leu Trp Phe Cys Val Glu Thr Arg Ala Ala Ser Val Gly Leu
 -10
 -5
 1
 CCT GGC GAT TTT CTC CAT CCC CCC AAG CTC AGC ACA CAG AAA GAC ATA 327
 Pro Gly Asp Phe Leu His Pro Pro Lys Leu Ser Thr Gln Lys Asp Ile
 10
 15
 20
 CTG ACA ATT TTG GCA AAT ACA ACC CTT CAG ATT ACT TGC AGG GGA CAG 375
 Leu Thr Ile Leu Ala Asn Thr Thr Leu Gln Ile Thr Cys Arg Gly Gln
 25
 30
 35
 CGG GAC CTG GAC TGG CTT TGG CCC AAT GCT CAG CGT GAT TCT GAG GAA 423
 Arg Asp Leu Asp Trp Leu Trp Pro Asn Ala Gln Arg Asp Ser Glu Glu
 40
 45
 50
 AGG GTA TTG GTG ACT GAA TGC GGC GGT GGT GAC AGT ATC TTC TGC AAA 471
 Arg Val Leu Val Thr Glu Cys Gly Gly Asp Ser Ile Phe Cys Lys
 55
 60
 65
 ACA CTC ACC ATT CCC AGG GTG GTT GGA AAT GAT ACT GGA GCC TAC AAG 519
 Thr Leu Thr Thr Ile Pro Arg Val Val Gly Asn Asp Thr Gly Ala Tyr Lys
 70
 75
 80
 85
 TGC TCG TAC CGG GAC GTC GAC ATA GCC TCC ACT GTT TAT GTC TAT GTT 567
 Cys Ser Tyr Arg Asp Val Asp Ile Ala Ser Thr Val Tyr Val Tyr Val
 90
 95
 100

615 CGA GAT TAC AGA TCA CCA TTC ATC GCC TCT GTC AGT GAC CAG CAT GGC.
 Arg Asp Tyr Arg Ser Pro Phe Ile Ala Ser Val Ser Asp Gln His Gly
 105 110 115
 663 ATC GTG TAC ATC ACC GAG AAC AAG AAC AAA ACT GTG GTG ATC CCC TGC
 Ile Val Tyr Ile Thr Glu Asn Lys Asn Lys Thr Val Val Ile Pro Cys
 120 125 130
 711 CGA GGG TCG ATT TCA AAC CTC AAT GTG TCT CTT TGC GCT AGG TAT CCA
 Arg Gly Ser Ile Ser Asn Leu Asn Val Ser Leu Cys Ala Arg Tyr Pro
 135 140 145
 759 GAA AAG AGA TTT GTT CCG GAT GGA AAC AGA ATT TCC TGG GAC AGC GAG
 Glu Lys Arg Phe Val Pro Asp Gly Asn Arg Ile Ser Trp Asp Ser Glu
 150 155 160 165
 807 ATA GGC TTT ACT CTC CCC AGT TAC ATG ATC AGC TAT GCC GGC ATG GTC
 Ile Gly Phe Thr Leu Pro Ser Tyr Met Ile Ser Tyr Ala Gly Met Val
 170 175 180
 855 TTC TGT GAG GCA AAG ATC AAT GAT GAA ACC TAT CAG TCT ATC ATG TAC
 Phe Cys Glu Ala Lys Ile Asn Asp Glu Thr Tyr Gln Ser Ile Met Tyr
 185 190 195
 903 ATA GTT GTG GTT GTA GGA TAT AGG ATT TAT GAT GTG ATT CTG AGC CCC
 Ile Val Val Val Val Gly Tyr Arg Ile Tyr Asp Val Ile Leu Ser Pro
 200 205 210
 951 CCG CAT GAA ATT GAG CTA TCT GCC GGA GAA AAA CTT GTC TTA AAT TGT
 Pro His Glu Ile Glu Leu Ser Ala Gly Glu Lys Leu Val Leu Asn Cys
 215 220 225
 999 ACA GCG AGA ACA GAG CTC AAT GTG GGG CTT GAT TTC ACC TGG CAC TCT
 Thr Ala Arg Thr Glu Leu Asn Val Gly Leu Asp Phe Thr Trp His Ser
 230 235 240 245
 1047 CCA CCT TCA AAG TCT CAT CAT AAG AAG ATT GTA AAC CGG GAT GTG AAA
 Pro Pro Ser Lys Ser His His Lys Lys Ile Val Asn Arg Asp Val Lys

250	255	260	1095
CCC TTT CCT GGG ACT GTG GCG AAG ATG TTT TTG AGC ACC TTG ACA ATA			
Pro Phe Pro Gly Thr Val Ala Lys Met Phe Leu Ser Thr Leu Thr Ile			
265	270	275	
GAA AGT GTG ACC AAG AGT GAC CAA GGG GAA TAC ACC TGT GTA GCG TCC			1143
Glu Ser Val Thr Lys Ser Asp Gln Gly Glu Tyr Thr Cys Val Ala Ser			
280	285	290	
AGT GGA CGG ATG ATC AAG AGA AAT AGA ACA TTT GTC CGA GTT CAC ACA			1191
Ser Gly Arg Met Ile Lys Arg Asn Arg Thr Phe Val Arg Val His Thr			
295	300	305	
AAG CCT TTT ATT GCT TTC GGT AGT GGG ATG AAA TCT TTG GTG GAA GCC			1239
Lys Pro Phe Ile Ala Phe Phe Gly Ser Gly Met Lys Ser Leu Val Glu Ala			
310	315	320	
ACA GTG GGC AGT CAA GTC CGA ATC CCT GTC AAG TAT CTC AGT TAC CCA			1287
Thr Val Gly Ser Gln Val Arg Ile Pro Val Lys Tyr Leu Ser Tyr Pro			
330	335	340	
GCT CCT GAT ATC AAA TGG TAC AGA AAT GGA AGG CCC ATT GAG TCC AAC			1335
Ala Pro Asp Ile Lys Trp Tyr Arg Asn Gly Arg Pro Ile Glu Ser Asn			
345	350	355	
TAC ACA ATG ATT GTT GGC GAT GAA CTC ACC ATC ATG GAA GTG ACT GAA			1383
Tyr Thr Met Ile Val Gly Asp Glu Leu Thr Ile Met Glu Val Thr Glu			
360	365	370	
AGA GAT GCA GGA AAC TAC ACG GTC ATC CTC ACC AAC CCC ATT TCA ATG			1431
Arg Asp Ala Gly Asn Tyr Thr Val Ile Leu Thr Asn Pro Ile Ser Met			
375	380	385	
GAG AAA CAG AGC CAC ATG GTC TCT CTG GTT GTG AAT GTC CCA CCC CAG			1479
Glu Lys Gln Ser His Met Val Ser Leu Val Val Asn Val Pro Pro Gln			
390	395	400	
		405	

1527 ATC GGT GAG AAA GCC TTG ATC TCG CCT ATG GAT TCC TAC CAG TAT GGG
 Ile Gly Glu Lys Ala Leu Ile Ser Pro Met Asp Ser Tyr Gln Tyr Gly
 410 415 420

1575 ACC ATG CAG ACA TTG ACA TGC ACA GTC TAC GCC AAC CCT CCC CTG CAC
 Thr Met Gln Thr Leu Thr Cys Thr Val Tyr Ala Asn Pro Pro Leu His
 425 430 435

1623 CAC ATC CAG TGG TAC TGG CAG CTA GAA GAA GCC TGC TCC TAC AGA CCC
 His Ile Gln Trp Tyr Trp Gln Leu Leu Glu Ala Cys Ser Tyr Arg Pro
 440 445 450

1671 GGC CAA ACA AGC CCG TAT GCT TGT AAA GAA TGG AGA CAC GTG GAG GAT
 Gly Gln Thr Ser Pro Tyr Ala Cys Lys Glu Trp Arg His Val Glu Asp
 455 460 465

1719 TTC CAG GGG GGA AAC AAG ATC GAA GTC ACC AAA AAC CAA TAT GCC CTG
 Phe Gln Gly Gly Asn Lys Ile Glu Val Thr Lys Asn Gln Tyr Ala Leu
 470 475 480 485

1767 ATT GAA GGA AAA AAC AAA ACT GTA AGT ACG CTG GTC ATC CAA GCT GCC
 Ile Glu Gly Lys Asn Lys Thr Val Ser Thr Leu Val Ile Gln Ala Ala
 490 495 500

1815 AAC GTG TCA GCG TTG TAC AAA TGT GAA GCC ATC AAC AAA GCG GGA CGA
 Asn Val Ser Ala Leu Tyr Lys Cys Glu Ala Ile Asn Lys Ala Gly Arg
 505 510 515

1863 GGA GAG AGG GTC ATC TCC TTC CAT GTG ATC AGG GGT CCT GAA ATT ACT
 Gly Glu Arg Val Ile Ser Phe His Val Ile Arg Gly Pro Glu Ile Thr
 520 525 530

1911 GTG CAA CCT GCT GCC CAG CCA ACT GAG CAG GAG AGT GTG TCC CTG TTG
 Val Gln Pro Ala Ala Gln Pro Thr Glu Gln Glu Ser Val Ser Leu Leu
 535 540 545

1959 TGC ACT GCA GAC AGA AAT ACG TTT GAG AAC CTC ACG TGG TAC AAG CTT
 Cys Thr Ala Asp Arg Asn Thr Phe Glu Asn Leu Thr Trp Tyr Lys Leu

550	555	560	565	
GGC TCA CAG GCA ACA TCG GTC CAC ATG GGC GAA TCA CTC ACA CCA GTT				2007
Gly Ser Gln Ala Thr Ser Val His Met Gly Glu Ser Leu Thr Pro Val				
570	575	580		
TGC AAG AAC TTG GAT GCT CTT TGG AAA CTG AAT GGC ACC ATG TTT TCT				2055
Cys Lys Asn Leu Asp Ala Leu Trp Lys Leu Asn Gly Thr Met Phe Ser				
585	590	595		
AAC AGC ACA AAT GAC ATC TTG ATT GTG GCA TTT CAG AAT GCC TCT CTG				2103
Asn Ser Thr Asn Asp Ile Leu Ile Val Ala Phe Gln Asn Ala Ser Leu				
600	605	610		
CAG GAC CAA GGC GAC TAT GTT TGC TCT GCT CAA GAT AAG AAG ACC AAG				2151
Gln Asp Gln Gly Asp Tyr Val Cys Ser Ala Gln Asp Lys Lys Thr Lys				
615	620	625		
AAA AGA CAT TGC CTG GTC AAA CAG CTC ATC ATC CTA GAG CGC ATG GCA				2199
Lys Arg His Cys Leu Val Lys Gln Leu Ile Ile Leu Glu Arg Met Ala				
630	635	640		
CCC ATG ATC ACC GGA AAT CTG GAG AAT CAG ACA ACA ACC ATT GGC GAG				2247
Pro Met Ile Thr Gly Asn Leu Glu Asn Gln Thr Thr Thr Ile Gly Glu				
650	655	660		
ACC ATT GAA GTG ACT TGC CCA GCA TCT GGA AAT CCT ACC CCA CAC ATT				2295
Thr Ile Glu Val Thr Cys Pro Ala Ser Gly Asn Pro Thr Pro His Ile				
665	670	675		
ACA TGG TTC AAA GAC AAC GAG ACC CTG GTA GAA GAT TCA GGC ATT GTA				2343
Thr Trp Phe Lys Asp Asn Glu Thr Leu Val Glu Asp Ser Gly Ile Val				
680	685	690		
CTG AGA GAT GGG AAC CGG AAC CTG ACT ATC CGC AGG GTG AGG AAG GAG				2391
Leu Arg Asp Gly Asn Arg Asn Leu Thr Ile Arg Val Arg Lys Glu				
695	700	705		

2439 GAT GGA GGC CTC TAC ACC TGC CAG GCC TGC AAT GTC CTT GGC TGT GCA
 Asp Gly Gly Leu Tyr Thr Cys Gln Ala Cys Asn Val Leu Gly Cys Ala 725
 710
 2487 AGA GCG GAG ACG CTC TTC ATA ATA GAA GGT GCC CAG GAA AAG ACC AAC
 Arg Ala Glu Thr Leu Phe Ile Ile Glu Gly Ala Gln Glu Lys Thr Asn 740
 730
 2535 TTG GAA GTC ATT ATC CTC GTC GGC ACT GCA GTG ATT GCC ATG TTC TTC
 Leu Glu Val Ile Ile Leu Val Gly Thr Ala Val Ile Ala Met Phe Phe 755
 745
 2583 TGG CTC CTT CTT GTC ATT CTC GTA CGG ACC GTT AAG CCG GCC AAT GAA
 Trp Leu Leu Leu Val Ile Leu Val Arg Thr Val Lys Arg Ala Asn Glu 770
 760
 2631 GGG GAA CTG AAG ACA GGC TAC TTG TCT ATT GTC ATG GAT CCA GAT GAA
 Gly Glu Leu Lys Thr Gly Tyr Leu Ser Ile Val Met Asp Pro Asp Glu 785
 775
 2679 TTG CCC TTG GAT GAG CGC TGT GAA CGC TTG CCT TAT GAT GCC AGC AAG
 Leu Pro Leu Asp Glu Arg Cys Glu Arg Leu Pro Tyr Asp Ala Ser Lys 805
 790
 2727 TGG GAA TTC CCC AGG GAC CGG CTG AAA CTA GGA AAA CCT CTT GGC CGC
 Trp Glu Phe Pro Arg Asp Arg Leu Lys Leu Gly Lys Pro Leu Gly Arg 820
 810
 2775 GGT GCC TTC GGC CAA GTG ATT GAG GCA GAC GCT TTT GGA ATT GAC AAG
 Gly Ala Phe Gly Gln Val Ile Glu Ala Asp Ala Phe Gly Ile Asp Lys 835
 825
 2823 ACA GCG ACT TGC AAA ACA GTA GCC GTC AAG ATG TTG AAA GAA GGA GCA
 Thr Ala Thr Cys Lys Thr Val Ala Val Lys Met Leu Lys Glu Gly Ala 850
 840
 2871 ACA CAC AGC GAG CAT CGA GCC CTC ATG TCT GAA CTC AAG ATC CTC ATC
 Thr His Ser Glu His Arg Ala Leu Met Ser Glu Leu Lys Ile Leu Ile

855	860	865	2919
CAC ATT GGT CAC CAT CTC AAT GTG AAC CTC CTA GGC GCC TGC ACC			
His Ile Gly His His Leu Asn Val Val Asn Leu Leu Gly Ala Cys Thr			
870	875	880	885
AAG CCG GGA GGG CCT CTC ATG GTG ATT GTG GAA TTC TCG AAG TTT GGA			
Lys Pro Gly Gly Pro Leu Met Val Ile Val Glu Phe Ser Lys Phe Gly			
890	895	900	2967
AAC CTA TCA ACT TAC TTA CGG GGC AAG AGA AAT GAA TTT GTT CCC TAT			
Asn Leu Ser Thr Tyr Leu Arg Gly Lys Arg Asn Glu Phe Val Pro Tyr			
905	910	915	3015
AAG AGC AAA GGG GCA CGC TTC CGC CAG GGC AAG GAC TAC GTT GGG GAG			
Lys Ser Lys Gly Ala Arg Phe Arg Gln Gly Lys Asp Tyr Val Gly Glu			
920	925	930	3063
CTC TCC GTG GAT CTG AAA AGA CGC TTG GAC AGC ATC ACC AGC AGC CAG			
Leu Ser Val Asp Leu Lys Arg Arg Leu Asp Ser Ile Thr Ser Ser Gln			
935	940	945	3111
AGC TCT GCC AGC TCA GGC TTT GTT GAG GAG AAA TCG CTC AGT GAT GTA			
Ser Ser Ala Ser Ser Gly Phe Val Glu Glu Lys Ser Leu Ser Asp Val			
950	955	960	3159
GAG GAA GAA GAA GCT TCT GAA GAA CTG TAC AAG GAC TTC CTG ACC TTG			
Glu Glu Glu Glu Ala Ser Glu Glu Leu Tyr Lys Asp Phe Leu Thr Leu			
970	975	980	3207
GAG CAT CTC ATC TGT TAC AGC TTC CAA GTG GCT AAG GGC ATG GAG TTC			
Glu His Leu Ile Cys Tyr Ser Phe Gln Val Ala Lys Gly Met Glu Phe			
985	990	995	3255
TTG GCA TCA AGG AAG TGT ATC CAC AGG GAC CTG GCA GCA AAC ATT			
Leu Ala Ser Arg Lys Cys Ile His Arg Asp Leu Ala Ala Arg Asn Ile			
1000	1005	1010	3303

CTC CTA TCG GAG AAG AAT GTG GTT AAG ATC TGT GAC TTC GGC TTG GCC 3351
 Leu Leu Ser Glu Lys Asn Val Val Lys Ile Cys Asp Phe Gly Leu Ala
 1015 1020 1025
 CGG GAC ATT TAT AAA GAC CCG GAT TAT GTC AGA AAA GGA GAT GCC CGA 3399
 Arg Asp Ile Tyr Lys Asp Pro Asp Tyr Val Arg Lys Gly Asp Ala Arg
 1030 1035 1040 1045
 CTC CCT TTG AAG TGG ATG GCC CCG GAA ACC ATT TTT GAC AGA GTA TAC 3447
 Leu Pro Leu Lys Trp Met Ala Pro Glu Thr Ile Phe Asp Arg Val Tyr
 1050 1055 1060
 ACA ATT CAG AGC GAT GTG TGG TCT TTC GGT GTG TTG CTC TGG GAA ATA 3495
 Thr Ile Gln Ser Asp Val Trp Ser Phe Gly Val Leu Leu Trp Glu Ile
 1065 1070 1075
 TTT TCC TTA GGT GCC TCC CCA TAC CCT GGG GTC AAG ATT GAT GAA GAA 3543
 Phe Ser Leu Gly Ala Ser Pro Tyr Pro Gly Val Lys Ile Asp Glu Glu
 1080 1085 1090
 TTT TGT AGG AGA TTG AAA GAA GGA ACT AGA ATG CGG GCT CCT GAC TAC 3591
 Phe Cys Arg Arg Leu Lys Glu Gly Thr Arg Met Arg Ala Pro Asp Tyr
 1095 1100 1105
 ACT ACC CCA GAA ATG TAC CAG ACC ATG CTG GAC TGC TGG CAT GAG GAC 3639
 Thr Thr Pro Glu Met Tyr Gln Thr Met Leu Asp Cys Trp His Glu Asp
 1110 1115 1120 1125
 CCC AAC CAG AGA CCC TCG TTT TCA GAG TTG GTG GAG CAT TTG GGA AAC 3687
 Pro Asn Gln Arg Pro Ser Phe Ser Glu Leu Val Glu His Leu Gly Asn
 1130 1135 1140
 CTC CTG CAA GCA AAT GCG CAG CAG GAT GGC AAA GAC TAT ATT GTT CTT 3735
 Leu Leu Gln Ala Asn Ala Gln Gln Asp Gly Lys Asp Tyr Ile Val Leu
 1145 1150 1155
 CCA ATG TCA GAG ACA CTG AGC ATG GAA GAG GAT TCT GGA CTC TCC CTG 3783
 Pro Met Ser Glu Thr Leu Ser Met Glu Glu Asp Ser Gly Leu Ser Leu

1160	1165	1170	
CCT ACC TCA CCT GTT TCC TGT ATG GAG GAA GAG GTG TGC GAC CCC			3831
Pro Thr Ser Pro Val Ser Cys Met Glu Glu Glu Val Cys Asp Pro	1180	1185	
1175			
AAA TTC CAT TAT GAC AAC ACA GCA GGA ATC AGT CAT TAT CTC CAG AAC			3879
Lys Phe His Tyr Asp Asn Thr Ala Gly Ile Ser His Tyr Leu Gln Asn	1195	1200	
1190			
AGT AAG CGA AAG AGC CGG CCA GTG AGT GTA AAA ACA TTT GAA GAT ATC			3927
Ser Lys Arg Lys Ser Arg Pro Val Ser Val Lys Thr Phe Glu Asp Ile	1210	1215	
			1220
CCA TTG GAG GAA CCA GAA GTA AAA GTG ATC CCA GAT GAC AGC CAG ACA			3975
Pro Leu Glu Glu Pro Glu Val Lys Val Ile Pro Asp Asp Ser Gln Thr	1225	1230	
			1235
GAC AGT GGG ATG GTC CTT GCA TCA GAA GAG CTG AAA ACT CTG GAA GAC			4023
Asp Ser Gly Met Val Leu Ala Ser Glu Glu Leu Lys Thr Leu Glu Asp	1240	1245	
			1250
AGG AAC AAA TTA TCT CCA TCT TTT GGT GGA ATG ATG CCC AGT AAA AGC			4071
Arg Asn Lys Leu Ser Pro Ser Phe Gly Gly Met Met Pro Ser Lys Ser	1255	1260	
			1265
AGG GAG TCT GTG GCC TCG GAA GGC TCC AAC CAG ACC AGT GGC TAC CAG			4119
Arg Glu Ser Val Ala Ser Glu Gly Ser Asn Gln Thr Ser Gly Tyr Gln	1270	1275	
			1280
TCT GGG TAT CAC TCA GAT GAC ACA GAC ACC ACC GTG TAC TCC AGC GAC			4167
Ser Gly Tyr His Ser Asp Asp Thr Asp Thr Thr Val Tyr Ser Ser Asp	1290	1295	
			1300
GAG GCA GGA CTT TTA AAG ATG GTG GAT GCT GCA GTT CAC GCT GAC TCA			4215
Glu Ala Gly Leu Leu Lys Met Val Asp Ala Ala Val His Ala Asp Ser	1305	1310	
			1315

GGG ACC ACA CTG CAG CTC ACC TCC TGT TTA AAT GGA AGT GGT CCT GTC 4263
Gly Thr Thr Leu Gln Leu Thr Ser Cys Leu Asn Gly Ser Gly Pro Val
1320 1325 1330

CCG GCT CCG CCC CCA ACT CCT GGA AAT CAC GAG AGA GGT GCT GCT TAGATTTTCA 4318
Pro Ala Pro Pro Thr Pro Gly Asn His Glu Arg Gly Ala Ala
1335 1340 1345

AGTGTGTGTC TTTCCACCAC CCGGAAGTAG CCACATTGGA TTTTCATTTT TGGAGGAGGG 4378

ACCTCAGACT GCAAGGAGCT TGTCCCTCAGG GCATTTCCAG AGAAGATGCC CATGACCCAA 4438

GAATGTGTG ACTCTACTCT CTTTTCATT CATTTAAAG TCCTATATAA TGTGCCCTGC 4498

TGTGGTCTCA CTACCAGTTA AAGCAAAAGA CTTTCAAACA CGTGGACTCT GTCCCTCCAAG 4558

AAGTGGCAAC GGCACCTCTG TGAACCTGGA TCGAATGGC AATGCTTTGT GTGTTGAGGA 4618

TGGGTGAGAT GTCCAGGGC CGAGTCTGTC TACCTTGGAG GCTTTGTGGA GGATGCGGCT 4678

ATGAGCCAAG TGTAAAGTGT GGGATGTGGA CTGGGAGGAA GGAAGGCGCA AGCCGTCCGG 4738

AGAGCGGTTG GAGCCTGCAG ATGCATTGTG CTGGCTCTGG TGGAGGTGGG CTTGTGGCCT 4798

GTCAGGAAAC GCAAAGGCGG CCGGCAGGGT TTGGTTTTGG AAGTTTGGG TGCTCTTCAC 4858

AGTCGGGTTA CAGGCGAGTT CCTGTGGCG TTTCCTACTC CTAATGAGAG TTCCTTCCGG 4918

ACTCTTACGT GTCTCCTGGC CTGGCCCCAG GAAGGAAATG ATGCAGCTTG CTCCTTCCTC 4978

ATCTCTCAGG CTGTGCCTTA ATTCAGAAACA CCAAAAGAGA GGAACGTCGG CAGAGGCTCC 5038

TGACGGGGCC GAAGAAATTGT GAGAACAGAA CAGAAACTCA GGGTTTCTGC TGGGTGGAGA 5098

CCCACGTGGC GCCCTGGTGG CAGGTCTGAG GGTTCCTCTGT CAAGTGGCGG TAAAGGCTCA 5158

GGCTGGTGTT CTTCTCTCTAT CTCCACTCCT GTCAGGCCCC CAAGTCCTCA GTATTTTAGC 5218

TTTGTGGCTT CCTGATGGCA GAAAAATCTT AATTGGTTGG TTGCTCTCC AGATAATCAC 5278
 TAGCCAGATT TCGAAATTAC TTTTITAGCCG AGGTTATGAT AACATCTACT GTATCCCTTTA 5338
 GAATTTTAAC CTATAAAACT ATGCTCTACTG GTTCTGCTCT GTGTGCTTAT GTTAAAAAAA 5398
 AAAAAAAA 5406

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1367 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Met Glu Ser Lys Gly Leu Leu Ala Val Ala Leu Trp Phe Cys Val Glu
 -19 -15 -10 -5
 Thr Arg Ala Ala Ser Val Gly Leu Pro Gly Asp Phe Leu His Pro Pro
 1 5 10
 Lys Leu Ser Thr Gln Lys Asp Ile Leu Thr Ile Leu Ala Asn Thr Thr
 15 20 25
 Leu Gln Ile Thr Cys Arg Gly Gln Arg Asp Leu Asp Trp Leu Trp Pro
 30 35 40 45
 Asn Ala Gln Arg Asp Ser Glu Glu Arg Val Leu Val Thr Glu Cys Gly
 50 55 60
 Gly Gly Asp Ser Ile Phe Cys Lys Thr Leu Thr Ile Pro Arg Val Val
 65 70 75

Gly Asn Asp Thr Gly Ala Tyr Lys Cys Ser Tyr Arg Asp Val Asp Ile
 80 85 90
 Ala Ser Thr Val Tyr Val Tyr Val Arg Asp Tyr Arg Ser Pro Phe Ile
 95 100 105
 Ala Ser Val Ser Asp Gln His Gly Ile Val Tyr Ile Thr Glu Asn Lys
 110 115 120 125
 Asn Lys Thr Val Val Ile Pro Cys Arg Gly Ser Ile Ser Asn Leu Asn
 130 135 140
 Val Ser Leu Cys Ala Arg Tyr Pro Glu Lys Arg Phe Val Pro Asp Gly
 145 150 155
 Asn Arg Ile Ser Trp Asp Ser Glu Ile Gly Phe Thr Leu Pro Ser Tyr
 160 165 170
 Met Ile Ser Tyr Ala Gly Met Val Phe Cys Glu Ala Lys Ile Asn Asp
 175 180 185
 Glu Thr Tyr Gln Ser Ile Met Tyr Ile Val Val Val Gly Tyr Arg
 190 195 200 205
 Ile Tyr Asp Val Ile Leu Ser Pro Pro His Glu Ile Glu Leu Ser Ala
 210 215 220
 Gly Glu Lys Leu Val Leu Asn Cys Thr Ala Arg Thr Glu Leu Asn Val
 225 230 235
 Gly Leu Asp Phe Thr Trp His Ser Pro Pro Ser Lys Ser His His Lys
 240 245 250
 Lys Ile Val Asn Arg Asp Val Lys Pro Phe Pro Gly Thr Val Ala Lys
 255 260 265
 Met Phe Leu Ser Thr Leu Thr Ile Glu Ser Val Thr Lys Ser Asp Gln
 270 275 280 285

Gly Glu Tyr Thr Cys Val Ala Ser Ser Gly Arg Met Ile Lys Arg Asn
 290 295 300
 Arg Thr Phe Val Arg Val His Thr Lys Pro Phe Ile Ala Phe Gly Ser
 305 310 315
 Gly Met Lys Ser Leu Val Glu Ala Thr Val Gly Ser Gln Val Arg Ile
 320 325 330
 Pro Val Lys Tyr Leu Ser Tyr Pro Ala Pro Asp Ile Lys Trp Tyr Arg
 335 340 345
 Asn Gly Arg Pro Ile Glu Ser Asn Tyr Thr Met Ile Val Gly Asp Glu
 350 355 360 365
 Leu Thr Ile Met Glu Val Thr Glu Arg Asp Ala Gly Asn Tyr Thr Val
 370 375 380
 Ile Leu Thr Asn Pro Ile Ser Met Glu Lys Gln Ser His Met Val Ser
 385 390 395
 Leu Val Val Asn Val Pro Pro Gln Ile Gly Glu Lys Ala Leu Ile Ser
 400 405 410
 Pro Met Asp Ser Tyr Gln Tyr Gly Thr Met Gln Thr Leu Thr Cys Thr
 415 420 425
 Val Tyr Ala Asn Pro Pro Leu His His Ile Gln Trp Tyr Trp Gln Leu
 430 435 440 445
 Glu Glu Ala Cys Ser Tyr Arg Pro Gly Gln Thr Ser Pro Tyr Ala Cys
 450 455 460
 Lys Glu Trp Arg His Val Glu Asp Phe Gln Gly Gly Asn Lys Ile Glu
 465 470 475
 Val Thr Lys Asn Gln Tyr Ala Leu Ile Glu Gly Lys Asn Lys Thr Val
 480 485 490

Ser Thr Leu Val Ile Gln Ala Ala Asn Val Ser Ala Leu Tyr Lys Cys
 495 500 505
 Glu Ala Ile Asn Lys Ala Gly Arg Gly Glu Arg Val Ile Ser Phe His
 510 515 520 525
 Val Ile Arg Gly Pro Glu Ile Thr Val Gln Pro Ala Ala Gln Pro Thr
 530 535 540
 Glu Gln Glu Ser Val Ser Leu Leu Cys Thr Ala Asp Arg Asn Thr Phe
 545 550 555
 Glu Asn Leu Thr Trp Tyr Lys Leu Gly Ser Gln Ala Thr Ser Val His
 560 565 570
 Met Gly Glu Ser Leu Thr Pro Val Cys Lys Asn Leu Asp Ala Leu Trp
 575 580 585
 Lys Leu Asn Gly Thr Met Phe Ser Ser Asn Ser Thr Asn Asp Ile Leu Ile
 590 595 600 605
 Val Ala Phe Gln Asn Ala Ser Leu Gln Asp Gln Gly Asp Tyr Val Cys
 610 615 620
 Ser Ala Gln Asp Lys Lys Thr Lys Lys Arg His Cys Leu Val Lys Gln
 625 630 635
 Leu Ile Ile Leu Glu Arg Met Ala Pro Met Ile Thr Gly Asn Leu Glu
 640 645 650
 Asn Gln Thr Thr Thr Ile Gly Glu Thr Ile Glu Val Thr Cys Pro Ala
 655 660 665
 Ser Gly Asn Pro Thr Pro His Ile Thr Thr Phe Lys Asp Asn Glu Thr
 670 675 680 685
 Leu Val Glu Asp Ser Gly Ile Val Leu Arg Asp Gly Asn Arg Asn Leu
 690 695 700

Thr Ile Arg Arg Val Arg Lys Glu Asp Gly Gly Leu Tyr Thr Cys Gln
 705 710 715
 Ala Cys Asn Val Leu Gly Cys Ala Arg Ala Glu Thr Leu Phe Ile Ile
 720 725 730
 Glu Gly Ala Gln Glu Lys Thr Asn Leu Glu Val Ile Ile Leu Val Gly
 735 740 745
 Thr Ala Val Ile Ala Met Phe Phe Trp Leu Leu Val Ile Leu Val
 750 755 760 765
 Arg Thr Val Lys Arg Ala Asn Glu Gly Glu Leu Lys Thr Gly Tyr Leu
 770 775 780
 Ser Ile Val Met Asp Pro Asp Glu Leu Pro Leu Asp Glu Arg Cys Glu
 785 790 795
 Arg Leu Pro Tyr Asp Ala Ser Lys Trp Glu Phe Pro Arg Asp Arg Leu
 800 805 810
 Lys Leu Gly Lys Pro Leu Gly Arg Gly Ala Phe Gly Gln Val Ile Glu
 815 820 825
 Ala Asp Ala Phe Gly Ile Asp Lys Thr Ala Thr Cys Lys Thr Val Ala
 830 835 840 845
 Val Lys Met Leu Lys Glu Gly Ala Thr His Ser Glu His Arg Ala Leu
 850 855 860
 Met Ser Glu Leu Lys Ile Leu Ile His Ile Gly His His Leu Asn Val
 865 870 875
 Val Asn Leu Leu Gly Ala Cys Thr Lys Pro Gly Gly Pro Leu Met Val
 880 885 890
 Ile Val Glu Phe Ser Lys Phe Gly Asn Leu Ser Thr Tyr Leu Arg Gly
 895 900 905

Lys Arg Asn Glu Phe Val Pro Tyr Lys Ser Lys Gly Ala Arg Phe Arg 925
 910 915
 Gln Gly Lys Asp Tyr Val Gly Glu Leu Ser Val Asp Leu Lys Arg Arg 940
 930 935
 Leu Asp Ser Ile Thr Ser Ser Gln Ser Ser Ala Ser Ser Gly Phe Val 955
 945 950
 Glu Glu Lys Ser Leu Ser Asp Val Glu Glu Glu Glu Ala Ser Glu Glu 970
 960 965
 Leu Tyr Lys Asp Phe Leu Thr Leu Glu His Leu Ile Cys Tyr Ser Phe 985
 975 980
 Gln Val Ala Lys Gly Met Glu Phe Leu Ala Ser Arg Lys Cys Ile His 1005
 990 995 1000
 Arg Asp Leu Ala Ala Arg Asn Ile Leu Leu Ser Glu Lys Asn Val Val 1020
 1010 1015
 Lys Ile Cys Asp Phe Gly Leu Ala Arg Asp Ile Tyr Lys Asp Pro Asp 1035
 1025 1030
 Tyr Val Arg Lys Gly Asp Ala Arg Leu Pro Leu Lys Trp Met Ala Pro 1050
 1040 1045
 Glu Thr Ile Phe Asp Arg Val Tyr Thr Ile Gln Ser Asp Val Trp Ser 1065
 1055 1060
 Phe Gly Val Leu Leu Trp Glu Ile Phe Ser Leu Gly Ala Ser Pro Tyr 1085
 1070 1075 1080
 Pro Gly Val Lys Ile Asp Glu Glu Phe Cys Arg Arg Leu Lys Glu Gly 1100
 1090 1095
 Thr Arg Met Arg Ala Pro Asp Tyr Thr Thr Pro Glu Met Tyr Gln Thr 1115
 1105 1110

Met Leu Asp Cys Trp His Glu Asp Pro Asn Gln Arg Pro Ser Phe Ser
1120 1125 1130
Glu Leu Val Glu His Leu Gly Asn Leu Leu Gln Ala Asn Ala Gln Gln
1135 1140 1145
Asp Gly Lys Asp Tyr Ile Val Leu Pro Met Ser Glu Thr Leu Ser Met
1150 1155 1160 1165
Glu Glu Asp Ser Gly Leu Ser Leu Pro Thr Ser Pro Val Ser Cys Met
1170 1175 1180
Glu Glu Glu Glu Val Cys Asp Pro Lys Phe His Tyr Asp Asn Thr Ala
1185 1190 1195
Gly Ile Ser His Tyr Leu Gln Asn Ser Lys Arg Lys Ser Arg Pro Val
1200 1205 1210
Ser Val Lys Thr Phe Glu Asp Ile Pro Leu Glu Glu Pro Glu Val Lys
1215 1220 1225
Val Ile Pro Asp Asp Ser Gln Thr Asp Ser Gly Met Val Leu Ala Ser
1230 1235 1240 1245
Glu Glu Leu Lys Thr Leu Glu Asp Arg Asn Lys Leu Ser Pro Ser Phe
1250 1255 1260
Gly Gly Met Met Pro Ser Lys Ser Arg Glu Ser Val Ala Ser Glu Gly
1265 1270 1275
Ser Asn Gln Thr Ser Gly Tyr Gln Ser Gly Tyr His Ser Asp Asp Thr
1280 1285 1290
Asp Thr Thr Val Tyr Ser Ser Asp Glu Ala Gly Leu Leu Lys Met Val
1295 1300 1305
Asp Ala Ala Val His Ala Asp Ser Gly Thr Thr Leu Gln Leu Thr Ser
1310 1315 1320 1325

Cys Leu Asn Gly Ser Gly Pro Val Pro Ala Pro Pro Pro Thr Pro Gly
1330 1335 1340

Asn His Glu Arg Gly Ala Ala
1345

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 96 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

AATTCGTCGA CTTTCTGTCA CCATGAGTGC ACTTCTGATC CTAGCCCTTG TGGGAGCTGC 60

TGTTGCTGAC TACAAAGATG ATGATGACAA GATCTA 96

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 96 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

AGCTTAGATC TTGTCATCAT CATCTTTGTA GTCAGCAACA GCAGCTCCCA CAGAGGCTAG 60

GATCAGAAGT GCACTCATGG TGACAGAAAG TCGACG 96

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 30 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

TGAGAAGATC TCAAACCAAG ACCTGCCTGT

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 34 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

CCAATGGCGG CCGCTCAGGA GATGTTGTCT TGGA

34

CLAIMS

What I claim is:

- 5 1. An isolated mammalian nucleic acid molecule encoding
 a receptor protein tyrosine kinase expressed in
 primitive hematopoietic cells and not expressed in
 mature hematopoietic cells.
- 10 2. A nucleic acid molecule according to claim 1 wherein
 the nucleic acid molecule is DNA.
3. A nucleic acid molecule according to claim 2 wherein
 the nucleic acid molecule is cDNA.
- 15 4. A nucleic acid molecule according to claim 1 wherein
 the nucleic acid molecule is RNA.
5. A nucleic acid molecule according to claim 1 that is
20 a mouse nucleic acid molecule.
6. A nucleic acid molecule according to claim 5 that is
 flk-2 comprising the sequence shown in Figure 1a.
- 25 7. A nucleic acid molecule according to claim 1 that is
 a human nucleic acid molecule.
8. A nucleic acid molecule according to claim 7 that is
 DNA.
- 30 9. A nucleic acid molecule according to claim 7 that is
 flk-2 comprising the sequence shown in Figure 1b.
- 35 10. An isolated acid nucleic molecule that is flk-2
 comprising the sequence shown in Figure 1a.
11. A nucleic acid molecule according to claim 10 wherein
 the nucleic acid molecule is DNA.
- 40 12. An isolated nucleic acid molecule that is flk-2

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comprising the sequence shown in Figure 1b.

13. A nucleic acid molecule according to claim 12 wherein the nucleic acid molecule is DNA.
- 5 14. An isolated nucleic molecule that is flk-1 having the sequence shown in Figure 2.
15. A nucleic acid molecule according to claim 14 wherein the nucleic acid molecule is DNA.
- 10 16. A nucleic acid molecule according to claim 14 wherein the nucleic acid molecule is cDNA.
- 15 17. A nucleic acid molecule according to claim 14 that has the corresponding sequence of RNA.
18. A vector comprising a mammalian nucleic acid molecule encoding a receptor protein tyrosine kinase expressed in primitive hematopoietic cells and not expressed in mature hematopoietic cells.
- 20 19. A vector comprising flk-1 having the nucleic acid sequence of Figure 2.
- 25 20. A vector comprising flk-2 having the nucleic acid sequence of Figure 1a or 1b.
21. A vector according to claim 18 wherein the vector is capable of being cloned in a host.
- 30 22. A vector according to claim 19 wherein the vector is capable of being cloned in a host.
23. A vector according to claim 20 wherein the vector is capable of being cloned in a host.
- 35 24. A vector according to claim 21 wherein the host is a prokaryotic host.

25. A vector according to claim 22 wherein the host is a prokaryotic host.
- 5 26. A vector according to claim 23 wherein the host is a prokaryotic host.
27. A vector according to claim 18 that is capable of expressing the nucleic acid molecule in a host.
- 10 28. A vector according to claim 19 that is capable of expressing flk-1 in a host.
29. A vector according to claim 20 that is capable of expressing flk-2 in a host.
- 15 30. A vector according to claim 27 wherein the host is a prokaryotic host.
31. A vector according to claim 28 wherein the host is a prokaryotic host.
- 20 32. A vector according to claim 29 wherein the host is a prokaryotic host.
- 25 33. A vector according to claim 27 wherein the host is a eucaryotic host.
34. A vector according to claim 28 wherein the host is a eucaryotic host.
- 30 35. A vector according to claim 29 wherein the host is a eucaryotic host.
- 35 36. An isolated protein tyrosine kinase expressed in primitive hematopoietic cells and not expressed in mature hematopoietic cells.
37. The protein tyrosine kinase according to claim 36 that is flk-2 having the sequence shown in Figure 1a or 1b.

SUBSTITUTE SHEET

38. The protein tyrosine kinase according to claim 36 that is human flk-2.
39. The protein tyrosine kinase according to claim 38 that is flk-2 having the sequence shown in Figure 1b.
40. An isolated protein tyrosine kinase that is flk-1 having the sequence shown in Figure 2.
41. A ligand that binds to a receptor protein tyrosine kinase expressed in primitive mammalian hematopoietic cells and not expressed in mature hematopoietic cells, wherein the ligand stimulates the proliferation and/or differentiation of the primitive hematopoietic cells.
42. A ligand that binds to the receptor protein tyrosine kinase having the amino acid sequence of flk-1 shown in Figure 2, wherein the ligand stimulates the proliferation and/or differentiation of cells that express flk-1.
43. A ligand that binds to the receptor protein tyrosine kinase having the amino acid sequence of flk-2 shown in Figure 1a or 1b, wherein the ligand stimulates the proliferation and/or differentiation of cells that express flk-2.
44. A nucleic acid molecule encoding a ligand that binds to a receptor protein tyrosine kinase expressed in primitive mammalian hematopoietic cells and not expressed in mature hematopoietic cells, wherein the ligand stimulates the proliferation and/or differentiation of the primitive hematopoietic cells.
45. A nucleic acid molecule encoding a ligand that binds to the receptor protein tyrosine kinase having the amino acid sequence of flk-1 shown in Figure 2, wherein the ligand stimulates the proliferation and/or differentiation of cells that express flk-1.

SUBSTITUTE SHEET

- 5 46. A nucleic acid molecule encoding a ligand that binds to the receptor protein tyrosine kinase having the amino acid sequence of flk-2 shown in Figure 1a or 1b, wherein the ligand stimulates the proliferation and/or differentiation of cells that express flk-2.
47. A nucleic acid molecule according to claim 44 wherein the nucleic acid molecule is DNA.
- 10 48. A nucleic acid molecule according to claim 44 wherein the nucleic acid molecule is cDNA.
49. A nucleic acid molecule according to claim 44 wherein the nucleic acid molecule is RNA.
- 15 50. A nucleic acid molecule according to claim 45 wherein the nucleic acid molecule is DNA.
51. A nucleic acid molecule according to claim 45 wherein the nucleic acid molecule is cDNA.
- 20 52. A nucleic acid molecule according to claim 45 wherein the nucleic acid molecule is RNA.
- 25 53. A nucleic acid molecule according to claim 46 wherein the nucleic acid molecule is DNA.
54. A nucleic acid molecule according to claim 46 wherein the nucleic acid molecule is cDNA.
- 30 55. A nucleic acid molecule according to claim 46 wherein the nucleic acid molecule is RNA.
- 35 56. A method of stimulating the proliferation and/or differentiation of primitive mammalian hematopoietic stem cells comprising contacting the stem cells with a ligand that binds to a receptor protein tyrosine kinase expressed in primitive mammalian hematopoietic cells and not expressed in mature hematopoietic cells.

57. A method of stimulating the proliferation and/or differentiation of primitive mammalian hematopoietic stem cells comprising contacting the stem cells with a ligand that binds to the receptor protein tyrosine kinase having the nucleic acid sequence of flk-1 shown in Figure 2.
58. A method of stimulating the proliferation and/or differentiation of primitive mammalian hematopoietic stem cells comprising contacting the stem cells with a ligand that binds to the receptor protein tyrosine kinase having the nucleic acid sequence of flk-2 shown in Figure 1a or 1b.
59. A method according to claim 56 wherein the stimulation occurs in vitro.
60. A method according to claim 57 wherein the stimulation occurs in vitro.
61. A method according to claim 58 wherein the stimulation occurs in vitro.
62. A method according to claim 56 wherein the stimulation occurs in vivo.
63. A method according to claim 57 wherein the stimulation occurs in vivo.
64. A method according to claim 58 wherein the stimulation occurs in vivo.
65. Murine cell line 2018 having ATCC accession number ATCC CRL 10907.
66. A recombinant nucleic acid molecule that is murine flk-2 having the sequence shown in Figure 1a.
67. A recombinant nucleic acid molecule comprising the

sequence shown in Figure 1a from nucleotide 1 to nucleotide 1662.

- 5 68. A recombinant nucleic acid molecule comprising the sequence shown in Figure 1a from nucleotide 31 to nucleotide 3006.
- 10 69. A recombinant nucleic acid molecule comprising the sequence shown in Figure 1a from nucleotide 112 to nucleotide 3006.
- 15 70. An isolated mRNA that encodes the murine flk-2 protein, said protein having the amino acid sequence shown in Figure 1a.
- 20 71. A recombinant nucleic acid molecule that is human flk-2 having the sequence shown in Figure 1b.
- 25 72. A recombinant nucleic acid molecule comprising the sequence shown in Figure 1b from nucleotide 1 to nucleotide 1689.
- 30 73. A recombinant nucleic acid molecule comprising the sequence shown in Figure 1b from nucleotide 58 to nucleotide 3036.
- 35 74. A recombinant nucleic acid molecule comprising the sequence shown in Figure 1b from nucleotide 139 to nucleotide 3036.
75. An isolated mRNA that encodes the human flk-2 protein, said protein having the amino acid sequence shown in Figure 1b.
76. A recombinant nucleic acid molecule that is murine flk-1 having the sequence shown in Figure 2.
77. A recombinant nucleic acid molecule comprising the sequence shown in Figure 2 from nucleotide 1 to

nucleotide 2493.

- 5 78. A recombinant nucleic acid molecule comprising the sequence shown in Figure 2 from nucleotide 208 to nucleotide 4308.
79. A recombinant nucleic acid molecule comprising the sequence shown in Figure 2 from nucleotide 265 to nucleotide 4308.
- 10 80. An isolated mRNA that encodes the murine flk-1 protein, said protein having the amino acid sequence shown in Figure 2.

1/19

Fig. 1a.1

GCGGCCTGGC TACCGCGCGC TCCGGAGGCC ATG CGG GCG TTG GCG CAG CGC AGC
 Met Arg Ala Leu Ala Gln Arg Ser
 -27 -25 -20

GAC CGG CGG CTG CTG CTG CTT GTT GTT TTG TCA GTA ATG ATT CTT GAG
 Asp Arg Arg Leu Leu Leu Leu Val Val Leu Ser Val Met Ile Leu Glu
 -15 -10 -5

ACC GTT ACA AAC CAA GAC CTG CCT GTG ATC AAG TGT GTT TTA ATC AGT
 Thr Val Thr Asn Gln Asp Leu Pro Val Ile Lys Cys Val Leu Ile Ser
 1 5 10

CAT GAG AAC AAT GGC TCA TCA GCG GGA AAG CCA TCA TCG TAC CGA ATG
 His Glu Asn Asn Gly Ser Ser Ala Gly Lys Pro Ser Ser Tyr Arg Met
 15 20 25

GTG CGA GGA TCC CCA GAA GAC CTC CAG TGT ACC CCG AGG CGC CAG AGT
 Val Arg Gly Ser Pro Glu Asp Leu Gln Cys Thr Pro Arg Arg Gln Ser
 30 35 40 45

GAA GGG ACG GTA TAT GAA GCG GCC ACC GTG GAG GTG GCC GAG TCT GGG
 Glu Gly Thr Val Tyr Glu Ala Ala Thr Val Glu Val Ala Glu Ser Gly
 50 55 60

TCC ATC ACC CTG CAA GTG CAG CTC GCC ACC CCA GGG GAC CTT TCC TGC
 Ser Ile Thr Leu Gln Val Gln Leu Ala Thr Pro Gly Asp Leu Ser Cys
 65 70 75

CTC TGG GTC TTT AAG CAC AGC TCC CTG GGC TGC CAG CCG CAC TTT GAT
 Leu Trp Val Phe Lys His Ser Ser Leu Gly Cys Gln Pro His Phe Asp
 80 85 90

TTA CAA AAC AGA GGA ATC GTT TCC ATG GCC ATC TTG AAC GTG ACA GAG
 Leu Gln Asn Arg Gly Ile Val Ser Met Ala Ile Leu Asn Val Thr Glu
 95 100 105

ACC CAG GCA GGA GAA TAC CTA CTC CAT ATT CAG AGC GAA CGC GCC AAC
 Thr Gln Ala Gly Glu Tyr Leu Leu His Ile Gln Ser Glu Arg Ala Asn
 110 115 120 125

TAC ACA GTA CTG TTC ACA GTG AAT GTA AGA GAT ACA CAG CTG TAT GTG
 Tyr Thr Val Leu Phe Thr Val Asn Val Arg Asp Thr Gln Leu Tyr Val
 130 135 140

CTA AGG AGA CCT TAC TTT AGG AAG ATG GAA AAC CAG GAT GCA CTG CTC
 Leu Arg Arg Pro Tyr Phe Arg Lys Met Glu Asn Gln Asp Ala Leu Leu
 145 150 155

TGC ATC TCC GAG GGT GTT CCG GAG CCC ACT GTG GAG TGG GTG CTC TGC
 Cys Ile Ser Glu Gly Val Pro Glu Pro Thr Val Glu Trp Val Leu Cys
 160 165 170

AGC TCC CAC AGG GAA AGC TGT AAA GAA GAA GGC CCT GCT GTT GTC AGA
 Ser Ser His Arg Glu Ser Cys Lys Glu Glu Gly Pro Ala Val Val Arg
 175 180 185

2/19

Fig. 1a.2

AAG Lys 190	GAG Glu	GAA Glu	AAG Lys	GTA Val	CTT Leu 195	CAT His	GAG Glu	TTG Leu	TTC Phe	GGA Gly 200	ACA Thr	GAC Asp	ATC Ile	AGA Arg	TGC Cys 205
TGT Cys	GCT Ala	AGA Arg	AAT Asn	GCA Ala 210	CTG Leu	GGC Gly	CGC Arg	GAA Glu	TGC Cys 215	ACC Thr	AAG Lys	CTG Leu	TTC Phe	ACC Thr 220	ATA Ile
GAT Asp	CTA Leu	AAC Asn 225	CAG Gln	GCT Ala	CCT Pro	CAG Gln	AGC Ser	ACA Thr 230	CTG Leu	CCC Pro	CAG Gln	TTA Leu	TTC Phe 235	CTG Leu	AAA Lys
GTG Val	GGG Gly 240	GAA Glu	CCC Pro	TTG Leu	TGG Trp	ATC Ile	AGG Arg 245	TGT Cys	AAG Lys	GCC Ala	ATC Ile	CAT His 250	GTG Val	AAC Asn	CAT His
GGA Gly 255	TTC Phe	GGG Gly	CTC Leu	ACC Thr	TGG Trp	GAG Glu 260	CTG Leu	GAA Glu	GAC Asp	AAA Lys 265	GCC Ala	CTG Leu	GAG Glu	GAG Glu	GGC Gly
AGC Ser 270	TAC Tyr	TTT Phe	GAG Glu	ATG Met	AGT Ser 275	ACC Thr	TAC Tyr	TCC Ser	ACA Thr	AAC Asn 280	AGG Arg	ACC Thr	ATG Met	ATT Ile 285	CGG Arg
ATT Ile	CTC Leu	TTG Leu	GCC Ala	TTT Phe 290	GTG Val	TCT Ser	TCC Ser	GTG Val	GGA Gly 295	AGG Arg	AAC Asn	GAC Asp	ACC Thr	GGA Gly 300	TAT Tyr
TAC Tyr	ACC Thr	TGC Cys	TCT Ser 305	TCC Ser	TCA Ser	AAG Lys	CAC His	CCC Pro 310	AGC Ser	CAG Gln	TCA Ser	GCG Ala	TTG Leu 315	GTG Val	ACC Thr
ATC Ile 320	CTA Leu	GAA Glu	AAA Lys	GGG Gly	TTT Phe	ATA Ile	AAC Asn 325	GCT Ala	ACC Thr	AGC Ser	TCG Ser	CAA Gln 330	GAA Glu	GAG Glu	TAT Tyr
GAA Glu 335	ATT Ile	GAC Asp	CCG Pro	TAC Tyr	GAA Glu 340	AAG Lys	TTC Phe	TGC Cys	TTC Phe	TCA Ser	GTC Val 345	AGG Arg	TTT Phe	AAA Lys	GCG Ala
TAC Tyr 350	CCA Pro	CGA Arg	ATC Ile	CGA Arg	TGC Cys 355	ACG Thr	TGG Trp	ATC Ile	TTC Phe 360	TCT Ser	CAA Gln	GCC Ala	TCA Ser	TTT Phe 365	CCT Pro
TGT Cys	GAA Glu	CAG Gln	AGA Arg	GGC Gly 370	CTG Leu	GAG Glu	GAT Asp	GGG Gly 375	TAC Tyr	AGC Ser	ATA Ile	TCT Ser	AAA Lys	TTT Phe 380	TGC Cys
GAT Asp	CAT His	AAG Lys 385	AAC Asn	AAG Lys	CCA Pro	GGA Gly	GAG Glu	TAC Tyr 390	ATA Ile	TTC Phe	TAT Tyr	GCA Ala	GAA Glu 395	AAT Asn	GAT Asp
GAC Asp	GCC Ala	CAG Gln	TTC Phe	ACC Thr	AAA Lys	ATG Met	TTC Phe 400	ACG Thr	CTG Leu 405	AAT Asn	ATA Ile	AGA Arg 410	AAG Lys	AAA Lys	CCT Pro

3/19

Fig. 1a.3

CAA	GTG	CTA	GCA	AAT	GCC	TCA	GCC	AGC	CAG	GCG	TCC	TGT	TCC	TCT	GAT	
Gln	Val	Leu	Ala	Asn	Ala	Ser	Ala	Ser	Gln	Ala	Ser	Cys	Ser	Ser	Asp	
	415					420					425					
GGC	TAC	CCG	CTA	CCC	TCT	TGG	ACC	TGG	AAG	AAG	TGT	TCG	GAC	AAA	TCT	
Gly	Tyr	Pro	Leu	Pro	Ser	Trp	Thr	Trp	Lys	Lys	Cys	Ser	Asp	Lys	Ser	
	430				435					440					445	
CCC	AAT	TGC	ACG	GAG	GAA	ATC	CCA	GAA	GGA	GTT	TGG	AAT	AAA	AAG	GCT	
Pro	Asn	Cys	Thr	Glu	Glu	Ile	Pro	Glu	Gly	Val	Trp	Asn	Lys	Lys	Ala	
				450					455					460		
AAC	AGA	AAA	GTG	TTT	GGC	CAG	TGG	GTG	TCG	AGC	AGT	ACT	CTA	AAT	ATG	
Asn	Arg	Lys	Val	Phe	Gly	Gln	Trp	Val	Ser	Ser	Ser	Thr	Leu	Asn	Met	
			465				470						475			
AGT	GAG	GCC	GGG	AAA	GGG	CTT	CTG	GTC	AAA	TGC	TGT	GCG	TAC	AAT	TCT	
Ser	Glu	Ala	Gly	Lys	Gly	Leu	Leu	Val	Lys	Cys	Cys	Ala	Tyr	Asn	Ser	
	480					485						490				
ATG	GGC	ACG	TCT	TGC	GAA	ACC	ATC	TTT	TTA	AAC	TCA	CCA	GGC	CCC	TTC	
Met	Gly	Thr	Ser	Cys	Glu	Thr	Ile	Phe	Leu	Asn	Ser	Pro	Gly	Pro	Phe	
	495					500					505					
CCT	TTC	ATC	CAA	GAC	AAC	ATC	TCC	TTC	TAT	GCG	ACC	ATT	GGG	CTC	TGT	
Pro	Phe	Ile	Gln	Asp	Asn	Ile	Ser	Phe	Tyr	Ala	Thr	Ile	Gly	Leu	Cys	
	510				515					520					525	
CTC	CCC	TTC	ATT	GTT	GTT	CTC	ATT	GTG	TTG	ATC	TGC	CAC	AAA	TAC	AAA	
Leu	Pro	Phe	Ile	Val	Val	Leu	Ile	Val	Leu	Ile	Cys	His	Lys	Tyr	Lys	
				530				535						540		
AAG	CAA	TTT	AGG	TAC	GAG	AGT	CAG	CTG	CAG	ATG	ATC	CAG	GTG	ACT	GGC	
Lys	Gln	Phe	Arg	Tyr	Glu	Ser	Gln	Leu	Gln	Met	Ile	Gln	Val	Thr	Gly	
			545				550						555			
CCC	CTG	GAT	AAC	GAG	TAC	TTC	TAC	GTT	GAC	TTC	AGG	GAC	TAT	GAA	TAT	
Pro	Leu	Asp	Asn	Glu	Tyr	Phe	Tyr	Val	Asp	Phe	Arg	Asp	Tyr	Glu	Tyr	
		560					565					570				
GAC	CTT	AAG	TGG	GAG	TTC	CCG	AGA	GAG	AAC	TTA	GAG	TTT	GGG	AAG	GTC	
Asp	Leu	Lys	Trp	Glu	Phe	Pro	Arg	Glu	Asn	Leu	Glu	Phe	Gly	Lys	Val	
	575					580					585					
CTG	GGG	TCT	GGC	GCT	TTC	GGG	AGG	GTG	ATG	AAC	GCC	ACG	GCC	TAT	GGC	
Leu	Gly	Ser	Gly	Ala	Phe	Gly	Arg	Val	Met	Asn	Ala	Thr	Ala	Tyr	Gly	
	590				595					600					605	
ATT	AGT	AAA	ACG	GGA	GTC	TCA	ATT	CAG	GTG	GCG	GTG	AAG	ATG	CTA	AAA	
Ile	Ser	Lys	Thr	Gly	Val	Ser	Ile	Gln	Val	Ala	Val	Lys	Met	Leu	Lys	
				610					615					620		
GAG	AAA	GCT	GAC	AGC	TGT	GAA	AAA	GAA	GCT	CTC	ATG	TCG	GAG	CTC	AAA	
Glu	Lys	Ala	Asp	Ser	Cys	Glu	Lys	Glu	Ala	Leu	Met	Ser	Glu	Leu	Lys	
			625					630					635			

4/19

Fig. 1a.4

ATG Met	ATG Met	ACC Thr	CAC His	CTG Leu	GGA Gly	CAC His	CAT His	GAC Asp	AAC Asn	ATC Ile	GTG Val	AAT Asn	CTG Leu	CTG Leu	GGG Gly
		640					645					650			
GCA Ala	TGC Cys	ACA Thr	CTG Leu	TCA Ser	GGG Gly	CCA Pro	GTG Val	TAC Tyr	TTG Leu	ATT Ile	TTT Phe	GAA Glu	TAT Tyr	TGT Cys	TGC Cys
		655				660					665				
TAT Tyr	GGT Gly	GAC Asp	CTC Leu	CTC Leu	AAC Asn	TAC Tyr	CTA Leu	AGA Arg	AGT Ser	AAA Lys	AGA Arg	GAG Glu	AAG Lys	TTT Phe	CAC His
					675					680					685
AGG Arg	ACA Thr	TGG Trp	ACA Thr	GAG Glu	ATT Ile	TTT Phe	AAG Lys	GAA Glu	CAT His	AAT Asn	TTC Phe	AGT Ser	TCT Ser	TAC Tyr	CCT Pro
				690					695					700	
ACT Thr	TTC Phe	CAG Gln	GCA Ala	CAT His	TCA Ser	AAT Asn	TCC Ser	AGC Ser	ATG Met	CCT Pro	GGT Gly	TCA Ser	CGA Arg	GAA Glu	GTT Val
			705					710					715		
CAG Gln	TTA Leu	CAC His	CCG Pro	CCC Pro	TTG Leu	GAT Asp	CAG Gln	CTC Leu	TCA Ser	GGG Gly	TTC Phe	AAT Asn	GGG Gly	AAT Asn	TCA Ser
		720					725					730			
ATT Ile	CAT His	TCT Ser	GAA Glu	GAT Asp	GAG Glu	ATT Ile	GAA Glu	TAT Tyr	GAA Glu	AAC Asn	CAG Gln	AAG Lys	AGG Arg	CTG Leu	GCA Ala
		735				740					745				
GAA Glu	GAA Glu	GAG Glu	GAG Glu	GAA Glu	GAT Asp	TTG Leu	AAC Asn	GTG Val	CTG Leu	ACG Thr	TTT Phe	GAA Glu	GAC Asp	CTC Leu	CTT Leu
					755					760					765
TGC Cys	TTT Phe	GCG Ala	TAC Tyr	CAA Gln	GTG Val	GCC Ala	AAA Lys	GGC Gly	ATG Met	GAA Glu	TTC Phe	CTG Leu	GAG Glu	TTC Phe	AAG Lys
			770						775					780	
TCG Ser	TGT Cys	GTC Val	CAC His	AGA Arg	GAC Asp	CTG Leu	GCA Ala	GCC Ala	AGG Arg	AAT Asn	GTG Val	TTG Leu	GTC Val	ACC Thr	CAC His
			785					790					795		
GGG Gly	AAG Lys	GTG Val	GTG Val	AAG Lys	ATC Ile	TGT Cys	GAC Asp	TTT Phe	GGA Gly	CTG Leu	GCC Ala	CGA Arg	GAC Asp	ATC Ile	CTG Leu
		800					805					810			
AGC Ser	GAC Asp	TCC Ser	AGC Ser	TAC Tyr	GTC Val	GTC Val	AGG Arg	GGC Gly	AAC Asn	GCA Ala	CGG Arg	CTG Leu	CCG Pro	GTG Val	AAG Lys
		815				820					825				
TGG Trp	ATG Met	GCA Ala	CCC Pro	GAG Glu	AGC Ser	TTA Leu	TTT Phe	GAA Glu	GGG Gly	ATC Ile	TAC Tyr	ACA Thr	ATC Ile	AAG Lys	AGT Ser
				835						840					845
GAC Asp	GTC Val	TGG Trp	TCC Ser	TAC Tyr	GGC Gly	ATC Ile	CTT Leu	CTC Leu	TGG Trp	GAG Glu	ATA Ile	TTT Phe	TCA Ser	CTG Leu	GGT Gly
				850					855					860	

5/19

Fig. 1a.5

GTG AAC CCT TAC CCT GGC ATT CCT GTC GAC GCT AAC TTC TAT AAA CTG
 Val Asn Pro Tyr Pro Gly Ile Pro Val Asp Ala Asn Phe Tyr Lys Leu
 865 870 875

ATT CAG AGT GGA TTT AAA ATG GAG CAG CCA TTC TAT GCC ACA GAA GGG
 Ile Gln Ser Gly Phe Lys Met Glu Gln Pro Phe Tyr Ala Thr Glu Gly
 880 885 890

ATA TAC TTT GTA ATG CAA TCC TGC TGG GCT TTT GAC TCA AGG AAG CGG
 Ile Tyr Phe Val Met Gln Ser Cys Trp Ala Phe Asp Ser Arg Lys Arg
 895 900 905

CCA TCC TTC CCC AAC CTG ACT TCA TTT TTA GGA TGT CAG CTG GCA GAG
 Pro Ser Phe Pro Asn Leu Thr Ser Phe Leu Gly Cys Gln Leu Ala Glu
 910 915 920 925

GCA GAA GAA GCA TGT ATC AGA ACA TCC ATC CAT CTA CCA AAA CAG GCG
 Ala Glu Glu Ala Cys Ile Arg Thr Ser Ile His Leu Pro Lys Gln Ala
 930 935 940

GCC CCT CAG CAG AGA GGC GGG CTC AGA GCC CAG TCG CCA CAG CGC CAG
 Ala Pro Gln Gln Arg Gly Gly Leu Arg Ala Gln Ser Pro Gln Arg Gln
 945 950 955

GTG AAG ATT CAC AGA GAA AGA AGT TAGCGAGGAG GCCTTGGACC CCGCCACCCCT
 Val Lys Ile His Arg Glu Arg Ser
 960 965

AGCAGGCTGT AGACCGCAGA GCCAAGATTA GCCTCGCCTC TGAGGAAGCG CCCTACAGCG
 CGTTGCTTCG CTGGACTTTT CTCTAGATGC TGTCTGCCAT TACTCCAAAG TGA CTCTCTAT
 AAAATCAAAC CTCTCCTCGC ACAGGCGGGA GAGCCAATAA TGAGACTTGT TGGTGAGCCC
 GCCTACCCTG GGGGCCTTTC CACGAGCTTG AGGGGAAAGC CATGTATCTG AAATATAGTA
 TATTCTTGTA AATACGTGAA ACAAACCAAA CCCGTTTTTTT GCTAAGGGAA AGCTAAATAT
 GATTTTTTAAA AATCTATGTT TTAAAATACT ATGTAACTTT TTCATCTATT TAGTGATATA
 TTTTATGGAT GGAAATAAAC TTTCTACTGT AAAAAAAAAA AAAAAAAAAA AAAAAA

6/19

Fig. 1b.1

CGAGGCGGCA TCCGAGGGCT GGGCCGGCGC CCTGGGGGAC CCCGGGCTCC GGAGGCC

ATG CCG GCG TTG GCG CGC GAC GCG GGC ACC GTG CCG CTG CTC GTT GTT
Met Pro Ala Leu Ala Arg Asp Ala Gly Thr Val Pro Leu Leu Val Val
-27 -25 -20 -15

TTT TCT GCA ATG ATA TTT GGG ACT ATT ACA AAT CAA GAT CTG CCT GTG
Phe Ser Ala Met Ile Phe Gly Thr Ile Thr Asn Gln Asp Leu Pro Val
-10 -5 1 5

ATC AAG TGT GTT TTA ATC AAT CAT AAG AAC AAT GAT TCA TCA GTG GGG
Ile Lys Cys Val Leu Ile Asn His Lys Asn Asn Asp Ser Ser Val Gly
10 15 20

AAG TCA TCA TCA TAT CCC ATG GTA TCA GAA TCC CCG GAA GAC CTC GGG
Lys Ser Ser Ser Tyr Pro Met Val Ser Glu Ser Pro Glu Asp Leu Gly
25 30 35

TGT GCG TTG AGA CCC CAG AGC TCA GGG ACA GTG TAC GAA GCT GCC GCT
Cys Ala Leu Arg Pro Gln Ser Ser Gly Thr Val Tyr Glu Ala Ala Ala
40 45 50

GTG GAA GTG GAT GTA TCT GCT TCC ATC ACA CTG CAA GTG CTG GTC GAT
Val Glu Val Asp Val Ser Ala Ser Ile Thr Leu Gln Val Leu Val Asp
55 60 65

GCC CCA GGG AAC ATT TCC TGT CTC TGG GTC TTT AAG CAC AGC TCC CTG
Ala Pro Gly Asn Ile Ser Cys Leu Trp Val Phe Lys His Ser Ser Leu
70 75 80 85

AAT TGC CAG CCA CAT TTT GAT TTA CAA AAC AGA GGA GTT GTT TCC ATG
Asn Cys Gln Pro His Phe Asp Leu Gln Asn Arg Gly Val Val Ser Met
90 95 100

GTC ATT TTG AAA ATG ACA GAA ACC CAA GCT GGA GAA TAC CTA CTT TTT
Val Ile Leu Lys Met Thr Glu Thr Gln Ala Gly Glu Tyr Leu Leu Phe
105 110 115

ATT CAG AGT GAA GCT ACC AAT TAC ACA ATA TTG TTT ACA GTG AGT ATA
Ile Gln Ser Glu Ala Thr Asn Tyr Thr Ile Leu Phe Thr Val Ser Ile
120 125 130

AGA AAT ACC CTG CTT TAC ACA TTA AGA AGA CCT TAC TTT AGA AAA ATG
Arg Asn Thr Leu Leu Tyr Thr Leu Arg Arg Pro Tyr Phe Arg Lys Met
135 140 145

GAA AAC CAG GAC GCC CTG GTC TGC ATA TCT GAG AGC GTT CCA GAG CCG
Glu Asn Gln Asp Ala Leu Val Cys Ile Ser Glu Ser Val Pro Glu Pro
150 155 160 165

ATC GTG GAA TGG GTG CTT TGC GAT TCA CAG GGG GAA AGC TGT AAA GAA
Ile Val Glu Trp Val Leu Cys Asp Ser Gln Gly Glu Ser Cys Lys Glu
170 175 180

GAA AGT CCA GCT GTT GTT AAA AAG GAG GAA AAA GTG CTT CAT GAA TTA
Glu Ser Pro Ala Val Val Lys Lys Glu Glu Lys Val Leu His Glu Leu
185 190 195

Fig. 1b.2

TTT Phe	GGG Gly	ACG Thr 200	GAC Asp	ATA Ile	AGG Arg	TGC Cys	TGT Cys 205	GCC Ala	AGA Arg	AAT Asn	GAA Glu	CTG Leu 210	GGC Gly	AGG Arg	GAA Glu
TGC Cys	ACC Thr 215	AGG Arg	CTG Leu	TTC Phe	ACA Thr	ATA Ile 220	GAT Asp	CTA Leu	AAT Asn	CAA Gln	ACT Thr 225	CCT Pro	CAG Gln	ACC Thr	ACA Thr
TTG Leu 230	CCA Pro	CAA Gln	TTA Leu	TTT Phe	CTT Leu 235	AAA Lys	GTA Val	GGG Gly	GAA Glu	CCC Pro 240	TTA Leu	TGG Trp	ATA Ile	AGG Arg	TGC Cys 245
AAA Lys	GCT Ala	GTT Val	CAT His	GTG Val 250	AAC Asn	CAT His	GGA Gly	TTC Phe	GGG Gly 255	CTC Leu	ACC Thr	TGG Trp	GAA Glu	TTA Leu 260	GAA Glu
AAC Asn	AAA Lys	GCA Ala	CTC Leu 265	GAG Glu	GAG Glu	GGC Gly	AAC Asn	TAC Tyr 270	TTT Phe	GAG Glu	ATG Met	AGT Ser	ACC Thr 275	TAT Tyr	TCA Ser
ACA Thr	AAC Asn 280	AGA Arg	ACT Thr	ATG Met	ATA Ile	CGG Arg	ATT Ile 285	CTG Leu	TTT Phe	GCT Ala	TTT Phe	GTA Val 290	TCA Ser	TCA Ser	GTG Val
GCA Ala 295	AGA Arg	AAC Asn	GAC Asp	ACC Thr	GGA Gly	TAC Tyr 300	TAC Tyr	ACT Thr	TGT Cys	TCC Ser	TCT Ser 305	TCA Ser	AAG Lys	CAT His	CCC Pro
AGT Ser 310	CAA Gln	TCA Ser	GCT Ala	TTG Leu	GTT Val 315	ACC Thr	ATC Ile	GTA Val	GGA Gly	AAG Lys 320	GGA Gly	TTT Phe	ATA Ile	AAT Asn	GCT Ala 325
ACC Thr	AAT Asn	TCA Ser	AGT Ser	GAA Glu 330	GAT Asp	TAT Tyr	GAA Glu	ATT Ile	GAC Asp 335	CAA Gln	TAT Tyr	GAA Glu	GAG Glu	TTT Phe 340	TGT Cys
TTT Phe	TCT Ser	GTC Val	AGG Arg 345	TTT Phe	AAA Lys	GCC Ala	TAC Tyr	CCA Pro 350	CAA Gln	ATC Ile	AGA Arg	TGT Cys 355	ACG Thr	TGG Trp	ACC Thr
TTC Phe	TCT Ser	CGA Arg 360	AAA Lys	TCA Ser	TTT Phe	CCT Pro	TGT Cys 365	GAG Glu	CAA Gln	AAG Lys	GGT Gly	CTT Leu 370	GAT Asp	AAC Asn	GGA Gly
TAC Tyr 375	AGC Ser	ATA Ile	TCC Ser	AAG Lys	TTT Phe	TGC Cys 380	AAT Asn	CAT His	AAG Lys	CAC His	CAG Gln 385	CCA Pro	GGA Gly	GAA Glu	TAT Tyr
ATA Ile 390	TTC Ph	CAT His	GCA Ala	GAA Glu	AAT Asn 395	GAT Asp	GAT Asp	GCC Ala	CAA Gln	TTT Phe 400	ACC Thr	AAA Lys	ATG Met	TTC Phe	ACG Thr 405
CTG Leu	AAT Asn	ATA Ile	AGA Arg	AGG Arg 410	AAA Lys	CCT Pro	CAA Gln	GTG Val	CTC Leu 415	GCA Ala	GAA Glu	GCA Ala	TCG Ser	GCA Ala	AGT Ser 420

8/19

Fig. 1b.3

CAG	GCG	TCC	TGT	TTC	TCG	GAT	GGA	TAC	CCA	TTA	CCA	TCT	TGG	ACC	TGG		
Gln	Ala	Ser	Cys	Phe	Ser	Asp	Gly	Tyr	Pro	Leu	Pro	Ser	Trp	Thr	Trp		
			425					430					435				
AAG	AAG	TGT	TCA	GAC	AAG	TCT	CCC	AAC	TGC	ACA	GAA	GAG	ATC	ACA	GAA		
Lys	Lys	Cys	Ser	Asp	Lys	Ser	Pro	Asn	Cys	Thr	Glu	Glu	Ile	Thr	Glu		
		440					445					450					
GGA	GTC	TGG	AAT	AGA	AAG	GCT	AAC	AGA	AAA	GTG	TTT	GGA	CAG	TGG	GTG		
Gly	Val	Trp	Asn	Arg	Lys	Ala	Asn	Arg	Lys	Val	Phe	Gly	Gln	Trp	Val		
	455					460					465						
TCG	AGC	AGT	ACT	CTA	AAC	ATG	AGT	GAA	GCC	ATA	AAA	GGG	TTC	CTG	GTC		
Ser	Ser	Ser	Thr	Leu	Asn	Met	Ser	Glu	Ala	Ile	Lys	Gly	Phe	Leu	Val		
470					475					480					485		
AAG	TGC	TGT	GCA	TAC	AAT	TCC	CTT	GGC	ACA	TCT	TGT	GAG	ACG	ATC	CTT		
Lys	Cys	Cys	Ala	Tyr	Asn	Ser	Leu	Gly	Thr	Ser	Cys	Glu	Thr	Ile	Leu		
			490					495						500			
TTA	AAC	TCT	CCA	GGC	CCC	TTC	CCT	TTC	ATC	CAA	GAC	AAC	ATC	TCA	TTC		
Leu	Asn	Ser	Pro	Gly	Pro	Phe	Pro	Phe	Ile	Gln	Asp	Asn	Ile	Ser	Phe		
			505					510					515				
TAT	GCA	ACA	ATT	GGT	GTT	TGT	CTC	CTC	TTC	ATT	GTC	GTT	TTA	ACC	CTG		
Tyr	Ala	Thr	Ile	Gly	Val	Cys	Leu	Leu	Phe	Ile	Val	Val	Leu	Thr	Leu		
	520						525					530					
CTA	ATT	TGT	CAC	AAG	TAC	AAA	AAG	CAA	TTT	AGG	TAT	GAA	AGC	CAG	CTA		
Leu	Ile	Cys	His	Lys	Tyr	Lys	Lys	Gln	Phe	Arg	Tyr	Glu	Ser	Gln	Leu		
	535					540					545						
CAG	ATG	GTA	CAG	GTG	ACC	GGC	TCC	TCA	GAT	AAT	GAG	TAC	TTC	TAC	GTT		
Gln	Met	Val	Gln	Val	Thr	Gly	Ser	Ser	Asp	Asn	Glu	Tyr	Phe	Tyr	Val		
550					555					560					565		
GAT	TTC	AGA	GAA	TAT	GAA	TAT	GAT	CTC	AAA	TGG	GAG	TTT	CCA	AGA	GAA		
Asp	Phe	Arg	Glu	Tyr	Glu	Tyr	Asp	Leu	Lys	Trp	Glu	Phe	Pro	Arg	Glu		
			570						575					580			
AAT	TTA	GAG	TTT	GGG	AAG	GTA	CTA	GGA	TCA	GGT	GCT	TTT	GGA	AAA	GTG		
Asn	Leu	Glu	Phe	Gly	Lys	Val	Leu	Gly	Ser	Gly	Ala	Phe	Gly	Lys	Val		
			585					590					595				
ATG	AAC	GCA	ACA	GCT	TAT	GGA	ATT	AGC	AAA	ACA	GGA	GTC	TCA	ATC	CAG		
Met	Asn	Ala	Thr	Ala	Tyr	Gly	Ile	Ser	Lys	Thr	Gly	Val	Ser	Ile	Gln		
	600					605						610					
GTT	GCC	GTC	AAA	ATG	CTG	AAA	GAA	AAA	GCA	GAC	AGC	TCT	GAA	AGA	GAG		
Val	Ala	Val	Lys	Met	Leu	Lys	Glu	Lys	Ala	Asp	Ser	Ser	Glu	Arg	Glu		
	615					620					625						
GCA	CTC	ATG	TCA	GAA	CTC	AAG	ATG	ATG	ACC	CAG	CTG	GGA	AGC	CAC	GAG		
Ala	Leu	Met	Ser	Glu	Leu	Lys	Met	Met	Thr	Gln	Leu	Gly	Ser	His	Glu		
630					635					640					645		

9/19

Fig. 1b.4

AAT ATT GTG AAC CTG CTG GGG GCG TGC ACA CTG TCA GGA CCA ATT TAC
 Asn Ile Val Asn Leu Leu Gly Ala Cys Thr Leu Ser Gly Pro Ile Tyr
 650 655 660

TTG ATT TTT GAA TAC TGT TGC TAT GGT GAT CTT CTC AAC TAT CTA AGA
 Leu Ile Phe Glu Tyr Cys Cys Tyr Gly Asp Leu Leu Asn Tyr Leu Arg
 665 670 675

AGT AAA AGA GAA AAA TTT CAC AGG ACT TGG ACA GAG ATT TTC AAG GAA
 Ser Lys Arg Glu Lys Phe His Arg Thr Trp Thr Glu Ile Phe Lys Glu
 680 685 690

CAC AAT TTC AGT TTT TAC CCC ACT TTC CAA TCA CAT CCA AAT TCC AGC
 His Asn Phe Ser Phe Tyr Pro Thr Phe Gln Ser His Pro Asn Ser Ser
 695 700 705

ATG CCT GGT TCA AGA GAA GTT CAG ATA CAC CCG GAC TCG GAT CAA ATC
 Met Pro Gly Ser Arg Glu Val Gln Ile His Pro Asp Ser Asp Gln Ile
 710 715 720 725

TCA GGG CTT CAT GGG AAT TCA TTT CAC TCT GAA GAT GAA ATT GAA TAT
 Ser Gly Leu His Gly Asn Ser Phe His Ser Glu Asp Glu Ile Glu Tyr
 730 735 740

GAA AAC CAA AAA AGG CTG GAA GAA GAG GAG GAC TTG AAT GTG CTT ACA
 Glu Asn Gln Lys Arg Leu Glu Glu Glu Glu Asp Leu Asn Val Leu Thr
 745 750 755

TTT GAA GAT CTT CTT TGC TTT GCA TAT CAA GTT GCC AAA GGA ATG GAA
 Phe Glu Asp Leu Leu Cys Phe Ala Tyr Gln Val Ala Lys Gly Met Glu
 760 765 770

TTT CTG GAA TTT AAG TCG TGT GTT CAC AGA GAC CTG GCC GCC AGG AAC
 Phe Leu Glu Phe Lys Ser Cys Val His Arg Asp Leu Ala Ala Arg Asn
 775 780 785

GTG CTT GTC ACC CAC GGG AAA GTG GTG AAG ATA TGT GAC TTT GGA TTG
 Val Leu Val Thr His Gly Lys Val Val Lys Ile Cys Asp Phe Gly Leu
 790 795 800 805

GCT CGA GAT ATC ATG AGT GAT TCC AAC TAT GTT GTC AGG GGC AAT GCC
 Ala Arg Asp Ile Met Ser Asp Ser Asn Tyr Val Val Arg Gly Asn Ala
 810 815 820

CGT CTG CCT GTA AAA TGG ATG GCC CCC GAA AGC CTG TTT GAA GGC ATC
 Arg Leu Pro Val Lys Trp Met Ala Pro Glu Ser Leu Phe Glu Gly Ile
 825 830 835

TAC ACC ATT AAG AGT GAT GTC TGG TCA TAT GGA ATA TTA CTG TGG GAA
 Tyr Thr Ile Lys Ser Asp Val Trp Ser Tyr Gly Ile Leu Leu Trp Glu
 840 845 850

ATC TTC TCA CTT GGT GTG AAT CCT TAC CCT GGC ATT CCG GTT GAT GCT
 Ile Phe Ser Leu Gly Val Asn Pro Tyr Pro Gly Ile Pro Val Asp Ala
 855 860 865

10/19

Fig. 1b.5

AAC TTC TAC AAA CTG ATT CAA AAT GGA TTT AAA ATG GAT CAG CCA TTT
 Asn Phe Tyr Lys Leu Ile Gln Asn Gly Phe Lys Met Asp Gln Pro Phe
 870 875 880 885

TAT GCT ACA GAA GAA ATA TAC ATT ATA ATG CAA TCC TGC TGG GCT TTT
 Tyr Ala Thr Glu Glu Ile Tyr Ile Ile Met Gln Ser Cys Trp Ala Phe
 890 895 900

GAC TCA AGG AAA CGG CCA TCC TTC CCT AAT TTG ACT TCG TTT TTA GGA
 Asp Ser Arg Lys Arg Pro Ser Phe Pro Asn Leu Thr Ser Phe Leu Gly
 905 910 915

TGT CAG CTG GCA GAT GCA GAA GAA GCG ATG TAT CAG AAT GTG GAT GGC
 Cys Gln Leu Ala Asp Ala Glu Glu Ala Met Tyr Gln Asn Val Asp Gly
 920 925 930

CGT GTT TCG GAA TGT CCT CAC ACC TAC CAA AAC AGG CGA CCT TTC AGC
 Arg Val Ser Glu Cys Pro His Thr Tyr Gln Asn Arg Arg Pro Phe Ser
 935 940 945

AGA GAG ATG GAT TTG GGG CTA CTC TCT CCG CAG GCT CAG GTC GAA GAT
 Arg Glu Met Asp Leu Gly Leu Leu Ser Pro Gln Ala Gln Val Glu Asp
 950 955 960 965

TCG TAGAGGAACA ATTTAGTTTT AAGGACTTCA TCCCTCCACC TATCCCTAAC
 Ser

AGGCTGTAGA TTACCAAAAC AAGATTAATT TCATCACTAA AAGAAAATCT ATTATCAACT
 GCTGCTTCAC CAGACTTTTC TCTAGAAGCC GTCTGCGTTT ACTCTTGTTT TCAAAGGGAC
 TTTTGTAATA TCAAATCATC CTGTCACAAG GCAGGAGGAG CTGATAATGA ACTTTATTGG
 AGCATTGATC TGCATCCAAG GCCTTCTCAG GCCGGCTTGA GTGAATTGTG TACCTGAAGT
 ACAGTATATT CTTGTAAATA CATAAAACAA AAGCATTTTG CTAAGGAGAA GCTAATATGA
 TTTTTTAAGT CTATGTTTTA AAATAATATG TAAATTTTTC AGCTATTTAG TGATATATTT
 TATGGGTGGG AATAAAATTT CTACTACAGA AAAAAAAAAA AAAAAAAAAA AAAAA

11/19

Fig. 2.1

CTGTGTCCCG CAGCCGGATA ACCTGGCTGA CCCGATTCCG CGGACACCCG TGCAGCCGCG
 GCTGGAGCCA GGGCGCCGGT GCCCGCGCTC TCCCCGGTCT TGCCTGCGG GGGCCGATAC
 CGCCTCTGTG ACTTCTTTGC GGGCCAGGGA CGGAGAAGGA GTCTGTGCCT GAGAACTGG
 GCTCTGTGCC CAGGCGCGAG GTGCAGG ATG GAG AGC AAG GGC CTG CTA GCT
 Met Glu Ser Lys Gly Leu Leu Ala
 -19 -15

GTC GCT CTG TGG TTC TGC GTG GAG ACC CGA GCC GCC TCT GTG GGT TTG
 Val Ala Leu Trp Phe Cys Val Glu Thr Arg Ala Ala Ser Val Gly Leu
 -10 -5 1 5

CCT GGC GAT TTT CTC CAT CCC CCC AAG CTC AGC ACA CAG AAA GAC ATA
 Pro Gly Asp Phe Leu His Pro Pro Lys Leu Ser Thr Gln Lys Asp Ile
 10 15 20

CTG ACA ATT TTG GCA AAT ACA ACC CTT CAG ATT ACT TGC AGG GGA CAG
 Leu Thr Ile Leu Ala Asn Thr Thr Leu Gln Ile Thr Cys Arg Gly Gln
 25 30 35

CGG GAC CTG GAC TGG CTT TGG CCC AAT GCT CAG CGT GAT TCT GAG GAA
 Arg Asp Leu Asp Trp Leu Trp Pro Asn Ala Gln Arg Asp Ser Glu Glu
 40 45 50

AGG GTA TTG GTG ACT GAA TGC GGC GGT GGT GAC AGT ATC TTC TGC AAA
 Arg Val Leu Val Thr Glu Cys Gly Gly Gly Asp Ser Ile Phe Cys Lys
 55 60 65

ACA CTC ACC ATT CCC AGG GTG GTT GGA AAT GAT ACT GGA GCC TAC AAG
 Thr Leu Thr Ile Pro Arg Val Val Gly Asn Asp Thr Gly Ala Tyr Lys
 70 75 80 85

TGC TCG TAC CGG GAC GTC GAC ATA GCC TCC ACT GTT TAT GTC TAT GTT
 Cys Ser Tyr Arg Asp Val Asp Ile Ala Ser Thr Val Tyr Val Tyr Val
 90 95 100

CGA GAT TAC AGA TCA CCA TTC ATC GCC TCT GTC AGT GAC CAG CAT GGC
 Arg Asp Tyr Arg Ser Pro Phe Ile Ala Ser Val Ser Asp Gln His Gly
 105 110 115

ATC GTG TAC ATC ACC GAG AAC AAG AAC AAA ACT GTG GTG ATC CCC TGC
 Ile Val Tyr Ile Thr Glu Asn Lys Asn Lys Thr Val Val Ile Pro Cys
 120 125 130

CGA GGG TCG ATT TCA AAC CTC AAT GTG TCT CTT TGC GCT AGG TAT CCA
 Arg Gly Ser Ile Ser Asn Leu Asn Val Ser Leu Cys Ala Arg Tyr Pro
 135 140 145

GAA AAG AGA TTT GTT CCG GAT GGA AAC AGA ATT TCC TGG GAC AGC GAG
 Glu Lys Arg Phe Val Pro Asp Gly Asn Arg Ile Ser Trp Asp Ser Glu
 150 155 160 165

ATA GGC TTT ACT CTC CCC AGT TAC ATG ATC AGC TAT GCC GGC ATG GTC
 Ile Gly Phe Thr Leu Pro Ser Tyr Met Ile Ser Tyr Ala Gly Met Val
 170 175 180

Fig. 2.2

TTC Phe	TGT Cys	GAG Glu	GCA Ala 185	AAG Lys	ATC Ile	AAT Asn	GAT Asp	GAA Glu 190	ACC Thr	TAT Tyr	CAG Gln	TCT Ser	ATC Ile 195	ATG Met	TAC Tyr	
ATA Ile	GTT Val 200	GTG Val	GTT Val	GTA Val	GGA Gly	TAT Tyr	AGG Arg 205	ATT Ile	TAT Tyr	GAT Asp	GTG Val	ATT Ile 210	CTG Leu	AGC Ser	CCC Pro	
CCG Pro	CAT His 215	GAA Glu	ATT Ile	GAG Glu	CTA Leu	TCT Ser 220	GCC Ala	GGA Gly	GAA Glu	AAA Lys	CTT Leu 225	GTC Val	TTA Leu	AAT Asn	TGT Cys	
ACA Thr 230	GCG Ala	AGA Arg	ACA Thr	GAG Glu	CTC Leu 235	AAT Asn	GTG Val	GGG Gly	CTT Leu	GAT Asp 240	TTC Phe	ACC Thr	TGG Trp	CAC His	TCT Ser 245	
CCA Pro	CCT Pro	TCA Ser	AAG Lys	TCT Ser 250	CAT His	CAT His	AAG Lys	AAG Lys	ATT Ile 255	GTA Val	AAC Asn	CGG Arg	GAT Asp	GTG Val 260	AAA Lys	
CCC Pro	TTT Phe	CCT Pro	GGG Gly 265	ACT Thr	GTG Val	GCG Ala	AAG Lys	ATG Met 270	TTT Phe	TTG Leu	AGC Ser	ACC Thr 275	TTG Leu	ACA Thr	ATA Ile	
GAA Glu	AGT Ser	GTG Val 280	ACC Thr	AAG Lys	AGT Ser	GAC Asp	CAA Gln 285	GGG Gly	GAA Glu	TAC Tyr	ACC Thr	TGT Cys 290	GTA Val	GCG Ala	TCC Ser	
AGT Ser	GGA Gly 295	CGG Arg	ATG Met	ATC Ile	AAG Lys	AGA Arg 300	AAT Asn	AGA Arg	ACA Thr	TTT Phe	GTC Val 305	CGA Arg	GTT Val	CAC His	ACA Thr	
AAG Lys 310	CCT Pro	TTT Phe	ATT Ile	GCT Ala	TTC Phe 315	GGT Gly	AGT Ser	GGG Gly	ATG Met	AAA Lys 320	TCT Ser	TTG Leu	GTG Val	GAA Glu	GCC Ala 325	
ACA Thr	GTG Val	GGC Gly	AGT Ser	CAA Gln 330	GTC Val	CGA Arg	ATC Ile	CCT Pro	GTG Val 335	AAG Lys	TAT Tyr	CTC Leu	AGT Ser	TAC Tyr 340	CCA Pro	
GCT Ala	CCT Pro	GAT Asp	ATC Ile 345	AAA Lys	TGG Trp	TAC Tyr	AGA Arg	AAT Asn 350	GGA Gly	AGG Arg	CCC Pro	ATT Ile 355	GAG Glu	TCC Ser	AAC Asn	
TAC Tyr	ACA Thr	ATG Met 360	ATT Ile	GTT Val	GGC Gly	GAT Asp	GAA Glu 365	CTC Leu	ACC Thr	ATC Ile	ATG Met	GAA Glu 370	GTG Val	ACT Thr	GAA Glu	
AGA Arg	GAT Asp 375	GCA Ala	GGA Gly	AAC Asn	TAC Tyr	ACG Thr 380	GTC Val	ATC Ile	CTC Leu	ACC Thr	AAC Asn 385	CCC Pro	ATT Ile	TCA Ser	ATG Met	
GAG Glu 390	AAA Lys	CAG Gln	AGC Ser	CAC His	ATG Met 395	GTC Val	TCT Ser	CTG Leu	GTT Val	GTG Val 400	AAT Asn	GTC Val	CCA Pro	CCC Pro	CAG Gln 405	

13 / 19

Fig. 2.3

ATC	GGT	GAG	AAA	GCC	TTG	ATC	TCG	CCT	ATG	GAT	TCC	TAC	CAG	TAT	GGG	
Ile	Gly	Glu	Lys	Ala	Leu	Ile	Ser	Pro	Met	Asp	Ser	Tyr	Gln	Tyr	Gly	
				410					415					420		
ACC	ATG	CAG	ACA	TTG	ACA	TGC	ACA	GTC	TAC	GCC	AAC	CCT	CCC	CTG	CAC	
Thr	Met	Gln	Thr	Leu	Thr	Cys	Thr	Val	Tyr	Ala	Asn	Pro	Pro	Leu	His	
			425					430					435			
CAC	ATC	CAG	TGG	TAC	TGG	CAG	CTA	GAA	GAA	GCC	TGC	TCC	TAC	AGA	CCC	
His	Ile	Gln	Trp	Tyr	Trp	Gln	Leu	Glu	Glu	Ala	Cys	Ser	Tyr	Arg	Pro	
		440					445					450				
GGC	CAA	ACA	AGC	CCG	TAT	GCT	TGT	AAA	GAA	TGG	AGA	CAC	GTG	GAG	GAT	
Gly	Gln	Thr	Ser	Pro	Tyr	Ala	Cys	Lys	Glu	Trp	Arg	His	Val	Glu	Asp	
	455					460					465					
TTC	CAG	GGG	GGA	AAC	AAG	ATC	GAA	GTC	ACC	AAA	AAC	CAA	TAT	GCC	CTG	
Phe	Gln	Gly	Gly	Asn	Lys	Ile	Glu	Val	Thr	Lys	Asn	Gln	Tyr	Ala	Leu	
470					475					480					485	
ATT	GAA	GGA	AAA	AAC	AAA	ACT	GTA	AGT	ACG	CTG	GTC	ATC	CAA	GCT	GCC	
Ile	Glu	Gly	Lys	Asn	Lys	Thr	Val	Ser	Thr	Leu	Val	Ile	Gln	Ala	Ala	
			490						495					500		
AAC	GTG	TCA	GCG	TTG	TAC	AAA	TGT	GAA	GCC	ATC	AAC	AAA	GCG	GGA	CGA	
Asn	Val	Ser	Ala	Leu	Tyr	Lys	Cys	Glu	Ala	Ile	Asn	Lys	Ala	Gly	Arg	
			505					510					515			
GGA	GAG	AGG	GTC	ATC	TCC	TTC	CAT	GTG	ATC	AGG	GGT	CCT	GAA	ATT	ACT	
Gly	Glu	Arg	Val	Ile	Ser	Phe	His	Val	Ile	Arg	Gly	Pro	Glu	Ile	Thr	
		520					525					530				
GTG	CAA	CCT	GCT	GCC	CAG	CCA	ACT	GAG	CAG	GAG	AGT	GTG	TCC	CTG	TTG	
Val	Gln	Pro	Ala	Ala	Gln	Pro	Thr	Glu	Gln	Glu	Ser	Val	Ser	Leu	Leu	
	535					540					545					
TGC	ACT	GCA	GAC	AGA	AAT	ACG	TTT	GAG	AAC	CTC	ACG	TGG	TAC	AAG	CTT	
Cys	Thr	Ala	Asp	Arg	Asn	Thr	Phe	Glu	Asn	Leu	Thr	Trp	Tyr	Lys	Leu	
550					555					560					565	
GGC	TCA	CAG	GCA	ACA	TCG	GTC	CAC	ATG	GGC	GAA	TCA	CTC	ACA	CCA	GTT	
Gly	Ser	Gln	Ala	Thr	Ser	Val	His	Met	Gly	Glu	Ser	Leu	Thr	Pro	Val	
			570						575					580		
TGC	AAG	AAC	TTG	GAT	GCT	CTT	TGG	AAA	CTG	AAT	GGC	ACC	ATG	TTT	TCT	
Cys	Lys	Asn	Leu	Asp	Ala	Leu	Trp	Lys	Leu	Asn	Gly	Thr	Met	Phe	Ser	
			585					590					595			
AAC	AGC	ACA	AAT	GAC	ATC	TTG	ATT	GTG	GCA	TTT	CAG	AAT	GCC	TCT	CTG	
Asn	S r	Thr	Asn	Asp	Ile	Leu	Ile	Val	Ala	Phe	Gln	Asn	Ala	Ser	Leu	
			600				605					610				
CAG	GAC	CAA	GGC	GAC	TAT	GTT	TGC	TCT	GCT	CAA	GAT	AAG	AAG	ACC	AAG	
Gln	Asp	Gln	Gly	Asp	Tyr	Val	Cys	Ser	Ala	Gln	Asp	Lys	Lys	Thr	Lys	
	615					620					625					

Fig. 2.4

AAA	AGA	CAT	TGC	CTG	GTC	AAA	CAG	CTC	ATC	ATC	CTA	GAG	CGC	ATG	GCA	630	635	640	645
Lys	Arg	His	Cys	Leu	Val	Lys	Gln	Leu	Ile	Ile	Leu	Glu	Arg	Met	Ala				
CCC	ATG	ATC	ACC	GGA	AAT	CTG	GAG	AAT	CAG	ACA	ACA	ACC	ATT	GGC	GAG	650	655	660	
Pro	Met	Ile	Thr	Gly	Asn	Leu	Glu	Asn	Gln	Thr	Thr	Thr	Ile	Gly	Glu				
ACC	ATT	GAA	GTG	ACT	TGC	CCA	GCA	TCT	GGA	AAT	CCT	ACC	CCA	CAC	ATT	665	670	675	
Thr	Ile	Glu	Val	Thr	Cys	Pro	Ala	Ser	Gly	Asn	Pro	Thr	Pro	His	Ile				
ACA	TGG	TTC	AAA	GAC	AAC	GAG	ACC	CTG	GTA	GAA	GAT	TCA	GGC	ATT	GTA	680	685	690	
Thr	Trp	Phe	Lys	Asp	Asn	Glu	Thr	Leu	Val	Glu	Asp	Ser	Gly	Ile	Val				
CTG	AGA	GAT	GGG	AAC	CGG	AAC	CTG	ACT	ATC	CGC	AGG	GTG	AGG	AAG	GAG	695	700	705	
Leu	Arg	Asp	Gly	Asn	Arg	Asn	Leu	Thr	Ile	Arg	Arg	Val	Arg	Lys	Glu				
GAT	GGA	GGC	CTC	TAC	ACC	TGC	CAG	GCC	TGC	AAT	GTC	CTT	GGC	TGT	GCA	710	715	720	725
Asp	Gly	Gly	Leu	Tyr	Thr	Cys	Gln	Ala	Cys	Asn	Val	Leu	Gly	Cys	Ala				
AGA	GCG	GAG	ACG	CTC	TTC	ATA	ATA	GAA	GGT	GCC	CAG	GAA	AAG	ACC	AAC	730	735	740	
Arg	Ala	Glu	Thr	Leu	Phe	Ile	Ile	Glu	Gly	Ala	Gln	Glu	Lys	Thr	Asn				
TTG	GAA	GTC	ATT	ATC	CTC	GTC	GGC	ACT	GCA	GTG	ATT	GCC	ATG	TTC	TTC	745	750	755	
Leu	Glu	Val	Ile	Ile	Leu	Val	Gly	Thr	Ala	Val	Ile	Ala	Met	Phe	Phe				
TGG	CTC	CTT	CTT	GTC	ATT	CTC	GTA	CGG	ACC	GTT	AAG	CGG	GCC	AAT	GAA	760	765	770	
Trp	Leu	Leu	Leu	Val	Ile	Leu	Val	Arg	Thr	Val	Lys	Arg	Ala	Asn	Glu				
GGG	GAA	CTG	AAG	ACA	GGC	TAC	TTG	TCT	ATT	GTC	ATG	GAT	CCA	GAT	GAA	775	780	785	
Gly	Glu	Leu	Lys	Thr	Gly	Tyr	Leu	Ser	Ile	Val	Met	Asp	Pro	Asp	Glu				
TTG	CCC	TTG	GAT	GAG	CGC	TGT	GAA	CGC	TTG	CCT	TAT	GAT	GCC	AGC	AAG	790	795	800	805
Leu	Pro	Leu	Asp	Glu	Arg	Cys	Glu	Arg	Leu	Pro	Tyr	Asp	Ala	Ser	Lys				
TGG	GAA	TTC	CCC	AGG	GAC	CGG	CTG	AAA	CTA	GGA	AAA	CCT	CTT	GGC	CGC	810	815	820	
Trp	Glu	Phe	Pro	Arg	Asp	Arg	Leu	Lys	Leu	Gly	Lys	Pro	Leu	Gly	Arg				
GGT	GCC	TTC	GGC	CAA	GTG	ATT	GAG	GCA	GAC	GCT	TTT	GGA	ATT	GAC	AAG	825	830	835	
Gly	Ala	Phe	Gly	Gln	Val	Ile	Glu	Ala	Asp	Ala	Phe	Gly	Ile	Asp	Lys				
ACA	GCG	ACT	TGC	AAA	ACA	GTA	GCC	GTC	AAG	ATG	TTG	AAA	GAA	GGA	GCA	840	845	850	
Thr	Ala	Thr	Cys	Lys	Thr	Val	Ala	Val	Lys	Met	Leu	Lys	Glu	Gly	Ala				

15/19
Fig. 2.5

ACA	CAC	AGC	GAG	CAT	CGA	GCC	CTC	ATG	TCT	GAA	CTC	AAG	ATC	CTC	ATC		
Thr	His	Ser	Glu	His	Arg	Ala	Leu	Met	Ser	Glu	Leu	Lys	Ile	Leu	Ile		
	855					860					865						
CAC	ATT	GGT	CAC	CAT	CTC	AAT	GTG	GTG	AAC	CTC	CTA	GGC	GCC	TGC	ACC		
His	Ile	Gly	His	His	Leu	Asn	Val	Val	Asn	Leu	Leu	Gly	Ala	Cys	Thr		
	870				875				880						885		
AAG	CCG	GGA	GGG	CCT	CTC	ATG	GTG	ATT	GTG	GAA	TTC	TCG	AAG	TTT	GGA		
Lys	Pro	Gly	Gly	Pro	Leu	Met	Val	Ile	Val	Glu	Phe	Ser	Lys	Phe	Gly		
				890					895						900		
AAC	CTA	TCA	ACT	TAC	TTA	CGG	GGC	AAG	AGA	AAT	GAA	TTT	GTT	CCC	TAT		
Asn	Leu	Ser	Thr	Tyr	Leu	Arg	Gly	Lys	Arg	Asn	Glu	Phe	Val	Pro	Tyr		
			905					910					915				
AAG	AGC	AAA	GGG	GCA	CGC	TTC	CGC	CAG	GGC	AAG	GAC	TAC	GTT	GGG	GAG		
Lys	Ser	Lys	Gly	Ala	Arg	Phe	Arg	Gln	Gly	Lys	Asp	Tyr	Val	Gly	Glu		
		920					925					930					
CTC	TCC	GTG	GAT	CTG	AAA	AGA	CGC	TTG	GAC	AGC	ATC	ACC	AGC	AGC	CAG		
Leu	Ser	Val	Asp	Leu	Lys	Arg	Arg	Leu	Asp	Ser	Ile	Thr	Ser	Ser	Gln		
						940					945						
AGC	TCT	GCC	AGC	TCA	GGC	TTT	GTT	GAG	GAG	AAA	TCG	CTC	AGT	GAT	GTA		
Ser	Ser	Ala	Ser	Ser	Gly	Phe	Val	Glu	Glu	Lys	Ser	Leu	Ser	Asp	Val		
					955					960					965		
GAG	GAA	GAA	GAA	GCT	TCT	GAA	GAA	CTG	TAC	AAG	GAC	TTC	CTG	ACC	TTG		
Glu	Glu	Glu	Glu	Ala	Ser	Glu	Glu	Leu	Tyr	Lys	Asp	Phe	Leu	Thr	Leu		
				970				975							980		
GAG	CAT	CTC	ATC	TGT	TAC	AGC	TTC	CAA	GTG	GCT	AAG	GGC	ATG	GAG	TTC		
Glu	His	Leu	Ile	Cys	Tyr	Ser	Phe	Gln	Val	Ala	Lys	Gly	Met	Glu	Phe		
			985					990					995				
TTG	GCA	TCA	AGG	AAG	TGT	ATC	CAC	AGG	GAC	CTG	GCA	GCA	CGA	AAC	ATT		
Leu	Ala	Ser	Arg	Lys	Cys	Ile	His	Arg	Asp	Leu	Ala	Ala	Arg	Asn	Ile		
			1000				1005						1010				
CTC	CTA	TCG	GAG	AAG	AAT	GTG	GTT	AAG	ATC	TGT	GAC	TTC	GGC	TTG	GCC		
Leu	Leu	Ser	Glu	Lys	Asn	Val	Val	Lys	Ile	Cys	Asp	Phe	Gly	Leu	Ala		
						1020					1025						
CGG	GAC	ATT	TAT	AAA	GAC	CCG	GAT	TAT	GTC	AGA	AAA	GGA	GAT	GCC	CGA		
Arg	Asp	Ile	Tyr	Lys	Asp	Pro	Asp	Tyr	Val	Arg	Lys	Gly	Asp	Ala	Arg		
					1035				1040						1045		
CTC	CCT	TTG	AAG	TGG	ATG	GCC	CCG	GAA	ACC	ATT	TTT	GAC	AGA	GTA	TAC		
Leu	Pro	Leu	Lys	Trp	Met	Ala	Pro	Glu	Thr	Ile	Phe	Asp	Arg	Val	Tyr		
				1050					1055						1060		
ACA	ATT	CAG	AGC	GAT	GTG	TGG	TCT	TTC	GGT	GTG	TTG	CTC	TGG	GAA	ATA		
Thr	Ile	Gln	Ser	Asp	Val	Trp	Ser	Phe	Gly	Val	Leu	Leu	Trp	Glu	Ile		
			1065					1070					1075				

16/19

Fig. 2.6

TTT TCC TTA GGT GCC TCC CCA TAC CCT GGG GTC AAG ATT GAT GAA GAA
 Phe Ser Leu Gly Ala Ser Pro Tyr Pro Gly Val Lys Ile Asp Glu Glu
 1080 1085 1090

TTT TGT AGG AGA TTG AAA GAA GGA ACT AGA ATG CGG GCT CCT GAC TAC
 Phe Cys Arg Arg Leu Lys Glu Gly Thr Arg Met Arg Ala Pro Asp Tyr
 1095 1100 1105

ACT ACC CCA GAA ATG TAC CAG ACC ATG CTG GAC TGC TGG CAT GAG GAC
 Thr Thr Pro Glu Met Tyr Gln Thr Met Leu Asp Cys Trp His Glu Asp
 1110 1115 1120 1125

CCC AAC CAG AGA CCC TCG TTT TCA GAG TTG GTG GAG CAT TTG GGA AAC
 Pro Asn Gln Arg Pro Ser Phe Ser Glu Leu Val Glu His Leu Gly Asn
 1130 1135 1140

CTC CTG CAA GCA AAT GCG CAG CAG GAT GGC AAA GAC TAT ATT GTT CTT
 Leu Leu Gln Ala Asn Ala Gln Gln Asp Gly Lys Asp Tyr Ile Val Leu
 1145 1150 1155

CCA ATG TCA GAG ACA CTG AGC ATG GAA GAG GAT TCT GGA CTC TCC CTG
 Pro Met Ser Glu Thr Leu Ser Met Glu Glu Asp Ser Gly Leu Ser Leu
 1160 1165 1170

CCT ACC TCA CCT GTT TCC TGT ATG GAG GAA GAG GAA GTG TGC GAC CCC
 Pro Thr Ser Pro Val Ser Cys Met Glu Glu Glu Glu Val Cys Asp Pro
 1175 1180 1185

AAA TTC CAT TAT GAC AAC ACA GCA GGA ATC AGT CAT TAT CTC CAG AAC
 Lys Phe His Tyr Asp Asn Thr Ala Gly Ile Ser His Tyr Leu Gln Asn
 1190 1195 1200 1205

AGT AAG CGA AAG AGC CGG CCA GTG AGT GTA AAA ACA TTT GAA GAT ATC
 Ser Lys Arg Lys Ser Arg Pro Val Ser Val Lys Thr Phe Glu Asp Ile
 1210 1215 1220

CCA TTG GAG GAA CCA GAA GTA AAA GTG ATC CCA GAT GAC AGC CAG ACA
 Pro Leu Glu Glu Pro Glu Val Lys Val Ile Pro Asp Asp Ser Gln Thr
 1225 1230 1235

GAC AGT GGG ATG GTC CTT GCA TCA GAA GAG CTG AAA ACT CTG GAA GAC
 Asp Ser Gly Met Val Leu Ala Ser Glu Glu Leu Lys Thr Leu Glu Asp
 1240 1245 1250

AGG AAC AAA TTA TCT CCA TCT TTT GGT GGA ATG ATG CCC AGT AAA AGC
 Arg Asn Lys Leu Ser Pro Ser Phe Gly Gly Met Met Pro Ser Lys Ser
 1255 1260 1265

AGG GAG TCT GTG GCC TCG GAA GGC TCC AAC CAG ACC AGT GGC TAC CAG
 Arg Glu Ser Val Ala Ser Glu Gly S r Asn Gln Thr Ser Gly Tyr Gln
 1270 1275 1280 1285

TCT GGG TAT CAC TCA GAT GAC ACA GAC ACC ACC GTG TAC TCC AGC GAC
 Ser Gly Tyr His Ser Asp Asp Thr Asp Thr Thr Val Tyr Ser Ser Asp
 1290 1295 1300

17/19

Fig. 2.7

GAG GCA GGA CTT TTA AAG ATG GTG GAT GCT GCA GTT CAC GCT GAC TCA
Glu Ala Gly Leu Leu Lys Met Val Asp Ala Ala Val His Ala Asp Ser
1305 1310 1315

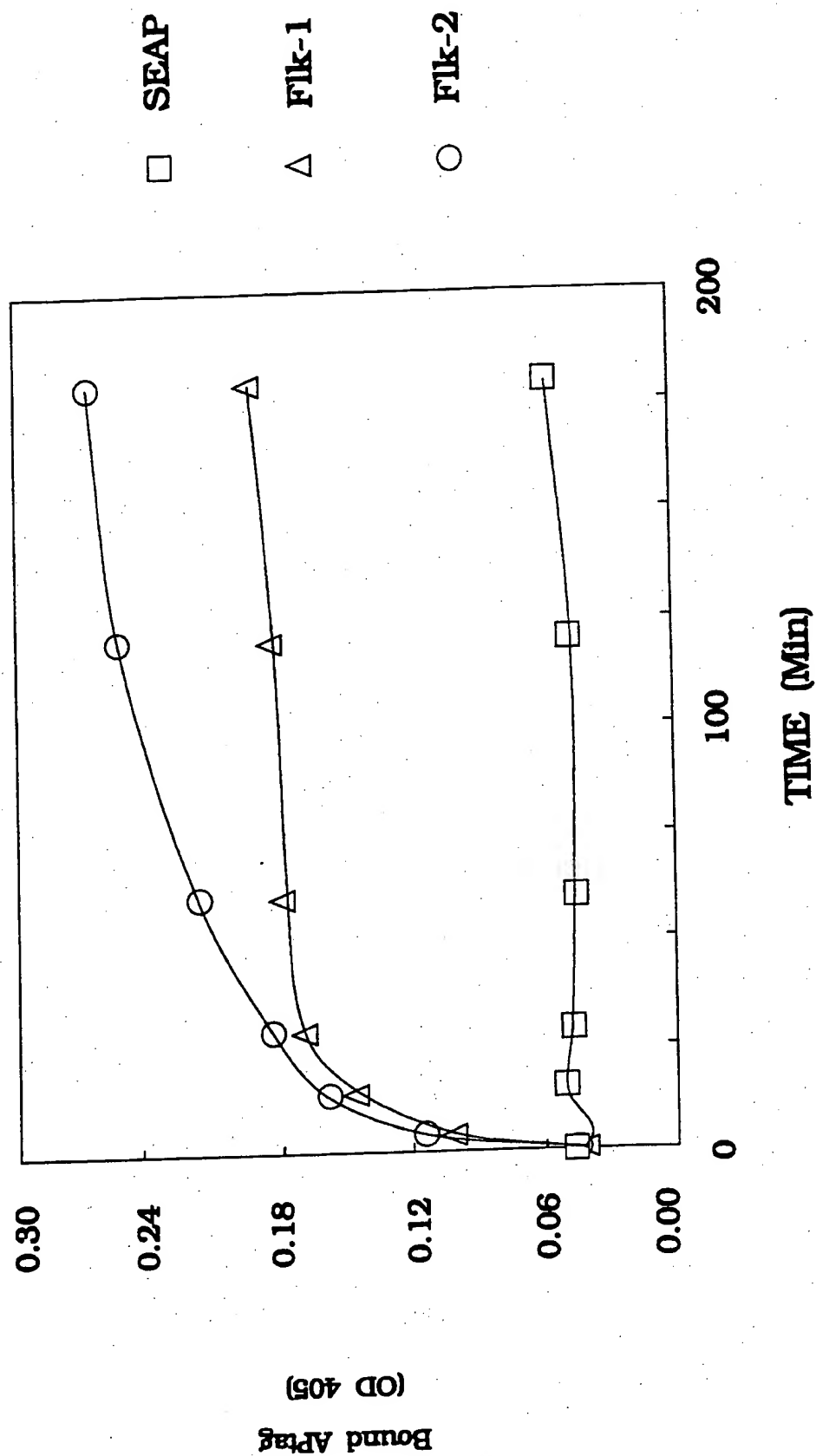
GGG ACC ACA CTG CAG CTC ACC TCC TGT TTA AAT GGA AGT GGT CCT GTC
Gly Thr Thr Leu Gln Leu Thr Ser Cys Leu Asn Gly Ser Gly Pro Val
1320 1325 1330

CCG GCT CCG CCC CCA ACT CCT GGA AAT CAC GAG AGA GGT GCT GCT TAG
Pro Ala Pro Pro Pro Thr Pro Gly Asn His Glu Arg Gly Ala Ala
1335 1340 1345

ATTTTCAAGT GTTGTTCTTT CCACCACCCG GAAGTAGCCA CATTTGATTT TCATTTTTTG
AGGAGGGACC TCAGACTGCA AGGAGCTTGT CCTCAGGGCA TTTCCAGAGA AGATGCCCCAT
GACCCAAGAA TGTGTTGACT CTACTCTCTT TTCCATTTCAT TTAAAAGTCC TATATAATGT
GCCCTGCTGT GGTCTCACTA CCAGTTAAAG CAAAAGACTT TCAAACACGT GGACTCTGTC
CTCCAAGAAG TGGCAACGGC ACCTCTGTGA AACTGGATCG AATGGGCAAT GCTTTGTGTG
TTGAGGATGG GTGAGATGTC CCAGGGCCGA GTCTGTCTAC CTTGGAGGCT TTGTGGAGGA
TGCGGCTATG AGCCAAGTGT TAAGTGTGGG ATGTGGACTG GGAGGAAGGA AGGCGCAAGC
CGTCCGGAGA GCGGTTGGAG CCTGCAGATG CATGTGTGCTG GCTCTGGTGG AGGTGGGCTT
GTGGCCTGTC AGGAAACGCA AAGGCGGCCG GCAGGGTTTG GTTTTGGAAG GTTTGCGTGC
TCTTCACAGT CGGGTTACAG GCGAGTTCCC TGTGGCGTTT CCTACTCCTA ATGAGAGTTC
CTTCCGGACT CTTACGTGTC TCCTGGCCTG GCCCCAGGAA GGAAATGATG CAGCTTGCTC
CTTCCTCATC TCTCAGGCTG TGCCTTAATT CAGAACACCA AAAGAGAGGA ACGTCGGCAG
AGGCTCCTGA CGGGGCCGAA GAATTGTGAG AACAGAACAG AAATCAGGG TTTCTGCTGG
GTGGAGACCC ACGTGGCGCC CTGGTGGCAG GTCTGAGGGT TCTCTGTCAA GTGGCGGTAA
AGGCTCAGGC TGGTGTCTT CCTCTATCTC CACTCCTGTC AGGCCCCCAA GTCCTCAGTA
TTTLAGCTTT GTGGCTTCCT GATGGCAGAA AAATCTTAAT TGGTTGGTTT GCTCTCCAGA
TAATCACTAG CCAGATTTCG AAATTACTTT TTAGCCGAGG TTATGATAAC ATCTACTGTA
TCCTTTAGAA TTTTAACCTA TAAACTATG TCTACTGGTT TCTGCCTGTG TGCTTATGTT
AAAAAAAAAAAA

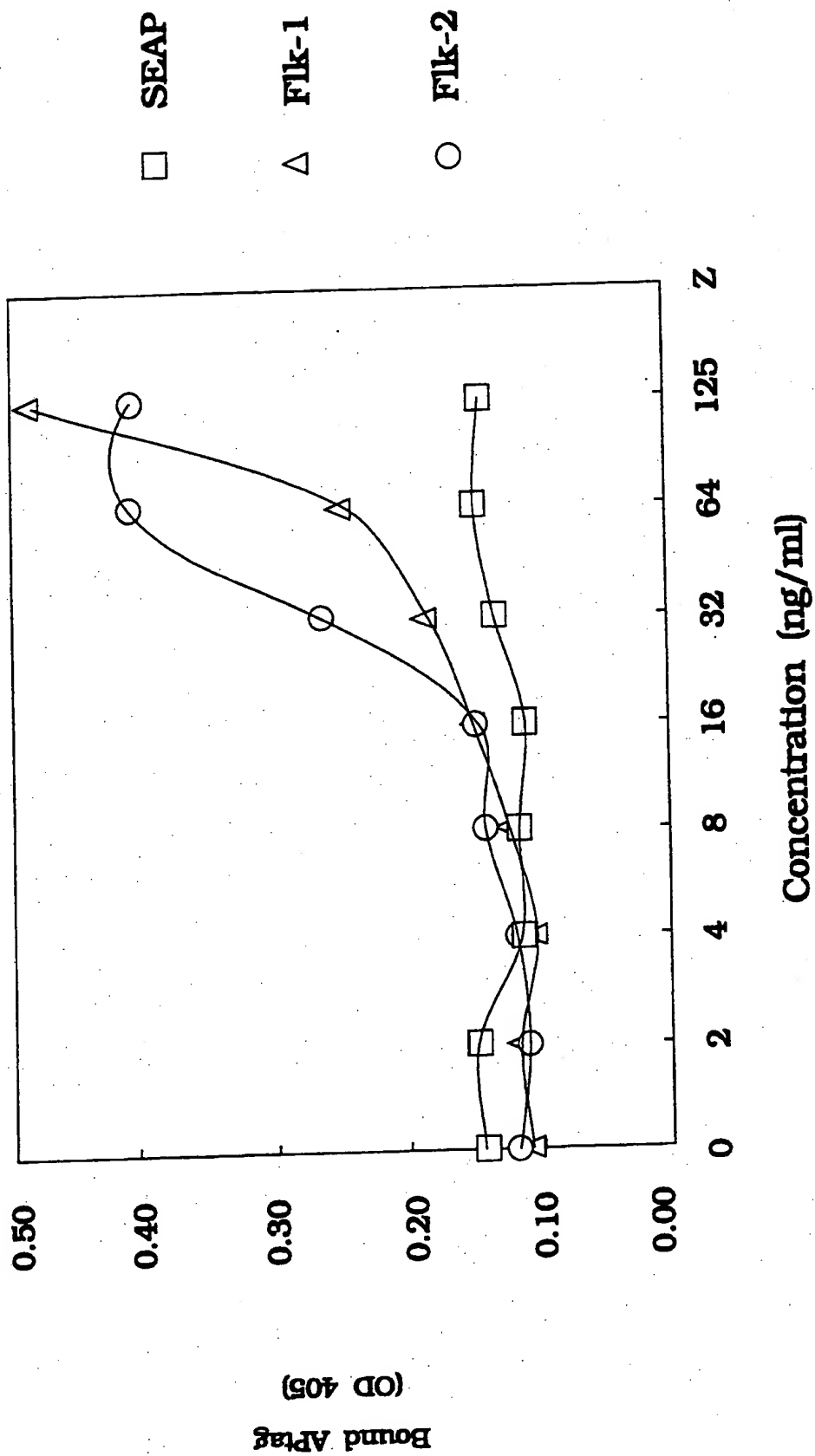
18/19

FIGURE 3



19/19

FIGURE 4



INTERNATIONAL SEARCH REPORT

International application No.
PCT/US92/09893

A. CLASSIFICATION OF SUBJECT MATTER

IPC(5) : C07H 15/12, 17/00; A61K 37/00; C07K 13/00, 15/00; C12N 5/00

US CL : 530/350, 387, 846; 536/27; 514/2; 435/240.2

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 530/350, 387, 846; 536/27; 514/2; 435/240.2

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

APS, DIALOG

search terms: protein tyrosine kinase, flt3, flk, hematopoiesis

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	Proceedings of the National Academy of Sciences, Vol. 86, issued March 1989, Wilks, "Two putative protein-tyrosine kinases identified by application of the polymerase chain reaction", pages 1603-1607, entire document.	1-64, 66-80
Y	Cell, Vol. 63, Issued 05 October, 1990, Flanagan et al., "The kit ligand: A cell surface molecule altered in steel mutant fibroblasts", pages 185-194, entire document.	41-64
X,P Y	Proceedings of the National Academy of Sciences, Vol. 88, Issued October 1991, Matthews et al., "A receptor tyrosine kinase cDNA isolated from a population of enriched primitive hematopoietic cells and exhibiting close genetic linkage to c-kit", pages 9026-9030, entire document.	14-17, 19,22,25,28, <u>31,34,76-80</u> 1 - 1 3 , 1 8 , 2 0 , 21,23,24,26,27,29,30,3 2,33,35-64,66-75
A	Science, Volume 241, Issued 01 July 1988, S.K. Hanks et al., "The protein kinase family: Conserved features and deduced phylogeny of the catalytic domains", pages 42-52, entire document.	1-64, 66-80

☒ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

* Special categories of cited documents:	*T	Inter document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
*A		document defining the general state of the art which is not considered to be part of particular relevance
*E		earlier document published on or after the international filing date
*L		document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
*O		document referring to an oral disclosure, use, exhibition or other means
*P		document published prior to the international filing date but later than the priority date claimed
	*X	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
	*Y	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
	*G	document member of the same patent family

Date of the actual completion of the international search

05 January 1993

Date of mailing of the international search report

02 FEB 1993

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Box PCT
Washington, D.C. 20231

Facsimile No. NOT APPLICABLE

Authorized officer

LORRAINE M. SPECTOR, PH.D.

Telephone No. (703) 308-1793

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US92/09893

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X.P Y	Cell, Volume 65, Issued 28 June 1991, W. Matthews et al., "A receptor tyrosine kinase specific to hematopoietic stem and progenitor cell-enriched populations", pages 1143-1152, entire document.	1-6,10-13, 18,20,21,23,24,26,27,2 9,30,32,33, <u>35,36-39,66-75</u> 7-9,41,43, 44,46-49, 53-55,56, 58,59,61,62,64,76-80
X.P Y	Oncogene, Volume 6, Issued 1991, O. Rosnet et al., "Murine Flt3, a gene encoding a novel tyrosine kinase receptor of the PDGFR/CSF1R family", pages 1641-1650, entire document.	1-6,10- 13,18,20,21,23,24,26,2 7,29,30,32,33, <u>35,36-39,66-75</u> 7-9,41,43, 44,46-49, 53-55,56,58, 59,61,62,64,76-80
X.P Y	Genomics, Volume 9, Issued 1991, O. Rosnet et al., "Isolation and chromosomal localization of a novel FMS-like tyrosine kinase gene", pages 380-385, entire document.	1-6,10-13, 18,20,21,23,24,26,27,2 9,30,32,33,35,36- <u>39,66-75</u> 7-9,41,43, 44,46-49, 53-55,56,58, 59,61,62,64, 76-80
A	Journal of Experimental Medicine, Volume 169, Issued May 1989, R.G. Andrews et al., "Precursors of colony-forming cells in humans can be distinguished from colony-forming cells by expression of the CD33 and CD34 antigens and light scatter properties", pages 1721-1731, entire document.	1-80
A	Cell, Volume 45, Issued 20 June 1986, I.R. Lemischka et al., "Developmental potential and dynamic behavior of hematopoietic stem cells", pages 917-927, entire document.	1-80
A	Cell, Volume 61, Issued 15 June 1990, C.T. Jordan et al., "Cellular and developmental properties of fetal hematopoietic stem cells", pages 953-963, entire document.	1-80
Y	Science, Volume 241, Issued 01 July 1988, G.J. Spangrude et al., "Purification and characterization of mouse hematopoietic stem cells", pages 58-62, entire document.	65
Y	R. HAY et al., "AMERICAN TYPE CULTURE COLLECTION CATALOGUE OF CELL LINES AND HYBRIDOMAS FIFTH EDITION, 1985", published 1985 by American Type Culture Collection (MD), see page 232, first entry.	65
Y	Proceedings of the National Academy of Sciences, Volume 84, Issued May 1987, B. Seed et al., "Molecular cloning of the CD2 antigen, the T-cell erythrocyte receptor, by a rapid immunoselection procedure", pages 3365-3369, entire document.	41-55

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US92/09893

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

2. ☐ Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:

3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:
Please See Extra Sheet.

1. ☒ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims. (Telephone Practice)
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:

4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☒ No protest accompanied the payment of additional search fees.

BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING

This ISA found multiple inventions as follows:

I. Claims 1-13, 18, 20, 21, 23, 24, 26, 27, 29, 30, 32, 33, 35-39, 41, 43, 44, 46-49, 53-56, 58, 59, 61, 62, 64-75, Drawn to flk-2, Class 530, subclasses 350, 387 and 846, Class 536, subclass 27, Class 514, subclass 2, and Class 435, subclass 240.2.

II. Claims 14-17, 19, 22, 25, 28, 31, 34, 40, 42, 45, 50-52, 57, 60, 63, 65, 76-80, drawn to flk-1, Class 530, subclasses 350, 387 and 846, Class 536, subclass 27, Class 514, subclass 2, and Class 435, subclass 240.2.

The claims of these two groups are drawn to distinct inventions which do not share a unifying technical feature. They are distinct proteins, with distinctly different amino acid sequences and patterns of expression. PCT Rules 13.1 and 13.2 do not provide for multiple products.